## A dormant microbial component in the development of pre-eclampsia<sup>1</sup>

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<sup>1</sup> Paper 8 in the series "The dormant blood microbiome in chronic, inflammatory diseases"

#### **Abstract**

Pre-eclampsia (PE) is a complex, multi-system disorder that remains a leading cause of morbidity and mortality in pregnancy. Four main classes of dysregulation accompany PE, and are widely considered to contribute to its severity. These are abnormal trophoblast invasion of the placenta, anti-angiogenic responses, oxidative stress, and inflammation. What is lacking, however, is an explanation of how these themselves are caused.

We here develop the unifying idea, and the considerable evidence for it, that the originating cause of PE (and of the four classes of dysregulation) is in fact microbial infection, that most such microbes are dormant and hence resist detection by conventional (replication-dependent) microbiology, and that by occasional resuscitation and growth it is they that are responsible for all the observable sequelae, including the continuing, chronic inflammation. In particular, bacterial products such as lipopolysaccharide (LPS), also known as endotoxin, are well known as highly inflammagenic and stimulate an innate (and possibly trained) immune response that exacerbates the inflammation further. The known need of microbes for free iron can explain the iron dysregulation that accompanies PE. We describe the main routes of infection (gut, oral, urinary tract infection) and the regularly observed presence of microbes in placental and other tissues in PE. Every known proteomic biomarker of "pre-eclampsia" that we assessed has in fact also been shown to be raised in response to infection. An infectious component to PE fulfils the Bradford Hill criteria for ascribing a disease to an environmental cause, and suggests a number of treatments, some of which have in fact been shown to be successful.

PE was classically referred to as endotoxaemia or toxaemia of pregnancy, and it is ironic that it seems that LPS and other microbial endotoxins really are involved. Overall, the recognition of an infectious component in the aetiology of PE mirrors that for ulcers and other diseases that were previously considered to lack one.

#### Insight, innovation, integration

Many descriptors of pre-eclampsia are widely accepted (e.g. abnormal trophoblast invasion, oxidative stress, inflammation and altered immune response, and anti-angiogenic responses). However, without knowing what causes them, they do not 'explain' the syndrome. The Biological Insight of this manuscript is that there is considerable evidence to the effect that each of these phenomena (hence PE) are caused by the resuscitation of dormant bacteria that shed (known and potent) inflammagens such as LPS, often as a consequence of iron availability. PE is thus seen as a milder form of sepsis. The Technological Innovations come from the use of molecular markers (of microbes and omics more generally, as well as novel markers of coagulopathies) to measure this. The Benefit of Integration comes from bringing together a huge number of disparate observations into a unifying theme.

#### Introduction

Pre-eclampsia. Pre-eclampsia (PE) is a multi-system disorder of pregnancy, characterised and indeed defined by the presence of hypertension after 20 weeks' gestation and before the onset of labour, or postpartum, with either proteinuria or any multisystem complication [1-8]. It is a common condition, affecting some 3-5% of nulliparous pregnant women [7; 9] and is characterised by high mortality levels [10-13]. There is no known cure other than delivery, and consequently PE also causes significant perinatal morbidity and mortality secondary to iatrogenic prematurity. There are a variety of known risk factors (Table 1), that may be of use in predicting a greater likelihood of developing PE, albeit there are so many, with only very modest correlations, that early-stage (especially first-trimester) prediction of late-stage PE remains very difficult [7; 14; 15].

It is striking that most of the 'risk factors' of Table 1 are in fact risk factors for multiple vascular or metabolic diseases, i.e. they merely pre-dispose the individual to a greater likelihood of manifesting the disease or syndrome (in this case PE). Indeed, some of them are diseases. This would be consistent with the well-known comorbidities e.g. between PE and later cardiovascular disease (e.g. [16-26]), between PE and intracerebral haemorrhage during pregnancy (OR 10.39 [27]), and between PE and stroke post-partum [28; 29]. The penultimate row of Table 1 lists a series of diseases that amount to comorbidities, although our interest was piqued by the observation that one third of patients with anti-phospholipid syndrome have PE, and infectious agents with known cross-reacting antigens are certainly one original (external) source of the triggers that cause the anti-phospholipid antibodies [30-33] (and see below). Similarly, in the case of urinary tract infection, the 'risk' factor is a genuine external trigger, a point (following the call [34] by Mignini and colleagues for systematic reviews) that we shall expand on considerably here.

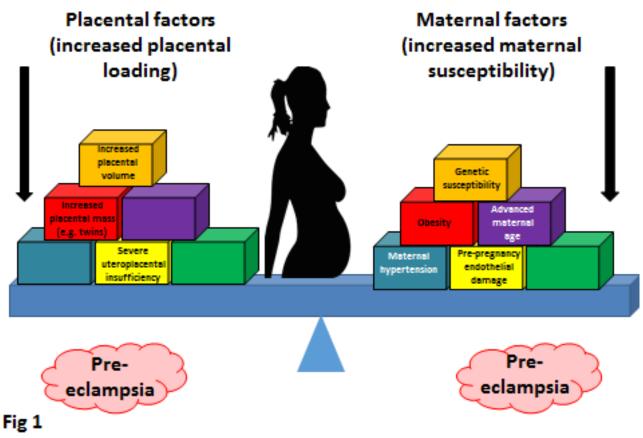
Table 1. Some known risk factors for pre-eclampsia (based in part on [2; 6; 35-37]). See also <a href="http://bestpractice.bmj.com/best-practice/monograph/326/diagnosis.html">http://bestpractice.bmj.com/best-practice/monograph/326/diagnosis.html</a>. Note that most of these are risk factors that might and do pre-dispose for other diseases (or are themselves diseases).

Risk factor	Comments	Selected references
Nulliparous women	Epidemiological observation; suggested biochemical explanations include soluble fmslike tyrosine kinase 1 (sFlt1): placental growth factor (PIGF) ratio	[2; 38]
Increased maternal age (especially >35 years)	Epidemiological observation, though may be related to existing age-related disease	[37; 39-42]
History of pre-eclampsia in previous pregnancy	Epidemiological observation, virtually akin to recurrence; among the strongest factors	[35; 43]
Multi-foetal gestation, i.e. twins, etc.	Extra demands on mother's circulation; larger placenta, danger of ischaemia? Relative risk ~3.5x in nulliparous	[44-46]
Obesity (esp BMI >35)	Can affect BP directly, also via intra-abdominal pressure; diabetogenic and inflammatory; possible role for asymmetric dimethylarginine	[47-50]
Booking diastolic BP > 80 mm Hg	An essential part of the later	[36]

	syndrome	
Booking proteinuria on at least 1	An essential part of the later	[36]
occasion, or ≥ 0.3g/24h	syndrome	
Family history of pre-eclampsia	2-5-fold increase in likelihood.	[51-56]
(mother or sister)	Genetic factors are said to	
	account for some 50% of the	
	variance, though few properly	
	controlled MZ/DZ twin studies	
	exist; when done the heritability	
	of PE can be lower to negligible	
Pre-existing medical conditions,	These are mainly seen as (other)	[5; 30; 31; 33; 57-64]
including chronic hypertension,	vascular diseases or	
diabetes mellitus,	comorbidities; however, anti-	
antiphospholipid syndrome,	phospholipid antibodies (Hughes'	
thrombophilia, autoimmune	syndrome) are of especial interest	
disease, renal disease, systemic	as they can have an infectious	
lupus erythematosus, infertility	origin; 1/3 of women with them	
	will develop PE, and they cause	
	recurrent pregnancy loss-	
Urinary tract infection	An infectious origin for PE is the	[65-67] and see below
	focus here, and not just from UTI	

In recent decades, intense investigation has led to the development of a two-stage aetiological model for pre-eclampsia, first proposed by Redman and colleagues [68], in which inadequate remodelling of the spiral arteries in early gestation results in poor placental development (stage one) and the resultant ischaemia/re-perfusion injury and oxidative stress eventually leads to maternal vascular endothelial cell dysfunction and the maternal manifestations of the disease (stage 2) [68-72]. However, many clinical inconsistencies challenge the simplicity of this model. For example, whilst the association between poor placentation and pre-eclampsia is well established, it is not specific. Poor placentation and foetal growth restriction (FGR) frequently present without maternal signs of pre-eclampsia. Moreover, FGR is not a consistent feature of pre-eclampsia. Whilst it is commonly seen in pre-eclampsia presenting at earlier gestations, in pre-eclampsia presenting at term, neonates are not growth restricted and may even be large for dates [73].

Thus, the two-stage model has been further refined by Roberts and others [72; 74; 75] to take into account the heterogeneous nature of pre-eclampsia and the varying contribution from mother and infant to the disorder. We now appreciate that normal pregnancy is characterised by a low-grade systemic inflammatory response and specific metabolic changes, and that virtually all of the features of normal pregnancy are simply exaggerated in pre-eclampsia [76-78]. There is also widespread acceptance that maternal constitutional and environmental factors (such as obesity) can interact to modulate the risk of pre-eclampsia. Thus, with profoundly reduced placental perfusion (or significant 'placental loading'), the generation of Stage 2 may require very little contribution from the mother to provide sufficient stress to elicit the maternal syndrome. In this setting, almost any woman will develop pre-eclampsia. Conversely, the woman with extensive predisposing constitutional sensitivity could develop pre-eclampsia with very little reduced perfusion, or minimal 'placental loading'. As with many complex disorders, multiple factors can affect disease development positively or negatively, with a convenient representation of the two main negative sources (foetal and maternal) being that of a see-saw [79], as in Fig 1.



Whilst this explains the inconsistencies of the two-stage model, the precise mechanisms 1) underlying the initial poor placentation and 2) linking placental stress and the maternal syndrome have still not been fully elucidated.

Much recent research in pre-eclampsia has focused on various angiogenic factors, including the proangiogenic factors vascular endothelial growth factor (VEGF) and placental growth factor (PIGF) and the two anti-angiogenic proteins, soluble endoglin (sEng) and soluble fms-like tyrosine kinase 1 (sFlt). Recent data suggest that alterations in circulating angiogenic factors play a pathogenic role in pre-eclampsia. These angiogenic factors tightly regulate angiogenesis and are also essential for maintenance of normal vessel health. Consequently, the synthesis and action of these factors and their receptors in the uterine bed and placenta are essential for normal placental development and pregnancy [80; 81]. In pre-eclampsia, increased levels of the anti-angiogenic sFlt-1 and sEng trap circulating VEGF, PIGF and transforming growth factor  $\beta$  (TGF $\beta$ ) respectively. A myriad of data supports the idea that circulating levels of these factors alone, or in combination, can be used to predict pre-eclampsia [82; 83] (and see below under PE biomarkers), but in line with the heterogeneous nature of pre-eclampsia, the data are somewhat inconsistent and their performance as biomarkers seems limited to disease with significant placental loading [7]. Therefore, angiogenic dysregulation would appear unlikely to be the sole link between the stressed placenta and endothelial dysfunction and the clinical manifestations of the disease.

Notwithstanding these many inconsistencies, the central role of the placenta as a source of 'toxin', in a condition regarded, and indeed often named, as 'toxaemia of pregnancy' [84-86] cannot be refuted. The uncertainty regarding the nature of the toxin(s) continues, and other placental sources of endothelial dysfunction include syncytiotrophoblast basement membrane fragments (STBM) [87] and endothelial progenitor cells (EPC) [88]; an increase of reactive oxygen species over scavenging by anti-oxidants [89; 90] has also been promoted.

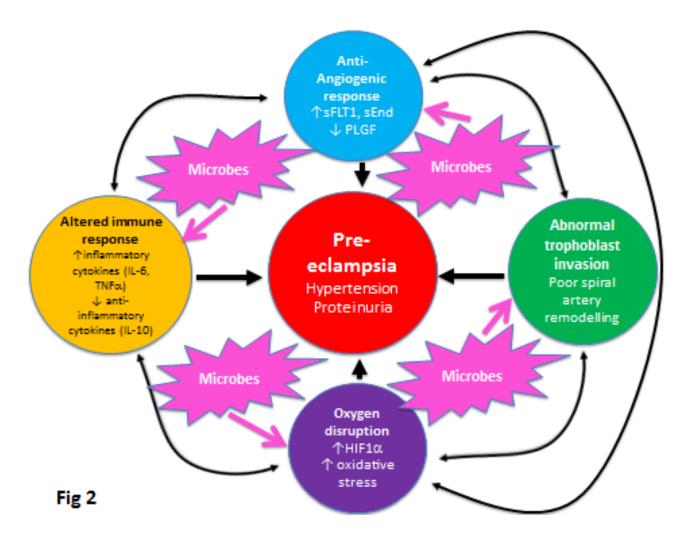
The Bradford Hill criteria for causation of a disease Y by an environmental factor X [91] are as follows:

(1) strength of association between X and Y; (2) consistency of association between X and Y; (3) specificity of association between X and Y; (4) experiments verify the relationship between X and Y; (5) modification of X alters the occurrence of Y; (6) biologically plausible cause and effect relationship.

In general terms [92], if we see that two things (A and B) co-vary in different circumstances, we might infer that A causes B, that B causes A, or that something else (C) causes both B and A, whether in series or parallel. To disentangle temporal relations requires a longitudinal study. The job of the systems biologist doing systems medicine is to uncover the chief actors and the means by which they interact [93], in this way fulfilling the Bradford Hill postulates, a topic to which we shall return at the end.

In infection microbiology, and long predating the Bradford Hill criteria, the essentially equivalent metrics are known (widely, but somewhat inaccurately [94]) as the Koch or Henle-Koch postulates (i.e. criteria). They involve assessing the correlation of a culturable organism with the presence of a disease, the cure of the disease (and its symptoms) upon removal of the organism, and the development of the disease with (re)inoculation of the organism. They are of great historical importance, but present us with three main difficulties here. The first is that we cannot apply the third of them to humans for obvious ethical reasons. The second (see also below) and related one is that we cannot usefully apply them in animal models because none of the existing models recapitulates human pre-eclampsia well. Finally, as widely recognised [94-101], they cannot be straightforwardly applied when dealing with dormant bacteria or bacteria that are otherwise refractory to culture.

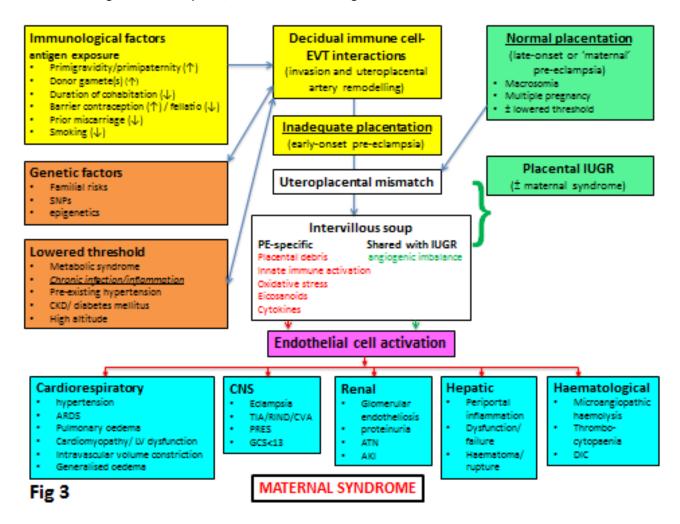
Our solution to this is twofold: (i) we can assess the first two using molecular methods if culturing does not work, and (ii) we exploit the philosophy of science principle known as 'coherence' [102-106]. This states that if a series of ostensibly unrelated findings are brought together into a self-consistent narrative, that narrative is thereby strengthened. Our systems approach purposely represents a 'coherence' in the sense given.



Overall, known biochemical associations with PE come into four main categories, viz. abnormal trophoblast invasion, oxidative stress, inflammation and altered immune response, and anti-angiogenic responses (Fig 2). Each of these can contribute directly to PE, and although they can interact with each other (black arrows), no external or causal source is apparent. Fig 2 has been redrawn from a very nice review by Pennington and colleagues [107], which indicates four main generally accepted 'causes' (or at least accompaniments) of PE as the four outer coloured circles. As illustrated with the black two-way arrows, many of these also interact with each other. What is missing, in a sense, is then what causes these causes, and that is the nub of our argument here. Since we now know (and describe below) that microbes can affect each of these four general mechanisms, we have added these routes to Fig 1 (using pink arrows) where dormant, resuscitating or growing microbes are known to contribute.

In a similar vein, Magee and colleagues [108] have nicely set down their related analysis of the causes and consequences of PE, with a central focus (redrawn in Fig 3) on endothelial cell activation. While bearing much similarity in terms of overall content to the analysis of Pennington and colleagues [107], and ours above, it again lacks a microbial or infection component as a <u>causative</u> element, but importantly does note

that infection and/or inflammation can serve to lower the threshold for PE in cases of inadequate placentation. In our view microbes can also enter following normal placentation if their dormant microbiome begins to wake up and/or to shed inflammagens.



Heritability. The question of the extent of heritability of PE (susceptibility) is of interest. Although this seems to vary widely in different studies (Table 1), a number of candidate gene studies [54; 109-112] imply that a susceptibility to PE is at least partly heritable, consistent with the variance in all the other 'risk factors' of Table 1 (and see [5]). As with all the other gene-association studies where phenotypic ('lifestyle') information is absent [113-115], it is not possible to ascribe the heritability to genetics alone, as opposed to an interaction of a genetic susceptibility (e.g. in the HLA system) with environmental factors [111], such as cytomegalovirus infection [116].

**Inflammation**. Pre-eclampsia is accompanied by oxidative stress [117] and inflammation, and thus shares a set of observable properties with many other (and hence related) inflammatory diseases, be they vascular (e.g. atherosclerosis), neurodegenerative (e.g. Alzheimer's, Parkinson's), or 'metabolic' (type 1 and 2 diabetes). It is thus at least plausible that they share some common aetiologies, as we argue here, and that knowledge of the aetiology of those diseases may give us useful clues for PE.

As well as raised levels of inflammatory cytokines, that constitute virtually a circular definition of inflammation, we and others have noted that <u>all</u> of these diseases are accompanied by dysregulation of iron metabolism [79; 118; 119], hypercoagulability and hypofibrinolysis [120; 121], blood microparticles

[119], and changes in the morphology of fibrin fibres (e.g. [122-127]) and of erythrocytes (e.g. [120; 125-130]).

In addition, we and others have recognised the extensive evidence for the role of a dormant blood and/or tissue microbiome in these [131-136] and related [137-140] diseases, coupled in part to the shedding of highly inflammagenic bacterial components such as Gram-negative lipopolysaccharides (LPS) and their Gram-positive cell wall equivalents such as lipoteichoic acids [141]. (We shall often use the term 'LPS' as a 'shorthand', to be illustrative of all of these kinds of highly inflammagenic molecules.)

The purpose of the present review, outlined as a 'mind map' in Fig 4, is thus to summarise the detailed and

## A 'mind map' of the review

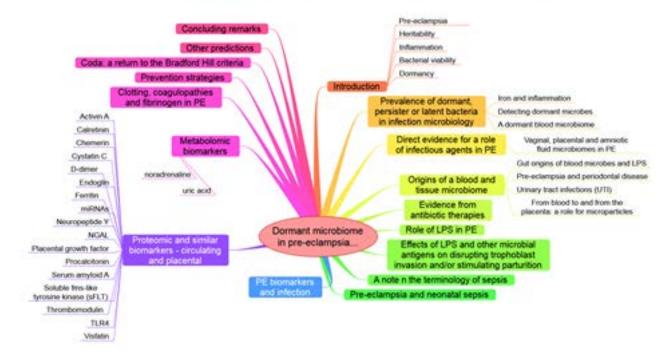
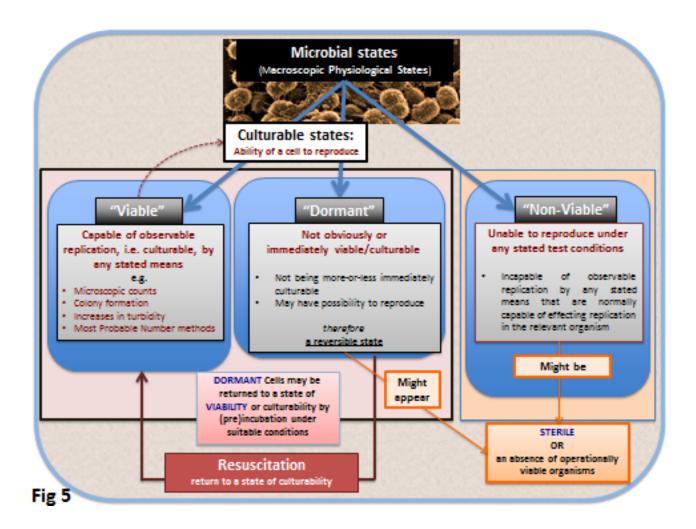


Fig 4

specific lines of evidence suggesting a very important role of a dormant microbial component in the aetiology of pre-eclampsia (and see also [131]). To do this, we must start by rehearsing what is meant by microbial dormancy.

Bacterial viability. In microbiology, we usually consider microbes as being in one of three 'physiological macrostates' (Fig 5). The definition of a 'viable' bacterium is normally based on its ability to replicate, i.e. 'viability' = culturability [142-144]. In this sense, classical microbiology has barely changed since the time of Robert Koch, with the presence of a 'viable' microorganism in a sample being assessed via its ability to form a visible colony on an agar plate containing suitable nutrients. However, it is well known, especially in environmental microbiology ('the great plate count anomaly' [145]), that only a small percentage of cells observable microscopically is typically culturable on agar plates. In principle this could be because they are or were 'irreversibly' non-culturable (operationally 'dead'), or because our culture media either kill them [146] or such media lack nutrients or signalling molecules necessary for their regrowth [147; 148] from an otherwise dormant state [149; 150]. Those statements are true even for microbes that appear in culture collections and (whose growth requirements) would be regarded as 'known'.



However, it is common enough in clinical microbiology that we detect the existence or presence of 'novel' microbial pathogens with obscure growth requirements before we learn to culture them; this is precisely what happened in the case of *Legionella pneumophila* [151-154], *Tropheryma whipplei* (Whipple's disease [155; 156]), and *Coxiella burnetii* (the causative agent of Q fever [157; 158]). Even *Helicobacter pylori* was finally brought into culture on agar plates only because an unusually long Easter holiday break meant that the plates were incubated for an extended period of five days (rather than the normal two) before being thrown out [159; 160]! Consequently, there is ample precedent for the presence of 'invisible' microbes to

go unremarked before they are discovered as the true cause of a supposedly non-infectious disease, even when they are perfectly viable (culturable) according to standard analyses.

**Dormancy** for a microbe is defined operationally as a state, commonly of low metabolic activity, in which the organism appears not to be viable in that it is unable to form a colony but where it is not dead in that it may revert to a state in which it can do so, via a process known as resuscitation [149; 150]. However, an important issue (and see above) is that <u>dormant</u> bacteria do not typically fulfil the Koch-Henle postulates [94; 96-98], and in order for them to do so it is necessary that they be grown or resuscitated. This is precisely what was famously done by Barry Marshall and Robin Warren when they showed that the <u>supposedly non-infectious</u> disease of gastric ulcers was in fact caused by a 'novel' organism called *Helicobacter pylori* [161; 162]. One of the present authors showed in laboratory cultures of actinobacteria that these too could enter a state of true dormancy [163; 164] (as is well known for *Mycobacterium tuberculosis*, e.g. [165-169]), and could be resuscitated by a secreted growth factor called Rpf [170-174]. This RPF family has a very highly conserved motif that is extremely immunogenic [175; 176], and it is presently under trials as a vaccine against *M. bovis*.

Prevalence of dormant, persistent or latent bacteria in infection microbiology. It is worth stressing here that the presence of dormant or latent bacteria in infection microbiology is well established; one third of humans carry dormant *Mycobacterium tuberculosis* (e.g. [165; 177-180]), most without reactivation, while probably 50-100% are infected with *H. pylori*, most without getting ulcers or worse [181; 182]. As with the risk factors in Table 1, the organisms are merely or equivalently 'risk factors' for those infectious diseases and are effectively seen as causative only when the disease is actually manifest.

In a similar vein, so-called persisters are <u>phenotypic</u> variants of infectious microbes that resist antibiotics and can effectively lie in hiding to resuscitate subsequently. This too is very well established (e.g. [132; 183-196]). In many cases they can hide intracellularly [197], where antibiotics often penetrate poorly [198] because the necessary transporters [199-202] are absent. This effectively provides for reservoirs of reinfection, e.g. for *Staphylococcus aureus* [203], *Bartonella* spp [204] and – most pertinently here – for the *Escherichia coli* involved in urinary tract (re)infection [205-208]. The same intracellular persistence is true for parasites such as *Toxoplasma gondii* [209].

Thus, the main point of the extensive prevalence of microbial dormancy and persistence is that microbes can appear to be absent when they are in fact present at high concentrations. This is true not only in cases where infection is recognised as the cause of disease but, as we here argue, such microbes may be an important part of diseases presently thought to lack an infectious component.

**Iron and inflammation**. It is well known that (with the possible exception of *Borrelia* [210; 211]) a lack of free iron normally limits microbial growth *in vivo* (e.g. [212-236]), and we have reviewed previously [79; 118; 119] the very clear iron dysregulation accompanying pre-eclampsia (e.g. [84; 237-249]).

This has led to the recognition [121; 132; 134] that the source of the <u>continuing</u> inflammation might be iron-based resuscitation of dormant microbes that could release well-known and highly potent inflammagens such as lipopolysaccharide (LPS). Indeed, we have shown that absolutely tiny (highly substoichiometric) amounts of LPS can have a massive effect on the blood clotting process [250], potentially inducing  $\beta$ -amyloid formation directly [251; 252] (something, interestingly, that can be mimicked in liquid crystals [253; 254]). The overall series of interactions envisaged (see also [132]) is shown in Fig 6.

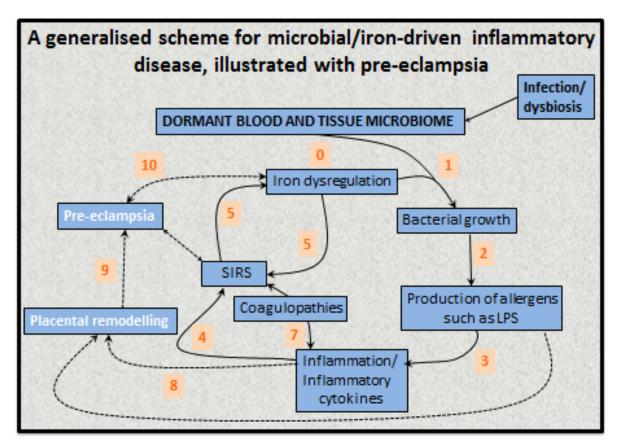


Fig 6

Detecting dormant microbes. By definition, dormant bacteria escape detection by classical methods of assessing viability that involve replication on agar plates. Other growth-associated methods include measurements involving changes in turbidity [255], including an important but now rather uncommon technique referred to as the 'most probable number' (MPN). The MPN involves diluting samples serially and assessing by turbidity changes the presence of growth/no growth. Look-up tables based on Poisson statistics enable estimation of the number of cells or propagules that were present. A particular virtue is that they allow dormant and 'initially viable' cells to be discriminated via 'dilution to extinction' [164], thereby avoiding many artefacts [150]. As mentioned above, preincubation in a weak nutrient broth [164; 256] was instrumental in allowing the discovery [170] of an autocrine 'wake-up' molecule necessary for the growth of many actinobacteria.

Other more classical means of detecting microbes, but not whether they were <u>culturable</u>, involved microscopy [183; 257-260] or flow cytometry [261] with or without various stains that reflected the

presence or otherwise of an intact cell wall/membrane [163; 262-269]. These stains are sometimes referred to as 'viability' stains, but this is erroneous as they do not measure 'culturability'. Readers may also come upon the term 'viable-but-not-culturable'; however, since viable = culturable, this is an oxymoron that we suggest is best avoided [150]. Other methods involved measurement of microbial products, e.g. CO<sub>2</sub> [270; 271], or changes in the conductivity or impedance of the growth medium [255; 272-274].

Most importantly, however, dormant (as well as culturable) cells may be detected by molecular means, nowadays most commonly through PCR and/or sequencing of the DNA encoding their small subunit ribosomal RNA (colloquially '16S') [275-289] or other suitable genes. It is clear that such methods will have a major role to play in detecting, identifying and quantifying the kinds of microbes that we argue lie at the heart of PE aetiology.

A dormant blood microbiome. Of course actual bacteraemia, the presence of replicable bacteria in blood, is highly life-threatening [290], but - as emphasised - viability assays do not detect dormant bacteria. When molecular detection methods are applied to human blood, it turns out that blood does indeed harbour a great many dormant bacteria (e.g. [291-301]); they may also be detected ultramicroscopically (e.g. [132-134; 183; 259; 292; 302]) or by flow cytometry [303], and dormant blood and tissue microbes probably underpin a great many chronic, inflammatory diseases normally considered to lack a microbial component [132-134; 137-140; 183; 259; 260; 294; 304-313]. Multiple arguments serve to exclude 'contaminants' as the source of the bacterial DNA [134]: 1. There are significant differences between the blood microbiomes of individuals harbouring disease states and nominally healthy controls, despite the fact that samples are treated identically; 2. The morphological type of organism (e.g. coccus vs bacillus) seems to be characteristic of particular diseases; 3. In many cases relevant organisms lurk intracellularly, which is hard to explain by contamination; 4. There are just too many diseases where bacteria have been found to play a role in the pathogenesis, that all of them may be caused by contamination; 5. The actual numbers of cells involved seem far too great to be explicable by contamination; given that blood contains ~5.109 erythrocytes.mL-1, if there was just one bacterial cell per 50,000 erythrocytes this will equate to 10<sup>5</sup> bacteria.mL<sup>-1</sup>. These are big numbers, and if the cells were culturable, that number of cells would be the same as that ordinarily defining bacteriuria.

A recent study by Damgaard and colleagues [298] is of particular interest here. Recognising the strong mismatch between the likelihood of an infection post-transfusion (very high [298]) and the likelihood of detecting culturable microbes in blood bank units (negligible, ca 0.1%) [298; 314], Damgaard *et al* reasoned that our methods of detecting and culturing these microbes might be the problem. Certainly, taking cells from a cooled blood bag and placing them onto an agar plate at room temperature that is directly exposed to atmospheric levels of gaseous  $O_2$  is a huge stress leading to the production of 'reactive oxygen species' [118; 315], that might plausibly kill any dormant, injured, or even viable microbes. Thus they incubated samples from blood on a rich medium (trypticase soy agar) for a full week, both aerobically and anaerobically. Subsequent PCR and sequencing allowed them to identify specific microbes in some 35-53% of the samples. Thus, very careful methods need to be deployed to help resuscitate bacteria from physiological states that normally resist culture, even when those bacteria are well-established species. This is very much beginning to happen in environmental microbiology (e.g. [147; 316-318]), and such organisms are rightly seen as important sources of novel bioactives [319; 320].

As reviewed previously [132-136], the chief sources of these blood microbes are the gut microbiome, the oral microbiome (periodontitis [321]), and via urinary tract infections. Consequently, if we are to argue that there is indeed a microbial component to pre-eclampsia, we should expect to see some literature evidence for it [66; 67; 131; 322-324]. In what follows we shall rehearse the fact that it is voluminous.

## Direct evidence for a role of infectious agents in PE

Although we recognise that many of the more molecular methods cannot distinguish culturable from dormant microbes, quite a number of studies have explicitly identified infection as a cause of PE (Table 2). The commonest microbe seems to be *H. pylori*; while it is most famously associated with gastric ulcers [161; 162; 325], there are many other extragastric manifestations (e.g. [326-334]). The Odds Ratio of no less than <u>26</u> in PE vs controls when the strains can produce CagA antigens is especially striking, not least because it provides a mechanistic link to poor trophoblast invasion via a mechanism involving host antibodies to CagA cross-reacting with trophoblasts [335; 336], and circulating [337] in microparticles [338] or endosomes [339; 340].

Table 2. Many studies have identified a much greater prevalence of infectious agents in the blood or urine of those exhibiting PE than in matched controls

Microbes	Comments	Reference
Chlamydia pneumoniae	IgG seroprevalence and gDNA associated with PE (p<0.0001)	[341]
	IgG (but not IgA or IgM) associated with PE, OR = 3.1.	[342]
	Significantly greater numbers with PE, and reversion under antichlamydial treatment	[343]
Chlamydia trachomatis	Increased risk of PE, OR = 7.2 or 1.6 based on serology	[344; 345]
Cytomegalovirus	RR for PE 1.5 if infected with CMV	[346] (see also [347])
Helicobacter pylori	Seropositivity or DNA. OR=2.7, or <b>26</b> if CagA seropositivity	[335] and editorial [348]
	IgG seropositivity 54%PE vs 21% controls	[349]
	Anti-CagA antibodies cross-react with trophoblasts and could inhibit placentation	[350]
	2.8x greater seropositivity in PE group	[351]
	OR=2.86 for seropositivity in PE, correlated with high malondialdehyde levels	[352]
	Wide-ranging review of many studies showing PE more prevalent after <i>Hp</i> infection	[353]
	Seropositivity PE:control = 84%:32% (p<0.001)	[354]
	OR for seropositivity 1.83 (p<0.001)	[355]
-	Seropositivity PE:control 86%:43%	[356]

	(p<0.001)	
Human papillomavirus	High-risk human papillomavirus (HR-HPV) presence implies an OR of 2.18 for PE.	[357]
Meta-analyses	Incidence of PE 19% with asymptomatic bacteriuria, vs 3% (primigravid) or 6% (multigravid) controls (p<0.005)	[358]
	UTI more than twice as likely in severe preeclamptics than in controls	[359]
	OR of 1.6 for PE if UTI present	[360]
	Increased risk of PE OR 1.57 for UTI, 1.76 for periodontal disease	[66]
	Early application of antibiotics in infection reduced PE by 52%	[322]
	Any overt infection led to an RR of 2 for PE	[67]
	UTI has OR of 3.2 for PE; OR = 4.3 if in third trimester	[361]
	UTI has OR of 1.3 for mild/moderate and 1.8 for severe PE	[362]
	Increased risk of PE with UTI (OR 1.22) or antibiotic prescription (OR 1.28)	[363]
	OR of 6.8 for symptomatic bacteriuria in PE vs controls	[364]
	OR 1.3-1.8 of mild or severe PE if exposed to UTI	[365]
	OR 1.4 for PE following UTI	[366]
	OR 1.3 for PE after UTI	[367]
	Meta-analyses showing associations between PD and PE	[368-370]
Plasmodium falciparum (malaria)	Indications that infection with malaria is associated with PE	[371]
	1.5 RR for PE if malarial	[372]
	Seasonality: 5.4-fold increase in eclampsia during malaria season	[373]
	Pre-eclampsia was significantly associated with malaria infection during pregnancy (p<0-03) and 69-7% of cases of pre-eclampsia with infected placenta might be attributable to malaria infection	[374]

In contrast to the situation in PE, albeit severe PE is associated with iatrogenic pre-term births, there is a widespread recognition (e.g. [375-402]) that infection is a common precursor to pre-term birth (PTB) in the absence of PE. The failure of antibiotics to help can be ascribed to their difficulty of penetrating to the

trophoblasts and placental regions. Unfortunately no proteomic biomarkers have yet been observed as predictive of PTB [403; 404]. In a similar vein, and if we are talking about a time of parturition that is very much more 'preterm', we are in the realm of miscarriages and spontaneous abortions and stillbirths, where infection again remains a major cause [405-408]. Here we note that <u>early or pre-emptive</u> antibiotic therapy <u>has</u> also proved of considerable value in improving outcomes after multiple spontaneous abortions [409].

#### Vaginal, placental and amniotic fluid microbiomes in PE

It might be natural to assume that the placenta is a sterile organ, like blood is supposed to be. However, various studies have shown the presence of microbes in tissues including the placenta [386; 395; 410-422], vagina [383; 423-429], uterus [387; 430; 431], amniotic fluid [422; 432-437], and follicular fluid [438; 439], and how these may vary significantly in PE (we do not discuss other pregnancy disorders such as small for gestational age (SGA) and intrauterine growth restriction (IUGR)). We list some of these in Table 3.

Table 3. Evidence for microbes in placental tissues, including those with PE.

Organisms	Comments	Reference
Multiple, including Actinobacillus actinomycetemcomitans, Fusobacterium nucleatum.	Many more in PE placentas relative to controls ( $p \le 0.0055$ )	[440]
Multiple	Half of second-trimester pregnancies have culturable or PCR-detectable bacteria/ mycoplasmas	[410]
Multiple	38% of placental samples were positive for selected bacteria and viruses	[441]
Bifidobacterium spp. and Lactobacillus rhamnosus	Bifidobacteria and L. rhamnosus (from gut) detected in 31/34 and 33/34 placental samples	[411]
Multiple	Detectable in 27% of all placentas and 54% of spontaneous preterm delivery	[412]
Multiple	16S/NGS, major review	[395]
Multiple	From 16S and NGS analysis of placental tissue of 7 PE patients (12.5%) (controls all negative)	[442]
E. coli and L. monocytogenes	When added <i>ex vivo</i> can migrate to extravillous trophoblasts	[415]
Multiple	Review, with some focus on preterm birth	[414]
Multiple	Overview, some focus on preterm birth	[416]
Multiple	Good recent overview, with possible implication of a physiological role	[418]
Multiple	320 placentas; changed microbiome as a function of excess gestational weight gain	[419]
Multiple	One third of placentas from	[420]

	preterm births were culture- positive	
Multiple	Major differences in placental microbiome in preterm birth	[421]
Plasmodium falciparum (malaria)	Increased likelihood of PE	[372]
Plasmodium falciparum (malaria)	Reviews of placental malaria	[443; 444]
Porphyromonas gingivalis	OR of PE = <u>6.3</u> if detected in umbilical cord	[445]
Porphyromonas gingivalis	OR <u>7.59</u> in placental tissues with hypertensive disorders	[446]
Treponema denticola	OR <u>9.39</u> in placental tissues with hypertensive disorders	[446]
Meta-analysis	Widespread occurrence of microbes in female genital tract during pregnancy	[400]

## Origins of a blood and tissue microbiome

As assessed previously [132-134] over a large literature, the chief source of blood microbes is the gut [418], with another major entry point being via the oral microbiome (especially in periodontitis, see below). For rheumatoid arthritis [135; 447-449] and diseases of pregnancy, urinary tract infection (see below and Table TT) also provides a major source.

#### Gut origins of blood microbes and LPS

We have recently rehearsed these issues elsewhere [132-134], so a brief summary will suffice. Clearly the gut holds trillions of microbes, with many attendant varieties of LPS [450], so even low levels of translocation (e.g. [451-453]), typically via Peyer's patches and M cells, provide a major source of the blood microbiome. This may be exacerbated by intra-abdominal hypertension that can indeed stimulate the translocation of LPS [454]. For reasons of space and scope, we do not discuss the origins and translocation of microbes in breast milk [455], nor the important question of the establishment of a well-functioning microbiome in the foetus and neonate [456], and the physiological role of the mother therein.

#### Pre-eclampsia and periodontal disease

One potential origin of microbes that might be involved in, or represent a major cause of, pre-eclampsia is the oral cavity, and in particular when there is oral disease (such as periodontitis and gum bleeding) that can allow microbes to enter the bloodstream. If this is a regular occurrence one would predict that PE would be much more prevalent in patients with pre-existing periodontitis (but cf. [457] for those in pregnancy) than in matched controls; this is indeed the case (Table 4).

Table 4. Periodontal disease (PD) and pre-eclampsia

Organisms	Comments	References
Meta-analyses	OR of PE increased 3.69-fold if PD	[458]
	before 32 weeks	
	OR of 3 for the development of	[459]
	PEif ureaplasmas present at first	
	antenatal visit	

OR 2.1 for preceding PE Extensive overview of role of oral health and periodontal disease in PE OR 3.71 for PE if history of periodontal treatment Excellent overview of likely relationship between PD and PE OR = 8.6 or 2.03 for PE if PD was present vs controls Strong association between PD and PE (P<0.01) Overview with many references OR for association between PD and PE (P<0.01) Overview with many references OR for association between PD and PE (P<0.03) Overview with many references OR for association between PD and PE -3.73. No correlation with TNF-c or II OR 2.46 PE:controls Excellent overviews, focusing on means of transport of microbes from mouth to reproductive tissue Relationship between C-reactive protein, PE and severity of PD Adjusted PE RR 5.8 for Women with periodontal disease and CRP >75th percentile compared to women without periodontal disease on the periodontal attachment level scores in the preciamptic group compared with controls (2.98 to 2.11 and the probing depth and clinical attachment level scores in the preciamptic group compared with controls (2.98 to 2.11 and the probing depth and clinical attachment level scores in the preciamptic group compared with controls (2.98 to 2.11 and the probing depth and clinical attachment level scores in the preciamptic group compared with controls (2.98 to 2.11 and the probing depth and clinical attachment level scores		OR 5.56 for PD preceding PE	[460]
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OR 3.71 for PE if history of periodontal treatment		·	
Periodontal treatment   Excellent overview of likely relationship between PD and PE   OR = 8.6 or 2.03 for PE if PD was present vs controls   Strong association between PD and PE   (A65)			
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·		preeclamptic group compared	
		with controls (2.98 vs 2.11 and	
3.33 vs 2.30, respectively).		3.33 vs 2.30, respectively).	

#### **Urinary tract infections (UTI)**

A particular feature of UTIs is the frequency of reinfection [483-490]. This is because the organisms can effectively 'hide' in bladder epithelial cells as so-called 'quiescent intracellular reservoirs' [206; 487; 489; 491-495] of (presumably) dormant cells that can resuscitate. This is why reinfection is often from the same strains that caused the original infection [496-500]. Other complications can include renal scarring [501]. Bacteriuria (often asymptomatic) is a frequent occurrence in pregnancy (e.g. [365; 367; 459; 502-508]), and the frequency of UTI as a source of microbes causing PE is clear from Table 2.

#### From blood to and from the placenta; a role for microparticles

We and others have noted the fact that many chronic, inflammatory disease are accompanied by the shedding of various antigens and other factors; typically they pass through the bloodstream as microparticles [119; 133; 509-514], sometimes known as endosomes [337; 339; 340; 510; 515] (and see later under miRNAs). Similarly, LPS is normally bound to proteins such as the LPS-binding protein and apoE (see [133]).

### **Evidence from antibiotic therapies**

Antibiotic drug prescriptions may be seen as a proxy for maternal infection, so if dormant (and resuscitating and growing) bacteria are a major part of PE aetiology one might imagine an association between antibiotic prescriptions and PE. According to an opposite argument, antibiotics and antibiotic prescriptions given for nominally unrelated infections (UTI, chest, etc, and in particular diseases requiring long-term anti-infective medication that might even last throughout a pregnancy) might have the beneficial side-effect of controlling the proliferation of dormant cells as they seek to resuscitate. There is indeed some good evidence for both of these, implying that it is necessary to look quite closely at the nature, timing and duration of the infections and of the anti-infective therapy relative to pregnancy. A summary is given in Table 5. A confounding factor can be that some (e.g. the antiretroviral) therapies are themselves quite toxic [516; 517]; while the OR for avoiding PE was 15.3 in one study of untreated HIV-infected individuals vs controls, implying (as is known) a strong involvement of the immune system in PE, the 'advantage' virtually disappeared upon triple-antiretroviral therapy [518]. Overall, it is hard to draw conclusions from antiretrovirals [519; 520]. However, we have included one HIV study in the Table. Despite a detailed survey, we found no reliable studies with diseases such as Lyme disease or tuberculosis, where treatment regimes are lengthy, that allowed a fair conclusion as to whether antibiotic treatment was protective against PE. However, we do highlight the absolutely stand-out study of Todros and colleagues [521], who noted that extended spiramycin treatment (of patients with Toxoplasma gondii) gave a greater than tenfold protection against PE, when the parasite alone had no effect [522]. This makes such an endeavour (assessing the utility of early or pre-emptive antibiotics in PE) potentially highly worthwhile.

Table 5. Examples of decreased PE following antibiotic therapies given for various reasons

Target organisms	Comments	Reference
HIV	OR of 0.65 for patients treated	[523]

	with mono- or triple anti- retroviral therapy	
Toxoplasma gondii	<u>Massive</u> (OR=0.092) protection against PE in patients treated with spiramycin	[521]
Various organisms	52% decrease in PE following 10- day antibiotic therapy	[322]

#### Role of LPS in PE

It is exceptionally well known that LPS (*sensu lato*) is highly inflammagenic, and since one of us recently reviewed that literature *in extenso* [133] this is not directly rehearsed here. However, since we are arguing that it has a major role in PE naturally or *in vivo*, we do need to ask whether the literature is consistent with this more focussed question. The answer is, of course, a resounding 'yes'. Notwithstanding that only primates, and really only humans, are afflicted by 'genuine' PE, so the genuine utility of rodent models is questionable [524], even if some can recapitulate elements of the disease [525; 526]. Hence, it is somewhat ironic that there are a number of animal models in which LPS (also known as 'endotoxin') is used experimentally to induce a condition resembling PE (e.g. [527-532], and see [533]). We merely argue that it is not a coincidence that exogenous administration of LPS has these effects, because we consider that it is in fact normally one of the main mediators of PE.

The standard sequelae of LPS activation, e.g. TLR signalling and cytokine production, also occur in PE [534; 535], bolstering the argument that this is precisely what is going on. In a similar vein, double stranded RNA-mediated activation of TLR3 and TLR7/8 can play a key role in the development of PE [536-538]. What is new here is our recognition that LPS and other inflammagens (e.g. [539-541]) may continue to be produced and shed by dormant and resuscitating bacteria that are generally invisible to classical microbiology.

# Effects of LPS and other microbial antigens on disrupting trophoblast invasion and/or stimulating parturition

As with other cases of cross-reactivity such as that of various antigens in *Proteus* spp that cause disease in rheumatoid arthritis [447-449], the assumption is that various microbial antigens can lead to the production of (auto-)antibodies that attack the host, in the present case of interest by stopping the placentation by trophoblasts. This is commonly referred to as 'molecular mimicry' (e.g. [542-545]), and may extend between molecular classes e.g. peptide/carbohydrate [546; 547]. Table 6 shows some molecular examples where this has been demonstrated.

Table 6. Molecular examples of bacterial antigens that can elicit antibodies that stop successful trophoblast implantation or stimulate parturition.

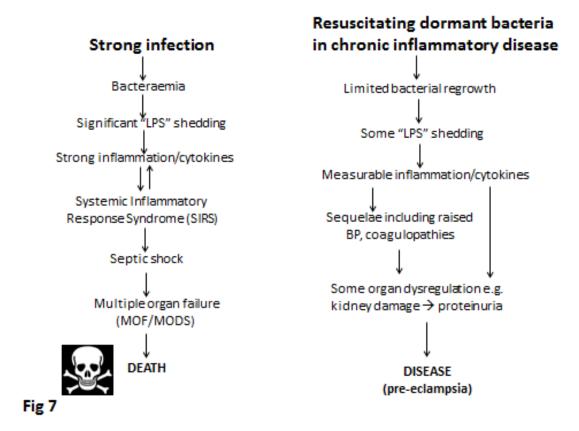
Organism	Antigen and comments	Reference
Gram-negatives	LPS can stimulate parturition, via corticotropin-releasing hormone	[548]
Gram-negatives	LPS can stimulate parturition, via MAPKinase	[535]

Helicobacter pylori	Anti-CagA antibodies cross-react	[336; 350]
	with trophoblasts and could	
	inhibit placentation	
Porphyromonas gingivalis	LPS inhibits trophoblast invasion	[480]
Various	Antiphospholipid antibodies (that	[549-552]
	can be induced by microbes, see	
	above)	

In many cases, the actual (and possibly microbial) antigens are unknown, and clearly the microbial elicitation of antibodies to anything that might contribute to PE points to multiple potential origins. To this end, we note that PE has also been associated with antibodies to angiotensin receptors [553-566], to smooth muscle [567; 568] (such blocking may be anti-inflammatory [569-571]), to adrenoceptors [572], to the M2 muscarinic receptor [573], and to Th17 [574](and see [575]). It is not unreasonable that epitope scanning of the antibody targets coupled to comparative sequence analysis of potential microbes might light up those responsible. In the case of Angiotensin II Type 1 Receptor Antibodies the epitope is considered [576] to be AFHYESQ, an epitope that also appears on parvovirus B19 capsid proteins; in the event, parvoviruses seem not to be the culprits here [577]. However, the role of these antibodies in activating the angiotensin receptor is also considered to underpin the lowering of the renin-angiotensin system that is commonly seen in PE [578-581], but which is typically raised during normal pregnancy.

Th-17 is of especial interest here, since these are the helper T (Th)-cell subset that produce IL-17. IL-17 is probably best known for its role in inflammation and automimmunity [575; 582-586]. However, it also has an important role in induction of the protective immune response against extracellular bacteria or fungal pathogens at mucosal surfaces [584; 587-599]. Th17 cells seem to participate in successful pregnancy processes and can be lower in PE [600-602], though more studies show them as higher [575; 603-611] or unchanged [612; 613]. One interpretation, consistent with the present thesis, is that the antimicrobial effects of placental IL-17 relative to T<sub>reg</sub> cells are compromised during PE [575; 609; 614].

A note on the terminology of sepsis. As one may suppose from the name, sepsis (and the use of words like 'antiseptic') was originally taken to indicate the presence of culturable organisms in (or in a sample taken from) a host, e.g. as in bacteraemia. Recognising that it is the <u>products</u> of bacteria, especially cell wall components, that cause the cytokine storms that eventually lead to death from all kinds of infection [615-618], 'sepsis' nowadays has more come to indicate the latter, as a stage (in the case of established infection) on a road that leads to septic shock and (eventually) to death (with a shockingly high mortality, and many failures of initially promising treatments, e.g. [619; 620], and despite the clear utility of iron chelation [79; 118; 621; 622]). In most cases significant numbers of culturable microbes are either unmeasured or absent, and like most authors we shall use 'sepsis' to imply the results of an infection whether the organisms are detected or otherwise. Overall, it is possible to see the stages of PE as a milder form of the sepsis cascade on the left-hand side of Figure 7. Fig 7 compares the classical route of sepsis-induced death with the milder versions that we see in PE; they are at least consistent with the idea that PE is strongly related to the more classical sepsis in degree rather than in kind.



**Pre-eclampsia and neonatal sepsis**. If PE is really based on infectious agents, it is reasonable that one might expect to see a greater incidence of neonatal sepsis (i.e. infection) following PE. While there are clearly other possible explanations (e.g. simply a weakened immune system, sometimes expressed as neutropaenia, after PE), there is certainly evidence that this is consistent with this suggestion [623-627].

**PE biomarkers and infection**. Because of the lengthy development of PE during pregnancy, there has long been a search for biomarkers (somewhat equivalent to the 'risk factors' discussed earlier) that might have predictive power, and some of these, at both metabolome [14; 628-635] and proteome [636-638] level, are starting to come forward. The typical experimental design is a case-control, in which markers that are raised or lowered significantly relative to the age-matched controls are considered to be candidate markers of PE. However, just as noted with leukocyte markers [76] and PCOS [639], that does not mean that they might not also be markers for other things too, such as infection [640]!

Thus, one prediction is that if dormant and resuscitating bacteria are responsible for PE then **at least some of these biomarkers should also be (known to be) associated with infection**. However, one obvious point is that the markers may appear only <u>after</u> infection, and this may itself be after the first trimester; clearly then these would not then be seen as 'first-trimester' biomarkers! There are many well-known inflammatory biomarkers that are part of the innate (and possibly trained [641]) immune response, such as the inflammatory cytokines CRP (cf. [642; 643]), IL-6 [644], IL-1 $\beta$  [645], TNF $\alpha$  [646], and macrophage migration inhibitory factor (MIF) [647], that are also all biomarkers of infection [648-652]. Certainly the fact that these increase in PE is consistent with a role for an infectious component. However, we shall mainly look at <u>other</u> biomarkers that are known to increase with PE, and see if they are also known to be biomarkers for (or at least changed in the presence of) infection (and see Th17/IL-17 above), and we next examine this. We shall see that pretty well every biomarker that is changed significantly in PE is also

changed following infection, a series of findings that we consider adds very strong weight to our arguments.

#### Proteomic and similar biomarkers - circulating and placental

What is really needed is a full systems biology strategy (see e.g. [93; 653-655]) that brings together the actors that interact then parametrises the nature of those interactions in a suitable encoding (e.g. SBML [656]) that permits their modelling, at least as an ODE model using software such as CellDesigner [657], COPASI [658] or Cytoscape [659]. Thus, to take a small example, "agonistic autoantibodies against the angiotensin II type 1 receptor autoantibodies (AT1-AA) are described. They induce NADPH oxidase and the MAPK/ERK pathway leading to NF-κB and tissue factor activation. AT1-AA are detectable in animal models of PE and are responsible for elevation of soluble fms-related tyrosine kinase-1 (sFlt1) and soluble endoglin (sEng), oxidative stress, and endothelin-1, all of which are enhanced in pre-eclamptic women. AT1-AA can be detected in pregnancies with abnormal uterine perfusion" [565]. Many such players have been invoked, and we next list some.

**Activin A**. Activin A is a member of the transforming growth factor (TGF)- $\beta$  superfamily. Its levels are raised significantly in PE [112; 660]. However, activin A is also well-established as a biomarker of infection [661-664].

*Calretinin*. In a proteomic study of pre-clamptic vs normal placentas [665], calretinin was one of the most differentially upregulated proteins ( $P = 1.6.10^{-13}$  for preterm PE vs controls,  $P = 8.9.10^{-7}$  for term PE vs controls), and in a manner that correlated with the severity of disease. While calretinin (normally more expressed in neural tissue and mesotheliomas [666]) is not normally seen as a marker of infection, it is in fact raised significantly when *Chlamydia pneumoniae* infects human mesothelial cells [667].

Chemerin is a relatively recently discovered adipokine, whose level can increase dramatically in the first trimester of pre-eclamptic pregnancies [668], and beyond [669]. Its levels are related to the severity of the pre-eclampsia [670-672]. Specifically, an ROC curve [673] analysis showed that a serum chemerin level >183.5 ng.mL<sup>-1</sup> predicted pre-eclampsia with 87.8% sensitivity and 75.7% specificity (AUC, 0.845; 95% CI, 0.811–0.875) [668]. Papers showing that chemerin is also increased by infection (hence inflammation) include [674; 675]; it even has antibacterial properties [676; 677], and was protective in a skin model of infection [678; 679]. In a study of patients with sepsis [680], circulating chemerin was increased 1.69-fold compared with controls (p = 0.012), and was also protective as judged by survival. These seem like particularly potent argument for a role of chemerin as a marker of infection rather than of pre-eclampsia per se, and for the consequent fact that PE follows infection and not vice versa.

*Cystatin C*. Not least because kidney function is impaired in PE, low MW proteins may serve as biomarkers for it. To this end, cystatin C (13 kDa) has been found to be raised significantly in PE [681-687]; it also contributed to the marker set in the SCOPE study [7; 15]. Notably, although it certainly can be raised during infection [688], it seems to be more of a marker of inflammation or kidney function [689; 690].

**D-dimer**. "D-dimer" is a term used to describe quite varying forms of fibrin degradation product(s) [691]. Given that PE is accompanied by coagulopathies, it is probably not surprising that D-dimer levels are raised in PE [692-696], though this is true for many conditions [697], and some of the assays would bear improvement [698; 699]. Needless to say, however, raised D-dimer levels are also a strong marker for infection [700; 701].

Endoglin. Endoglin is the product of a gene implicated [702; 703] in the rare disease Hereditary Haemorrhagic Telangiectasia. The role of endoglin remains somewhat enigmatic [704]. However, endoglin levels were 2.5-fold higher in pre-eclamptic placentas compared to normal pregnancies (15.4 ± 2.6 versus 5.7 ± 1.0, p < 0.01). After the onset of clinical disease, the mean serum level of soluble endoglin in women with preterm PE was 46.4 ng.mL<sup>-1</sup>, as compared with 9.8 ng.mL<sup>-1</sup> in controls (P<0.001) [83]. Women with a particular endoglin SNP (AA) were 2.29 times more likely to develop PE than those with the GG genotype (P = 0.008) [705], and endoglin is seen as a reasonably good marker for PE [83; 660; 706-709] (cf. [710]). Again, endoglin levels are raised following infection by a variety of organisms [711-714], with a particularly clear example that it is a marker of infection coming from the fact that there is raised endoglin only in infected vs aseptic loosening in joints following arthroplasty [715]. In general, it seems likely that these circulating (anti)angiogenic factors are more or less markers of endothelial cell damage, just as we have described for serum ferritin [119].

*Ferritin*. The natural iron transporter in blood is transferrin (e.g. [716-721]), present at ca 1-2g.L<sup>-1</sup>, with ferritin being an intracellular iron storage molecule, so one is led to wonder why there is even any serum ferritin at all [119; 722]. The answer is almost certainly that it is a leakage molecule from damaged cells [119], and when in serum it is found to have lost its iron content [723-726]. Serum ferritin is, as expected, raised during PE [237; 239; 242; 246; 248; 727; 728] and in many other inflammatory diseases [119], including infection (e.g. [729; 730] and above).

miRNAs. MicroRNAs are a relatively novel and highly important class of ~22nt noncoding, regulatory molecules [731-734]. Some are placenta-specific, and those in the circulation (often in endo/exosomes [735-737]) can be identified during pregnancy [738-741], potentially providing a minimally invasive readout of placental condition [742-744]. There is aberrant expression of placenta-specific microRNAs (miRNAs) in PE including miR-517a/b and miR-517c [745-751] and miR-1233 [752]. C19MC is one of the largest miRNA gene clusters in humans, maps to chromosome 19q13.41, and spans a ~100 kb long region. C19MC miRNAs are processed from the cluster [753], are primate-specific, conserved in humans, and comprise 46 miRNA genes, including the miR-517 family [754]. miR-517 is known to be antiviral [755; 756], while miR-517a overexpression is apoptotic [757] and can inhibit trophoblast invasion [758]. Importantly for our argument, miR-517 molecules are overexpressed following infection [759; 760].

**Neuropeptide Y**. Although, as its name suggests, neuropeptide Y is a neurotransmitter it is also correlated with stress. Certainly it is related to noradrenaline (see below) that may itself be responsible for the raised BP in PE [761]. It is also raised in sepsis, where it is considered to counterbalance the vasodilation characteristic of septic shock (e.g. [762; 763]). The apparent paradox of a raised BP in PE and a lowered one in septic shock is considered to be related to the very different concentrations of endotoxin involved (Fig 7).

NGAL (lipocalin 2, siderocalin). NGAL (neutrophil gelatinase-associated lipocalin) is a lipocalin that is capable of binding catecholate-based siderophores (see [118; 764; 765]). As such it is anti-microbial, and is also an inflammatory or sepsis biomarker [766; 767]. Given our interest in iron, it is not surprising that it is changed during PE. While one study suggested it to be decreased in PE [768], a great many other studies showed it to be increased significantly in PE, and typically in a manner that correlated with PE severity [686; 769-777]. Pertinently to PE, it is also well established as an early biomarker of acute kidney injury (AKI) [778-781]. However, it is not a specific biomarker for AKI vs sepsis [779; 782-790] and its origin in sepsis differs [791; 792]. Of course it can be the sepsis that leads to the AKI [793; 794]. Fairly obviously, while it does tend to be increased during PE, we again see its direct role as an antimicrobial and marker of sepsis as highly supportive of our present thesis.

Placental growth factor (PIGF). This is a member of the vascular endothelial growth factor (VEGF) Family, that despite its name has a great many activities [795]. It is often considered in parallel with endoglin and sFlt, with a high sFlt:PIGF ratio being considered as especially discriminatory for PE [796-807], i.e. a lower PIGF can be diagnostic of PE [710; 808-810]. PIGF tends to be raised in sepsis unrelated to pregnancy [811; 812], while its lowering in PE may be due to the excess sFLT that decreases it [795; 813; 814]. In one study of a patient with CMV infection and PE it was in fact raised [815], while in the case of IUGR it was massively lowered [816]. PIGF alone is thus probably not a useful general marker for either PE or sepsis if one is trying to disentangle them, although it has clear promise when PE is superimposed on CKD [810; 817].

**Procalcitonin**. Procalcitonin is the 116 amino acid polypeptide precursor of calcitonin, a calcium regulatory hormone. It is another marker that has been observed to be raised (according to severity) in preeclamptics [693; 818; 819] (but cf. [820]). However, it too is a known marker of bacterial infections or sepsis [818; 821-829].

Serum amyloid A. This is an inflammatory biomarker, that was shown to increase fourfold in PE in one study [830], was significantly raised in another [819], but not in a third [831]. However, it is a well-established (and potent) biomarker for infection/sepsis (e.g. [832-845]). Defective amyloid processing may be a hallmark of PE more generally [846], and of course amyloid can be induced by various microbes [309; 311; 847; 848] and their products [250].

Soluble fms-like tyrosine kinase-1 (sFlt). The soluble fms-like tryrosine kinase (sFlt) receptor is a splice variant of the VEGF receptor [706]. It is raised considerably in PE [660; 708; 796; 801; 803; 849-852], and may be causal [525; 566; 853-856]. Needless to say, by now, we can see that it is also a very clear marker of infection [708; 857; 858], whose levels even correlate with the severity of sepsis [859-861]. Of particular note is the fact that sFLT is actually anti-inflammatory [860].

*Thrombomodulin*. Soluble thrombomodulin was recognised early as an endothelial damage biomarker, and is raised in PE [862-872]. Interestingly, it has been found to have significant efficacy in the treatment of sepsis(-based DIC) [873-881].

TLR4 upregulation in preeclamptic placentas [882] is entirely consistent with infection and the 'danger model' as applied to PE [883]. As well as LPS activation (reviewed in [133]), the heat shock protein 60 of *Chlamydia* also activates TLR4 [131].

*Visfatin* is another adipokine that is raised in PE, approximately two-fold in the study of Fasshauer and colleagues [884], and 1.5-fold in that of Adali and colleagues [885]. However, it was little different in a third study [886] while in a different study it was rather lower in pre-eclampsia than in controls [887]. This kind of phenomenon rather lights up the need for excellent quality studies, including ELISA reagents, when making assessments of this type.

Fairly obviously, the conclusion that this long list of biomarkers that are raised in PE might be <u>specific</u> 'PE' biomarkers is challenged very strongly by the finding that they are, in fact, all known markers of infection, a finding that in our view strongly bolsters the case for an infectious component in PE.

In a similar vein, there are a number of other sepsis markers (where sepsis is varied via or occurs as an independent variable) that we would predict are likely to be visible as raised in PE patient. These might include [652; 888] PAI-1, sE-selectin [889] and sVCAM-1 [859]. In particular, Presepsin looks like a

potentially useful marker for sepsis [826; 827; 890-895] but we can find no literature on its use as a PE biomarker, where we predict that it may also be raised.

#### **Metabolomic biomarkers**

For fundamental reasons connected with metabolic control and its formal, mathematical analysis [896-899], changes in the metabolome are both expected [900] and found [901-904] to be amplified relative to those in the transcriptome and proteome. For similar reasons, and coupled to evolution's selection for robustness [905-911] (i.e. homeostasis) in metabolic networks, we do not normally expect to find single metabolic biomarkers for a complex disease or syndrome. Since our initial metabolomic analyses [628], the technology has improved considerably [912-915], a full human metabolic network reconstruction has been published [911; 916-918] in the style of that done for yeast [919], and a number of candidate metabolomics biomarkers for PE have been identified reproducibly on an entirely separate validation set [14; 629].

This latter, LC-MS-based, study [14] found a cohort of 14 metabolites from the first trimester that when combined gave an OR of 23 as being predictive of third trimester PE. For convenience, we list them in Table 7. Note that because they were characterised solely via their mass there are some uncertainties in the exact identification in some cases, and that untargeted metabolomics of this type has a moderately high limit of detection (maybe  $10~\mu M$ ) such that many potentially discriminatory metabolites are below the limit of detection.

Table 7. 14 metabolites contributing to a pre-eclamptic 'signature' [14]

Metabolite	Up or down in PE	Average mass (Da)	Chemspider identifier
5-hydroxytryptophan	Down	220.225	141
Monosaccharide	Up	Unspecified	Unspecified
Decanoylcarnitine	Up	315.448	8420677
Methylglutaric and/or adipic acid	Down	146.141	11549 / 191
Oleic acid	Up	282.461	393217
Docosahexaenoic acid and/or docosatriynoic acid	Up	328.488	393183 / absent
γ-Butyrolactone and/or oxolan-3-one (Dihydrofuran-3-one)	Up	86.089	7029/ 461367
2-Oxovaleric acid and/or oxo-methylbutanoic acid	Up	116.115	67142 / absent
Acetoacetic acid	Up	102.089	94
Hexadecenoyleicosatetraenoyl- sn-glycerol	Up	n/a	Absent
Di-(octadecadienoyl)-sn- glycerol	Up	616.954	4942782
Sphingosine-1-phosphate	Up	379.472	4446673
Sphinganine 1-phosphate	Up	381.488	559277
Vitamin D₃ derivatives	Up	n/a	unspecified

A number of features of interest emerge from this.

- 1. All the markers save 5-hydroxytryptophan and adipic/methylglutaric acid were raised in PE; 5-hydroxytryptophan is a precursor of serotonin (which in some studies [920] has been seen to be mildly elevated in PE).
- 2. Markers came from multiple classes of metabolite or areas of metabolism, including amino acids, carbohydrates, carnitines, dicarboxylic acids, fatty acids (especially), (phospho)lipids and sterols.
- 3.  $\gamma$ -Butyrolactone derivatives can act as signalling molecules for a variety of bacteria [921; 922].
- 4. In common with many other inflammatory diseases [138], Vitamin D<sub>3</sub> levels (usually measured as 25(OH)vitD or calcidiol) are often lower in PE [923-927](cf. [928-930]), consistent with the levels of their derivatives being raised. However, the direction of causality inflammation ←→ vitamin D levels is not yet known [931] (see also [136; 138; 930]).
- 5. None of these metabolites was among four metabolites proposed as first trimester biomarkers in two other (smaller) studies from different groups [634; 932].
- 6. Sphingolipid metabolism can be deranged in PE [933] (also in Parkinson's [934]).

As well as the non-targeted metabolomics noted above, a number of other small molecule biomarkers have been turned up by more conventional measurements.

*Noradrenaline (norepinephrine)*. An interesting early study [935] found that venous plasma noradrenaline was raised by 67% in pre-eclamptics vs controls. Similar data were found by others [936]. This is of particular interest in the present context since noradrenaline is well established as highly growth stimulatory to Gram-negative microorganisms (e.g. [937-941]), in part by acting as a siderophore [942-944]. It also raises the levels of neuropeptide Y [761], and as a stress hormone [945], is of course well known for its role in raising blood pressure, a hallmark of PE.

There is relatively little metabolomics work in sepsis, but in one study carnitine and sphingolipid metabolism were also modified during sepsis [946], while in another [947] a suite of molecules were decreased during acute sepsis. However, the patients involved here were quite close to death, so it is not clear that comparisons between the metabolome in PE and in dying patients are that worthwhile.

We also note a recent and rather interesting suggestion by Eggers [948] that the maternal release of adrenaline (rather than noradrenaline) may have an important aetiological role in PE, although as with the rest of our thesis here it is not there indicated as to what causes the adrenaline to rise (although infection and inflammation can of course do so).

*Uric acid*. Hyperuricemia is a moderately common finding in preeclamptic pregnancies, and may even be involved in its pathogenesis (see e.g. [949-955]). However, it does not seem to be very specific [956-960], and is seemingly not an early biomarker (and it did not appear in our own study [14]). Its lack of specificity is illustrated by the fact that there is considerable evidence for the roles of purinergic signalling [961], and especially the role of uric acid, in Alzheimer's and Parkinson's disease [962-964], as well as in a variety of other kinds of inflammatory processes, including pro-inflammatory cytokine production [965; 966], the *Plasmodium falciparum*-induced inflammatory response [967], the mechanistic basis for the action of alum as an adjuvant [968], and even peanut allergy [969-971]. As is common in case-control studies when just one disease (e.g. PE) is studied, artificially high levels of sensitivity and (especially) specificity may appear when other patients with other diseases are not considered.

#### Clotting, coagulopathies and fibrinogen in PE.

In much of our previous work (e.g. [119-127]), we have noted that each of these chronic, inflammatory diseases is accompanied by changes in fibrin fibre morphologies, coagulopathies and changes in erythrocytes that are both substantial and characteristic. They can variously be mimicked by adding unliganded iron or LPS. As is well known, LPS itself is a strong inducer of coagulation, whether via tissue factor or otherwise (e.g. [972-981]), and will bind to fibrin strongly [252; 982]. The morphological methods have not yet to our knowledge been performed on blood from pre-eclamptics, whether as a diagnostic or a prognostic, though we note that clotting factors came top in one GWAS looking for gene-PE associations [111]. Fibrinogen itself is a TLR4 ligand [983], is raised in PE [984-987], and we note the extensive evidence for coagulopathies during pregnancies with PE (e.g. [25; 121; 511; 692; 988-1000]). In the worst cases these are the very frightening Disseminated Intravascular Coagulation (DIC) [980; 1001-1005], that can, of course, also emerge as a consequence of sepsis [1006-1012]. Variations in the plasminogen activator inhibitor-1 may contribute to the hypofibrinolysis observed [1013-1015].

We recently showed that LPS can potently induce amyloid formation in fibrin (see [251; 1016]). Thus, In addition, we note the increasing recognition that amyloid proteins themselves, that may occur as a result of coagulaopathies, are themselves both inflammatory (e.g. [540; 640; 1017-1022]) and cytotoxic (e.g. [250; 1023-1027]), and this that can of itself contribute strongly to the death of e.g. trophoblasts.

Related to clotting parameters are three other 'old' but easily measured variables that probably reflect inflammation [1028] and that have been suggested to differ in PE from normotensives and may have some predictive power. The first two are the erythrocyte sedimentation rate (ESR) [1029; 1030] and the red cell distribution width (RDW) [1031] (but cf. [1032]). Interestingly, the former was the only variable that was predictive of a subsequent stroke following sub-arachnoid haemorrhage [1033]. The third relates to the morphology of erythrocytes (that may in part underpin the other two). We and others have shown in a series of studies (e.g. [127-129; 1034-1037] that erythrocyte morphology diverges very considerably from that 'classical' discoid shape adopted by normal healthy cells, and that this can be a strong indicator of disease [130]. In extreme cases (e.g. [126; 1038-1043]), including following infection [1044], this results in eryptosis, the suicidal death of erythrocytes. It is of interest that ceramide, a precursor of sphingosine-1-phosphate (Table 7), is raised in various diseases such as Parkinson's and may serve to stimulate eryptosis [1045]. Although we know of no direct measurements to date, there is evidence that eryptosis may play a significant role in PE [1046]

**Prevention Strategies**. Apart from low-dose aspirin (that may have little effect [1047-1050] unless initiated relatively early in pregnancy [1051-1055]), and low-dose calcium [1056], there are relatively few treatment options in present use [1057-1060]. (Magnesium sulphate [1061-1063] has been used as a treatment for eclampsia.)

In the history of science or medicine, some treatments are empirical, while others are considered to have a mechanistic basis. The general assumption is that the more we know about the originating aetiology of a disease or syndrome the more likely we are to be able to treat its causes effectively, and not just its symptoms. Clearly, too, clinicians are rightly loth to give complex and potentially teratogenic treatments to pregnant women when this can be avoided [1064; 1065]. However, the surprising lack of systematic data with antibiotics [1066], modulo one particularly spectacular success [521], suggests that we ought to be performing trials with safe antibiotics on women at special risk [1067]. These must take care to avoid any Jarisch-Herxheimer reaction [1068-1070] due to the release from microbes induced by antibiotics of inflammagens like LPS [1071-1074]. A related strategy recognises that some FDA-approved drugs can actually exert powerful antibiotic effects *in vivo* (but not on petri plates) by modifying the host [1075].

Because of the known oxidative stress accompanying PE, it had been assumed that antioxidants such as vitamin C (ascorbate) might be preventive; however, this turned out not to be the case (even the opposite) for ascorbate [1047; 1076]. Probably this is because in the presence of unliganded iron, ascorbate is in fact pro-oxidant [118]. However, polyphenolic antioxidants that actually act by chelating iron [79; 118] seem to be more effective [1077].

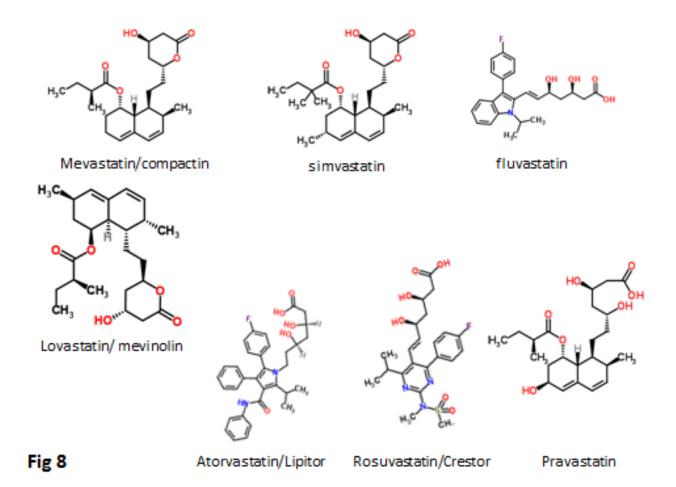
Another area that we and others have previously highlighted recognises the ability of non-siderophoric iron chelators to act as iron-withholding agents and thereby limit the growth of bacteria. Again, a prediction is that women with iron overload diseases should be more susceptible to pre-eclampsia, a prediction that is borne out for  $\alpha$ -thalassaemia [1078; 1079] though not apparently for hereditary haemochromatosis [1080]. However, the extent of use of chelators and degree of control of free iron thereby obtained is rarely recorded in any detail, so in truth it is difficult to draw conclusions.

The general benefits of nutritional iron chelators such as blueberries and other fruits and vegetables containing anthocyanins have been discussed elsewhere (e.g. [79; 118; 1081]).

How significant coagulopathies are to the aetiology of PE development (as opposed to providing merely an accompaniment) is not entirely clear, but on the basis that they are then anticoagulants would potentially assist, just as thrombomodulin does in DIC accompanying sepsis [879; 881; 1008; 1012]. Of course one of many effects of low-dose aspirin is to act as an anticoagulant. There is also evidence for the efficacy of heparin [5; 1058; 1082-1087], which is especially interesting given our highlighting of the role of coagulopathies in PE. Those anticoagulants that avoid bleeding [1088] are obviously of particular interest, while anything stopping the fibrin forming  $\beta$ -amyloid [251; 252] should serve as an especially useful anti-inflammatory anticoagulant.

With a change in focus from function-first to target-first-based drug discovery [909], there has been an assumption that because a drug is (i) found to bind potently to a molecular target and (ii) has efficacy at a physiological level *in vivo*, the first process is thus responsible for the second. This has precisely no basis in logic (it is a logical fault known variously as "affirming the consequent" or "post hoc ergo propter hoc" [1089]). This is because the drug might be acting physiologically by any other means, since drug binding to proteins is typically quite promiscuous (e.g. [1090-1094]). Indeed, the average known number of binding sites for marketed drugs is six [201; 1095]. In particular, it is likely, from a network or systems pharmacology perspective (e.g. [911; 1096-1099], that successful drugs (like aspirin) are successful precisely because they hit multiple targets. The so-called 'statins' provide a particularly good case in point [118].

It had long been known that the enzyme HMGCoA reductase exerted strong control on the biosynthetic flux to cholesterol, and that inhibiting it might lower the flux and steady-state cholesterol levels (as indeed it does). Notwithstanding that cholesterol alone is a poor predictor of cardiovascular disease [1100-1102]), especially in the normal range, HMGCoA reductase inhibitors have benefits in terms of decreasing the adverse events of various types of cardiovascular disease [1103]. Following an original discovery of natural products such as compactin (mevastatin) and lovastatin containing a group related to hydroxymethylglutaric acid (rather than a CoA version) that inhibited the enzyme [1104]), many variants with this (hydroxyl)methylglutaric substructure came to be produced, with the much larger 'rest' of the molecule being considerably divergent (see Fig 8, where the MW values vary from 390.5 (mevastatin) to 558.6 (atorvastatin)). Despite this wide structural diversity (Fig 8) they are still collectively known as 'statins', and despite the wildly illogical assumption that they might all work in the same way(s). The fact that different statins can cause a variety of distinct expression profiles [1105] is anyway utterly inconsistent with a unitary mode of action. In particular, in this latter study, statins clustered into whether they were (fluvastatin, lovastatin and simvastatin) or were not (atorvastatin, pravastatin and rosuvastatin) likely to induce the side effect of rhabdomyolysis or any other myopathy. Clearly, any choice of 'statin' should come from the latter group, with pravastatin and rosuvastatin being comparatively hydrophilic.



The epidemiological fact of improved survival despite the comparative irrelevance of cholesterol levels to atherosclerotic plaque formation and heart disease in the normal range provides an apparent paradox [1106]. This is easily solved by the recognition (e.g. [1107-1120], and many other references and reviews) that 'statins' are in fact anti-inflammatory. They may also be antimicrobial/anti-septic, whether directly or

otherwise [1121-1125], and we also note the role of cholesterol in mopping up endotoxin [1126]. Finally, here, it needs to be recognised that statins do themselves serve to lower iron levels [1127-1129], and (while oddly this seems not to have been tested directly) simple inspection of their structures (Fig 8) implies that the better ones (with their multiple OH groups) might in fact chelate iron directly.

In consequence, a number of authors have indicated the potential utility of statins in treating PE [107; 525; 1130-1140], and pravastatin has been the subject of a number of favourable studies [525; 1131; 1133; 1136; 1138; 1141; 1142], including in humans [1131; 1143-1145]. Pravastatin seems more than ripe for a proper, randomised clinical trial [1130].

Another 'vascular' class of drugs that has been proposed for treating PE is represented by those of the family of vasodilatory phosphodiesterase5 inhibitors such as sildenafil (Viagra) and vardenafil (Levitra), as it is reasonable that they might improve endothelial function, especially if started early in pregnancy [1146]. Thus vardefanil restores endothelial function by increasing placental growth factor [1147], and sildenafil has shown promise in a number of animal studies [1148-1153] and in human tissues [1154; 1155], with a clinical trial ongoing [1156]. In particular [1153], it was able to normalise the metabolomics changes observed in a mouse model (the COMT<sup>-/-</sup> model) of PE.

Antihypertensive therapy for PE has been reviewed by Abalos and colleagues [1157] and Magee and colleagues [108]. Anti-hypertensives did halve the incidence of hypertension but had no effect on PE. Methyldopa is one of the most commonly used anti-hypertensives in pregnancy, but it may also stimulate eryptosis [1158]; alternative drugs were considered to be better [1157] for hypertension. Nifedipine [1159] and labetalol [1160] are considered a reasonable choice. There was also a slight reduction in the overall risk of developing proteinuria/pre-eclampsia when beta blockers and calcium channel blockers considered together (but not alone) were compared with methyldopa [1157]. In mice, olmesartan (together with captopril) proved usefully anti-hypertensive [1161]; this is of interest because olmesartan is also an agonist of the vitamin D receptor [1162]. However, it was not mentioned in either [1157] or [108].

LPS itself has long been recognised as a target of inflammatory diseases. Unfortunately, despite initially promising trials of an anti-LPS antibody known as Centoxin [1163], it was eventually withdrawn, apparently because of a combination of ineffectiveness [1164; 1165] and toxicity [1166; 1167]. LPS is rather hydrophobic, and thus it is hard to make even monoclonal antibodies very selective for such targets, such that the toxicity was probably because of its lack of specificity between lipid A and other hydrophobic ligands [1168]. Other possible treatments based on LPS, such as 'sushi peptides' [1169-1176] (or variants [1177; 1178]) and LPS-binding protein were covered elsewhere [133].

If an aberrant or dysbiotic gut microbiome is the source of the microbes that underpin PE, it is at least plausible that the gut microbiome should be predictive of PE [370], but we know of no suitably powered study that has been done to assess this, and this would clearly be worthwhile. However, in a study of primiparous women, the OR for getting severe PE was only 0.6 if probiotic milk drinks containing lactobacilli were consumed daily [1179]. This too seems an area well worth following up.

From a metabolomics point of view, the molecules seen to be raised in PE may either be biomarkers of the disease aetiology <u>or</u> of the body's attempts to respond to the disease (and this is true generally [1180]). Thus it is of great interest that sphingosine-1-phosphate (S1P) was raised in PE (see [14] and Table 7). S1P is mainly vasoconstrictive [1181; 1182], but agonists of the sphingosine-1-phosphate1 receptor (that is involved in endothelial cell function) seemed to have considerable value in combatting the cytokine storm that followed infection-driven sepsis [1183-1188]. The detailed mechanism seems not to be known, but in the context of infection, a need for S1P and other sphingolipids for successful pregnancies [1189; 1190]

(see also Parkinson's [934]), and the induction of PE by its disruption [933; 1191-1195]), some serious investigation of the potential protective effects of S1PR1 agonists seems highly warranted.

Among other small molecules, melatonin has shown some promise in the treatment of septic shock, by lowering inflammatory cytokine production [1196] (and see [1197] for neonatal oxidative stress), and a trial is in prospect for PE [1198].

Lipoxin  $A_4$  (LXA<sub>4</sub>) is considered to be an endogenous stop signal in inflammation. While recognising the difficulties with rodent PE models (above), we note that in one study, the effect of BML-111 (a synthetic analogue of LXA<sub>4</sub>) was tested on experimental PE induced in rats by low-dose endotoxin (LPS), and showed highly beneficial effects [530].

Coda – a return to the Bradford Hill criteria. Returning to the Bradford Hill criteria for ascribing causation of a disease to an environmental factor [91], we can now ask whether a detectable (if largely dormant) microbiome X, that is more likely to replicate with free iron, and can anyway secrete or shed a variety of inflammatory components such as LPS, represents a plausible and major aetiological factor for PE (Y):

- (1) what is the strength of association between X and Y? We found an overwhelming co-occurrence of microbes or their products and PE
- (2) what is the consistency of association between X and Y? Almost wherever we looked, whether via periodontal disease, urinary tract infection, or other means of ingress, we could find a microbial component in PE
- (3) What is the specificity of association between X and Y? Insufficient data are available to ascribe PE solely to one type of organism; by contrast the data rather indicate that a variety of microbes, each capable of shedding inflammatory molecules such as LPS, can serve to stimulate or exacerbate PE.
- (4) experiments verify the relationship between X and Y. It is unethical to do these in humans in terms of purposely infecting pregnant women, but data from antibiotics show the expected improvements.
- (5) modification of X alters the occurrence of Y; this is really as (4)
- (6) biologically plausible cause and effect relationship. Yes, this is where we think the ideas set down here are entirely consistent with current thinking on the main causes of PE. What we add in particular is the recognition that bacteria (and other microbes) that may be invisible to culture are both present and responsible, by established means, for the inflammation and other sequelae (and especially the coagulopathies) seen as causative accompaniments to PE.

#### Other predictions

Classical clinical microbiology, involving mainly replication-based methods, is evolving rapidly to assess the microbial content of samples on the basis of DNA sequences [288; 1199], including 16S rDNA [279; 280; 282; 284; 285; 287; 289; 1200], suitable protein-encoding housekeeping genes (e.g. [1201-1206]), and, increasingly, full genome sequences [1207]. In the future, we can thus expect a considerable increase in molecular assessments of the microbiological content of blood, urine and tissues, and this will obviously be a vital part of the experimental assessment and development of the ideas presented here. Molecular methods will also be used to assess maternal circulating DNA [1208-1210] and RNA [1211] in terms of both its presence and sequencing, as well as the use of digital PCR [1212].

Since PE has such a strong vascular component, we also predict that measurements designed to detect coagulopathies will increase in importance, for both diagnosis and prognosis, and for assessing treatments.

New drugs designed to kill <u>non-growing</u> bacteria [1213-1217] or to overcome amyloid coagulopathies [1218-1222] will be needed, and will come to the fore.

Finally, we consider that real progress in understanding PE from a systems biology perspective means that it must be modelled accordingly, and this must be a major goal.

**Concluding remarks**. We have brought together a large and widely dispersed literature to make the case that an important aetiological role in pre-eclampsia is played by dormant microbes, or at least ones that are somewhat refractory to culture, and that these can awaken, shed inflammagens such as LPS, and thereby initiate inflammatory cascades. (The sequelae of these, involving cytokines, coagulopathies, and so on, are well enough accepted.) The case is founded on a large substructure of interlocking evidence, but readers might find the following elements as discussed above especially persuasive and/or worthy of follow-up:

- The regular presence of detectable microbes in pre-eclamptic placentas (e.g. [395; 418; 419])
- The fact that endotoxin (LPS) can act as such a mimic for invoking PE in experimental models
- The fact that every known proteomic biomarker suggested for PE has also been shown to increase during infection.
- The significant number of papers reviewing a link between infection and PE (e.g. [66; 67; 131; 363])
- The almost complete absence (one case) of PE in patients treated with spiramycin [521]

Any and all of these provide powerful strategies for testing whether PE is, in fact, like gastric ulcers [159; 161; 162; 1223], essentially initiated as an infectious disease.

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## **Legends to Figures**

Figures (as aide memoire for us)

- Figure 1. Two main sources (foetal and maternal) can drive a pregnancy towards pre-eclampsia.
- Figure 2. There are four main 'causes' of pre-eclampsia, represented by the coloured outer circles, and these too can interact with each other. That part of the figure is redrawn from [107]. In addition, we note here, as the theme of this review, that microbes can themselves cause each of the features in the outer coloured circles to manifest.
- Figure 3. Another detailed representation of factors known to cause or accompany PE, redrawn from [108]
- Figure 4. A mind map of the overall structure of the review
- Figure 5. The chief physiological macrostates exhibited by microorganisms
- Figure 6. An 11-stage systems biology model of the factors that we consider cause initially formant microbes to manifest the symptoms (and disease) of pre-eclampsia
- Figure 7. Pre-eclampsia bears some similarities to, and may be considered as a milder form of, the changes that occur during genuine sepsis leading to a systematic inflammatyory response syndrome, septic shock and multiple organ disfunction.
- Figure 8. Some structures of various statins

## **Legends to Tables**

- Table 1. Some known risk factors for pre-eclampsia (based in part on [2; 6; 35-37]). See also <a href="http://bestpractice.bmj.com/best-practice/monograph/326/diagnosis.html">http://bestpractice.bmj.com/best-practice/monograph/326/diagnosis.html</a>. Note that most of these are risk factors that might and do pre-dispose for other diseases (or are themselves diseases).
- Table 2. Many studies have identified a much greater prevalence of infectious agents in the blood or urine of those exhibiting PE than in matched controls
- Table 3. Evidence for microbes in placental tissues, including those with PE.
- Table 4. Periodontal disease (PD) and pre-eclampsia
- Table 5. Examples of decreased PE following antibiotic therapies given for various reasons
- Table 6. Molecular examples of bacterial antigens that can elicit antibodies that stop successful trophoblast implantation or stimulate parturition.
- Table 7. 14 metabolites contributing to a pre-eclamptic 'signature' [14]

## References

- [1] Steegers, E. A. P., von Dadelszen, P., Duvekot, J. J. & Pijnenborg, R. (2010). Pre-eclampsia. *Lancet* **376**, 631-44.
- [2] North, R. A., McCowan, L. M., Dekker, G. A., Poston, L., Chan, E. H., Stewart, A. W., Black, M. A., Taylor, R. S., Walker, J. J., Baker, P. N. & Kenny, L. C. (2011). Clinical risk prediction for pre-eclampsia in nulliparous women: development of model in international prospective cohort. *BMJ* **342**, d1875.
- [3] Uzan, J., Carbonnel, M., Piconne, O., Asmar, R. & Ayoubi, J. M. (2011). Pre-eclampsia: pathophysiology, diagnosis, and management. *Vasc Health Risk Manag* **7**, 467-74.
- [4] Kenny, L. (2012). Improving diagnosis and clinical management of pre-eclampsia. *MLO Med Lab Obs* **44**, 12, 14.
- [5] Desai, P. (2013). Obstetric vasculopathies. Jaypee, New Delhi.
- [6] Chaiworapongsa, T., Chaemsaithong, P., Yeo, L. & Romero, R. (2014). Pre-eclampsia part 1: current understanding of its pathophysiology. *Nat Rev Nephrol* **10**, 466-80.
- [7] Kenny, L. C., Black, M. A., Poston, L., Taylor, R., Myers, J. E., Baker, P. N., McCowan, L. M., Simpson, N. A. B., Dekker, G. A., Roberts, C. T., Rodems, K., Noland, B., Raymundo, M., Walker, J. J. & North, R. A. (2014). Early pregnancy prediction of preeclampsia in nulliparous women, combining clinical risk and biomarkers: the Screening for Pregnancy Endpoints (SCOPE) international cohort study. *Hypertension* 64, 644-52.
- [8] Sircar, M., Thadhani, R. & Karumanchi, S. A. (2015). Pathogenesis of preeclampsia. *Curr Opin Nephrol Hypertens* **24,** 131-8.
- [9] Abalos, E., Cuesta, C., Grosso, A. L., Chou, D. & Say, L. (2013). Global and regional estimates of preeclampsia and eclampsia: a systematic review. *Eur J Obstet Gynecol Reprod Biol* **170**, 1-7.
- [10] Khan, K. S., Wojdyla, D., Say, L., Gulmezoglu, A. M. & Van Look, P. F. (2006). WHO analysis of causes of maternal death: a systematic review. *Lancet* **367**, 1066-74.
- [11] Duley, L. (2009). The global impact of pre-eclampsia and eclampsia. Semin Perinatol 33, 130-7.
- [12] Abalos, E., Cuesta, C., Carroli, G., Qureshi, Z., Widmer, M., Vogel, J. P., Souza, J. P. & W. H. O. Multicountry Survey on Maternal Newborn Health Research Network. (2014). Pre-eclampsia, eclampsia and adverse maternal and perinatal outcomes: a secondary analysis of the World Health Organization Multicountry Survey on Maternal and Newborn Health. *BJOG* 121 Suppl 1, 14-24.
- [13] Ghulmiyyah, L. & Sibai, B. (2012). Maternal mortality from preeclampsia/eclampsia. *Semin Perinatol* **36,** 56-9.
- [14] Kenny, L. C., Broadhurst, D. I., Dunn, W., Brown, M., Francis-McIntyre, S., North, R. A., McGowan, L., Roberts, C., Cooper, G. J. S., Kell, D. B. & Philip N Baker on behalf of the SCOPE consortium. (2010). Robust early pregnancy prediction of later preeclampsia using metabolomic biomarkers *Hypertension* **56**, 741-749.
- [15] Karumanchi, S. A. & Granger, J. P. (2016). Preeclampsia and Pregnancy-Related Hypertensive Disorders. *Hypertension* **67**, 238-42.
- [16] Irgens, H. U., Reisaeter, L., Irgens, L. M. & Lie, R. T. (2001). Long term mortality of mothers and fathers after pre-eclampsia: population based cohort study. *BMJ* **323**, 1213-7.
- [17] Bellamy, L., Casas, J. P., Hingorani, A. D. & Williams, D. J. (2007). Pre-eclampsia and risk of cardiovascular disease and cancer in later life: systematic review and meta-analysis. *BMJ* **335**, 974.
- [18] Craici, I. M., Wagner, S. J., Hayman, S. R. & Garovic, V. D. (2008). Pre-eclamptic pregnancies: an opportunity to identify women at risk for future cardiovascular disease. *Womens Health (Lond Engl)*4. 133-5
- [19] Romundstad, P. R., Magnussen, E. B., Smith, G. D. & Vatten, L. J. (2010). Hypertension in pregnancy and later cardiovascular risk: common antecedents? *Circulation* **122**, 579-84.

- [20] Powe, C. E., Levine, R. J. & Karumanchi, S. A. (2011). Preeclampsia, a disease of the maternal endothelium: the role of antiangiogenic factors and implications for later cardiovascular disease. *Circulation* **123**, 2856-69.
- [21] Skjaerven, R., Wilcox, A. J., Klungsøyr, K., Irgens, L. M., Vikse, B. E., Vatten, L. J. & Lie, R. T. (2012). Cardiovascular mortality after pre-eclampsia in one child mothers: prospective, population based cohort study. *BMJ* **345**, e7677.
- [22] Ananth, C. V. & Lawrence Cleary, K. (2013). Pre-eclampsia and cardiovascular disease: more questions than answers? *BJOG* **120**, 920-3.
- [23] Brennan, L. J., Morton, J. S. & Davidge, S. T. (2014). Vascular dysfunction in preeclampsia. *Microcirculation* **21**, 4-14.
- [24] Chen, C. W., Jaffe, I. Z. & Karumanchi, S. A. (2014). Pre-eclampsia and cardiovascular disease. *Cardiovasc Res* **101**, 579-86.
- [25] Tannetta, D. S., Hunt, K., Jones, C. I., Davidson, N., Coxon, C. H., Ferguson, D., Redman, C. W. G., Gibbins, J. M., Sargent, I. L. & Tucker, K. L. (2015). Syncytiotrophoblast Extracellular Vesicles from Pre-Eclampsia Placentas Differentially Affect Platelet Function. *PLoS One* 10, e0142538.
- [26] Mol, B. W. J., Roberts, C. T., Thangaratinam, S., Magee, L. A., de Groot, C. J. M. & Hofmeyr, G. J. (2016). Pre-eclampsia. *Lancet* **387**, 999-1011.
- [27] Bateman, B. T., Schumacher, H. C., Bushnell, C. D., Pile-Spellman, J., Simpson, L. L., Sacco, R. L. & Berman, M. F. (2006). Intracerebral hemorrhage in pregnancy: frequency, risk factors, and outcome. *Neurology* **67**, 424-9.
- [28] Kajantie, E., Eriksson, J. G., Osmond, C., Thornburg, K. & Barker, D. J. P. (2009). Pre-eclampsia is associated with increased risk of stroke in the adult offspring: the Helsinki birth cohort study. *Stroke* **40**, 1176-80.
- [29] Tang, C. H., Wu, C. S., Lee, T. H., Hung, S. T., Yang, C. Y., Lee, C. H. & Chu, P. H. (2009). Preeclampsia-eclampsia and the risk of stroke among peripartum in Taiwan. *Stroke* **40**, 1162-8.
- [30] Harel, M., Aron-Maor, A., Sherer, Y., Blank, M. & Shoenfeld, Y. (2005). The infectious etiology of the antiphospholipid syndrome: links between infection and autoimmunity. *Immunobiology* **210**, 743-7.
- [31] Shoenfeld, Y., Blank, M., Cervera, R., Font, J., Raschi, E. & Meroni, P. L. (2006). Infectious origin of the antiphospholipid syndrome. *Ann Rheum Dis* **65**, 2-6.
- [32] Sherer, Y., Blank, M. & Shoenfeld, Y. (2007). Antiphospholipid syndrome (APS): where does it come from? *Best Pract Res Clin Rheumatol* **21,** 1071-8.
- [33] Sène, D., Piette, J. C. & Cacoub, P. (2008). Antiphospholipid antibodies, antiphospholipid syndrome and infections. *Autoimmun Rev* **7**, 272-7.
- [34] Mignini, L. E., Villar, J. & Khan, K. S. (2006). Mapping the theories of preeclampsia: the need for systematic reviews of mechanisms of the disease. *Am J Obstet Gynecol* **194**, 317-21.
- [35] Duckitt, K. & Harrington, D. (2005). Risk factors for pre-eclampsia at antenatal booking: systematic review of controlled studies. *British Medical Journal* **330**, 565-567.
- [36] Baker, P. N. & Kenny, L. C. (2011). Obstetrics by ten teachers. CRC Press, Boca Raton, FL.
- [37] Ananth, C. V., Keyes, K. M. & Wapner, R. J. (2013). Pre-eclampsia rates in the United States, 1980-2010: age-period-cohort analysis. *BMJ* **347**, f6564.
- [38] Bdolah, Y., Elchalal, U., Natanson-Yaron, S., Yechiam, H., Bdolah-Abram, T., Greenfield, C., Goldman-Wohl, D., Milwidsky, A., Rana, S., Karumanchi, S. A., Yagel, S. & Hochner-Celnikier, D. (2014). Relationship between nulliparity and preeclampsia may be explained by altered circulating soluble fms-like tyrosine kinase 1. *Hypertens Pregnancy* **33**, 250-9.
- [39] Saftlas, A. F., Olson, D. R., Franks, A. L., Atrash, H. K. & Pokras, R. (1990). Epidemiology of Preeclampsia and Eclampsia in the United-States, 1979-1986. *Am J Obstet Gynecol* **163**, 460-465.
- [40] Zhang, J., Zeisler, J., Hatch, M. C. & Berkowitz, G. (1997). Epidemiology of pregnancy-induced hypertension. *Epidemiol Rev* **19**, 218-32.
- [41] Lamminpää, R., Vehvilainen-Julkunen, K., Gissler, M. & Heinonen, S. (2012). Preeclampsia complicated by advanced maternal age: a registry-based study on primiparous women in Finland 1997-2008. BMC Pregnancy Childbirth 12, 47.
- [42] Carolan, M. (2013). Maternal age >= 45 years and maternal and perinatal outcomes: A review of the evidence. *Midwifery* **29**, 479-489.

- [43] Trogstad, L., Magnus, P. & Stoltenberg, C. (2011). Pre-eclampsia: Risk factors and causal models. *Best Pract Res Clin Obstet Gynaecol* **25**, 329-42.
- [44] Coonrod, D. V., Hickok, D. E., Zhu, K. M., Easterling, T. R. & Daling, J. R. (1995). Risk-Factors for Preeclampsia in Twin Pregnancies a Population-Based Cohort Study. *Obstet Gynecol* **85**, 645-650.
- [45] Campbell, D. M. & MacGillivray, I. (1999). Preeclampsia in twin pregnancies: Incidence and outcome. *Hypertens Pregnancy* **18**, 197-207.
- [46] Bdolah, Y., Lam, C., Rajakumar, A., Shivalingappa, V., Mutter, W., Sachs, B. P., Lim, K. H., Bdolah-Abram, T., Epstein, F. H. & Karumanchi, S. A. (2008). Twin pregnancy and the risk of preeclampsia: bigger placenta or relative ischemia? *Am J Obstet Gynecol* **198**.
- [47] Bodnar, L. M., Ness, R. B., Harger, G. F. & Roberts, J. M. (2005). Inflammation and triglycerides partially mediate the effect of prepregnancy body mass index on the risk of preeclampsia. *Am J Epidemiol* **162**, 1198-206.
- [48] Roberts, J. M., Bodnar, L. M., Patrick, T. E. & Powers, R. W. (2011). The role of obesity in preeclampsia. *Pregnancy Hypertens* **1,** 6-16.
- [49] Jeyabalan, A. (2013). Epidemiology of preeclampsia: impact of obesity. Nutr Rev 71, S18-S25.
- [50] Sugerman, H. J. (2014). Effect of obesity on incidence of preeclampsia. *American Journal of Obstetrics and Gynecology* **210**, 375-375.
- [51] Thornton, J. G. & Macdonald, A. M. (1999). Twin mothers, pregnancy hypertension and pre-eclampsia. *Brit J Obstet Gynaec* **106**, 570-575.
- [52] Ros, H. S., Lichtenstein, P., Lipworth, L. & Cnattingius, S. (2000). Genetic effects on the liability of developing pre-eclampsia and gestational hypertension. *American Journal of Medical Genetics* **91**, 256-260.
- [53] Williams, P. J. & Broughton Pipkin, F. (2011). The genetics of pre-eclampsia and other hypertensive disorders of pregnancy. *Best Pract Res Clin Obstet Gynaecol* **25**, 405-17.
- [54] Valenzuela, F. J., Perez-Sepulveda, A., Torres, M. J., Correa, P., Repetto, G. M. & Illanes, S. E. (2012). Pathogenesis of preeclampsia: the genetic component. *J Pregnancy* **2012**, 632732.
- [55] Boyd, H. A., Tahir, H., Wohlfahrt, J. & Melbye, M. (2013). Associations of Personal and Family Preeclampsia History With the Risk of Early-, Intermediate- and Late-Onset Preeclampsia. *Am J Epidemiol* **178**, 1611-1619.
- [56] Roten, L. T., Thomsen, L. C. V., Gundersen, A. S., Fenstad, M. H., Odland, M. L., Strand, K. M., Solberg, P., Tappert, C., Araya, E., Bærheim, G., Lyslo, I., Tollaksen, K., Bjørge, L. & Austgulen, R. (2015). The Norwegian preeclampsia family cohort study: a new resource for investigating genetic aspects and heritability of preeclampsia and related phenotypes. *Bmc Pregnancy and Childbirth* 15.
- [57] Asherson, R. A. & Shoenfeld, Y. (2000). The role of infection in the pathogenesis of catastrophic antiphospholipid syndrome--molecular mimicry? *J Rheumatol* **27**, 12-4.
- [58] Asherson, R. A. & Cervera, R. (2003). Antiphospholipid antibodies and infections. *Ann Rheum Dis* **62**, 388-93.
- [59] Blank, M., Asherson, R. A., Cervera, R. & Shoenfeld, Y. (2004). Antiphospholipid syndrome infectious origin. *J Clin Immunol* **24**, 12-23.
- [60] Clark, E. A., Silver, R. M. & Branch, D. W. (2007). Do antiphospholipid antibodies cause preeclampsia and HELLP syndrome? *Curr Rheumatol Rep* **9**, 219-25.
- [61] Espinosa, G., Cervera, R. & Asherson, R. A. (2007). Catastrophic antiphospholipid syndrome and sepsis. A common link? *J Rheumatol* **34**, 923-6.
- [62] Zinger, H., Sherer, Y., Goddard, G., Berkun, Y., Barzilai, O., Agmon-Levin, N., Ram, M., Blank, M., Tincani, A., Rozman, B., Cervera, R. & Shoenfeld, Y. (2009). Common infectious agents prevalence in antiphospholipid syndrome. *Lupus* **18**, 1149-53.
- [63] Kutteh, W. H. (2014). Antiphospholipid antibody syndrome and reproduction. *Curr Opin Obstet Gynecol* **26**, 260-265.
- [64] Kutteh, W. H. & Hinote, C. D. (2014). Antiphospholipid Antibody Syndrome. *Obstet Gyn Clin N Am* **41**, 113-+.
- [65] Schieve, L. A., Handler, A., Hershow, R. & Davis, F. (1994). Urinary-Tract Infection during Pregnancy Its Association with Maternal Morbidity and Perinatal Outcome. *American Journal of Public Health* **84**, 405-410.

- [66] Conde-Agudelo, A., Villar, J. & Lindheimer, M. (2008). Maternal infection and risk of preeclampsia: systematic review and metaanalysis. *Am J Obstet Gynecol* **198,** 7-22.
- [67] Rustveld, L. O., Kelsey, S. F. & Sharma, R. (2008). Association between maternal infections and preeclampsia: a systematic review of epidemiologic studies. *Matern Child Health J* 12, 223-42.
- [68] Redman, C. W. G. (1991). Current topic: pre-eclampsia and the placenta. Placenta 12, 301-8.
- [69] Burton, G. J. & Jauniaux, E. (2004). Placental oxidative stress: from miscarriage to preeclampsia. *J Soc Gynecol Investig* **11**, 342-352.
- [70] Redman, C. W. G. & Sargent, I. L. (2005). Latest advances in understanding preeclampsia. *Science* **308**, 1592-1594.
- [71] Roberts, J. M. & Bell, M. J. (2013). If we know so much about preeclampsia, why haven't we cured the disease? *J Reprod Immunol* **99**, 1-9.
- [72] Redman, C. W., Sargent, I. L. & Staff, A. C. (2014). IFPA Senior Award Lecture: making sense of pre-eclampsia two placental causes of preeclampsia? *Placenta* **35 Suppl**, S20-5.
- [73] Xiong, X., Demianczuk, N. N., Saunders, L. D., Wang, F. L. & Fraser, W. D. (2002). Impact of Preeclampsia and gestational hypertension on birth weight by gestational age. Am J Epidemiol 155, 203-209.
- [74] Roberts, J. M. & Hubel, C. A. (2009). The two stage model of preeclampsia: variations on the theme. *Placenta* **30 Suppl A,** S32-7.
- [75] Redman, C. W. G. (2014). The six stages of pre-eclampsia. Pregnancy Hypertens 4, 246.
- [76] Sacks, G. P., Studena, K., Sargent, K. & Redman, C. W. G. (1998). Normal pregnancy and preeclampsia both produce inflammatory changes in peripheral blood leukocytes akin to those of sepsis. *Am J Obstet Gynecol* **179**, 80-6.
- [77] Redman, C. W. G. & Sargent, I. L. (2003). Pre-eclampsia, the placenta and the maternal systemic inflammatory response--a review. *Placenta* **24 Suppl A,** S21-7.
- [78] Hubel, C. A. (2006). Dyslipidemia and pre-eclampsia. In *Pre-eclampsia-aetiology and clinical practice*. (ed. M. A. Belfort and F. Lydall), pp. 164–182. Cambridge University Press, Cambridge.
- [79] Kell, D. B. (2010). Towards a unifying, systems biology understanding of large-scale cellular death and destruction caused by poorly liganded iron: Parkinson's, Huntington's, Alzheimer's, prions, bactericides, chemical toxicology and others as examples. *Arch Toxicol* **577**, 825-889.
- [80] Yancopoulos, G. D., Davis, S., Gale, N. W., Rudge, J. S., Wiegand, S. J. & Holash, J. (2000). Vascular-specific growth factors and blood vessel formation. *Nature* **407**, 242-248.
- [81] Maynard, S., Epstein, F. H. & Karumanchi, S. A. (2008). Preeclampsia and angiogenic imbalance. *Annual Review of Medicine* **59**, 61-78.
- [82] Levine, R. J., Maynard, S. E., Qian, C., Lim, K. H., England, L. J., Yu, K. F., Schisterman, E. F., Thadhani, R., Sachs, B. P., Epstein, F. H., Sibai, B. M., Sukhatme, V. P. & Karumanchi, S. A. (2004). Circulating angiogenic factors and the risk of preeclampsia. *N Engl J Med* **350**, 672-83.
- [83] Levine, R. J., Lam, C., Qian, C., Yu, K. F., Maynard, S. E., Sachs, B. P., Sibai, B. M., Epstein, F. H., Romero, R., Thadhani, R. & Karumanchi, S. A. (2006). Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *N Engl J Med* **355**, 992-1005.
- [84] Entman, S. S. & Richardson, L. D. (1983). Clinical applications of the altered iron kinetics of toxemia of pregnancy. *Am J Obs Gynecol* **146**, 568-574.
- [85] MacDonald, A. B. (1986). Human fetal borreliosis, toxemia of pregnancy, and fetal death. *Zentralbl Bakteriol Mikrobiol Hyg A* **263**, 189-200.
- [86] Loudon, I. (1991). Some historical aspects of toxaemia of pregnancy. A review. *Br J Obstet Gynaecol* **98**, 853-8.
- [87] Meziani, F., Tesse, A., David, E., Martinez, M. C., Wangesteen, R., Schneider, F. & Andriantsitohaina, R. (2006). Shed membrane particles from preeclamptic women generate vascular wall inflammation and blunt vascular contractility. *Am J Pathol* **169**, 1473-83.
- [88] Sugawara, J., Mitsui-Saito, M., Hayashi, C., Hoshiai, T., Senoo, M., Chisaka, H., Yaegashi, N. & Okamura, K. (2005). Decrease and senescence of endothelial progenitor cells in patients with preeclampsia. J Clin Endocrinol Metab 90, 5329-32.
- [89] Wang, Y. & Walsh, S. W. (1998). Placental mitochondria as a source of oxidative stress in pre-eclampsia. *Placenta* **19**, 581-6.

- [90] Schumacker, P. T. (2003). Current paradigms in cellular oxygen sensing. Adv Exp Med Biol 543, 57-71.
- [91] Bradford Hill, A. (1965). Environment and Disease: Association or Causation? *Proc R Soc Med* **58,** 295-300.
- [92] Karmon, A. & Pilpel, Y. (2016). Biological causal links on physiological and evolutionary time scales. *Elife* 5.
- [93] Kell, D. B. (2006). Metabolomics, modelling and machine learning in systems biology: towards an understanding of the languages of cells. The 2005 Theodor Bücher lecture. *FEBS J* **273**, 873-894.
- [94] Gradmann, C. (2014). A spirit of scientific rigour: Koch's postulates in twentieth-century medicine. *Microbes Infect* **16**, 885-92.
- [95] Byrd, A. L. & Segre, J. A. (2016). Adapting Koch's postulates. *Science* **351**, 224-226.
- [96] Falkow, S. (1988). Molecular Koch's postulates applied to microbial pathogenicity. *Rev Infect Dis* **10 Suppl 2,** S274-6.
- [97] Falkow, S. (2004). Molecular Koch's postulates applied to bacterial pathogenicity a personal recollection 15 years later. *Nat Rev Microbiol* **2**, 67-72.
- [98] Fredricks, D. N. & Relman, D. A. (1996). Sequence-based identification of microbial pathogens a reconsideration of Koch's postulates. *Clin Micr Rev* **9**, 18-33.
- [99] Lowe, A. M., Yansouni, C. P. & Behr, M. A. (2008). Causality and gastrointestinal infections: Koch, Hill, and Crohn's. *Lancet Infect Dis* **8**, 720-6.
- [100] Seal, J. B., Morowitz, M., Zaborina, O., An, G. & Alverdy, J. C. (2010). The molecular Koch's postulates and surgical infection: a view forward. *Surgery* **147**, 757-65.
- [101] Segre, J. A. (2013). What does it take to satisfy Koch's postulates two centuries later? Microbial genomics and *Propionibacteria acnes*. *J Invest Dermatol* **133**, 2141-2.
- [102] Thagard, P. (1989). Explanatory Coherence. Behav Brain Sci 12, 435-502.
- [103] Thagard, P. & Verbeurgt, K. (1998). Coherence as constraint satisfaction. Cogn Sci 22, 1-24.
- [104] Thagard, P. (1999). How scientists explain disease. Princeton University Press, Princeton, NJ.
- [105] Thagard, P. (2007). Coherence, truth, and the development of scientific knowledge. *Philosophy of Science* **74**, 28-47.
- [106] Thagard, P. (2008). Explanatory Coherence. *Reasoning: Studies of Human Inference and Its Foundations*, 471-513.
- [107] Pennington, K. A., Schlitt, J. M., Jackson, D. L., Schulz, L. C. & Schust, D. J. (2012). Preeclampsia: multiple approaches for a multifactorial disease. *Dis Model Mech* **5**, 9-18.
- [108] Magee, L. A., Pels, A., Helewa, M., Rey, E., von Dadelszen, P. & Canadian Hypertensive Disorders of Pregnancy Working Group. (2014). Diagnosis, evaluation, and management of the hypertensive disorders of pregnancy. *Pregnancy Hypertens* **4**, 105-45.
- [109] Goddard, K. A. G., Tromp, G., Romero, R., Olson, J. M., Lu, Q., Xu, Z., Parimi, N., Nien, J. K., Gomez, R., Behnke, E., Solari, M., Espinoza, J., Santolaya, J., Chaiworapongsa, T., Lenk, G. M., Volkenant, K., Anant, M. K., Salisbury, B. A., Carr, J., Lee, M. S., Vovis, G. F. & Kuivaniemi, H. (2007). Candidategene association study of mothers with pre-eclampsia, and their infants, analyzing 775 SNPs in 190 genes. *Hum Hered* **63**, 1-16.
- [110] Jebbink, J., Wolters, A., Fernando, F., Afink, G., van der Post, J. & Ris-Stalpers, C. (2012). Molecular genetics of preeclampsia and HELLP syndrome a review. *Biochim Biophys Acta* **1822**, 1960-9.
- [111] Fong, F. M., Sahemey, M. K., Hamedi, G., Eyitayo, R., Yates, D., Kuan, V., Thangaratinam, S. & Walton, R. T. (2014). Maternal genotype and severe preeclampsia: a HuGE review. *Am J Epidemiol* **180**, 335-45
- [112] Williamson, R. D., O'Keeffe, G. W. & Kenny, L. C. (2015). Activin signalling and pre-eclampsia: from genetic risk to pre-symptomatic biomarker. *Cytokine* **71**, 360-5.
- [113] Maher, B. (2008). The case of the missing heritability. Nature 456, 18-21.
- [114] Manolio, T. A., Collins, F. S., Cox, N. J., Goldstein, D. B., Hindorff, L. A., Hunter, D. J., McCarthy, M. I., Ramos, E. M., Cardon, L. R., Chakravarti, A., Cho, J. H., Guttmacher, A. E., Kong, A., Kruglyak, L., Mardis, E., Rotimi, C. N., Slatkin, M., Valle, D., Whittemore, A. S., Boehnke, M., Clark, A. G., Eichler, E. E., Gibson, G., Haines, J. L., Mackay, T. F., McCarroll, S. A. & Visscher, P. M. (2009). Finding the missing heritability of complex diseases. *Nature* **461**, 747-53.

- [115] Zuk, O., Hechter, E., Sunyaev, S. R. & Lander, E. S. (2012). The mystery of missing heritability: Genetic interactions create phantom heritability. *Proc Natl Acad Sci U S A*.
- [116] Carreiras, M., Montagnani, S. & Layrisse, Z. (2002). Preeclampsia: A multifactorial disease resulting from the interaction of the feto-maternal HLA genotype and HCMV infection. *Am J Reprod Immunol* **48**, 176-183.
- [117] McCarthy, C. M. & Kenny, L. C. (2016). Immunostimulatory role of mitochondrial DAMPs: alarming for pre-eclampsia? *Am J Reprod Immunol*.
- [118] Kell, D. B. (2009). Iron behaving badly: inappropriate iron chelation as a major contributor to the aetiology of vascular and other progressive inflammatory and degenerative diseases. *BMC Med Genom* **2**, 2
- [119] Kell, D. B. & Pretorius, E. (2014). Serum ferritin is an important disease marker, and is mainly a leakage product from damaged cells. *Metallomics* **6,** 748-773.
- [120] Pretorius, E. & Kell, D. B. (2014). Diagnostic morphology: biophysical indicators for iron-driven inflammatory diseases. *Integrative Biol* **6**, 486-510.
- [121] Kell, D. B. & Pretorius, E. (2015). The simultaneous occurrence of both hypercoagulability and hypofibrinolysis in blood and serum during systemic inflammation, and the roles of iron and fibrin(ogen). *Integr Biol* **7**, 24-52.
- [122] Pretorius, E., Swanepoel, A. C., Oberholzer, H. M., van der Spuy, W. J., Duim, W. & Wessels, P. F. (2011). A descriptive investigation of the ultrastructure of fibrin networks in thrombo-embolic ischemic stroke. *J Thromb Thrombolysis* **31**, 507-13.
- [123] Pretorius, E., Steyn, H., Engelbrecht, M., Swanepoel, A. C. & Oberholzer, H. M. (2011). Differences in fibrin fiber diameters in healthy individuals and thromboembolic ischemic stroke patients. *Blood Coagul Fibrinolysis* 22, 696-700.
- [124] Pretorius, E., Oberholzer, H. M., van der Spuy, W. J., Swanepoel, A. C. & Soma, P. (2011). Qualitative scanning electron microscopy analysis of fibrin networks and platelet abnormalities in diabetes. *Blood Coagul Fibrinol* **22**, 463-7.
- [125] Pretorius, E., du Plooy, J., Soma, P. & Gasparyan, A. Y. (2014). An ultrastructural analysis of platelets, erythrocytes, white blood cells, and fibrin network in systemic lupus erythematosus. *Rheumatol Int* **34**, 1005-1009.
- [126] Pretorius, E., Swanepoel, A. C., Buys, A. V., Vermeulen, N., Duim, W. & Kell, D. B. (2014). Eryptosis as a marker of Parkinson's disease. *Aging* **6**, 788-819.
- [127] Pretorius, E., Bester, J., Vermeulen, N., Alummoottil, S., Soma, P., Buys, A. V. & Kell, D. B. (2015). Poorly controlled type 2 diabetes is accompanied by significant morphological and ultrastructural changes in both erythrocytes and in thrombin-generated fibrin: implications for diagnostics. *Cardiovasc Diabetol* **13**, 30.
- [128] Bester, J., Buys, A. V., Lipinski, B., Kell, D. B. & Pretorius, E. (2013). High ferritin levels have major effects on the morphology of erythrocytes in Alzheimer's disease. *Front Aging Neurosci* **5**, 00088.
- [129] Pretorius, E., Bester, J., Vermeulen, N., Lipinski, B., Gericke, G. S. & Kell, D. B. (2014). Profound morphological changes in the erythrocytes and fibrin networks of patients with hemochromatosis or with hyperferritinemia, and their normalization by iron chelators and other agents. *PLoS One* **9**, e85271.
- [130] Pretorius, E., Olumuyiwa-Akeredolu, O. O., Mbotwe, S. & Bester, J. (2016). Erythrocytes and their role as health indicator: Using structure in a patient-orientated precision medicine approach. *Blood Rev.*
- [131] Todros, T., Vasario, E. & Cardaropoli, S. (2007). Preeclampsia as an infectious disease. *Exp Rev Obs Gynecol* **2**, 735-741.
- [132] Kell, D. B., Potgieter, M. & Pretorius, E. (2015). Individuality, phenotypic differentiation, dormancy and 'persistence' in culturable bacterial systems: commonalities shared by environmental, laboratory, and clinical microbiology. *F1000Research* **4**, 179.
- [133] Kell, D. B. & Pretorius, E. (2015). On the translocation of bacteria and their lipopolysaccharides between blood and peripheral locations in chronic, inflammatory diseases: the central roles of LPS and LPS-induced cell death *Integr Biol* **7**, 1339-1377.
- [134] Potgieter, M., Bester, J., Kell, D. B. & Pretorius, E. (2015). The dormant blood microbiome in chronic, inflammatory diseases. *FEMS Microbiol Rev* **39**, 567-591.

- [135] Pretorius, E., Akeredolu, O.-O., Soma, P. & Kell, D. B. (2016). Major involvement of bacterial components in rheumatoid arthritis and its accompanying oxidative stress, systemic inflammation and hypercoagulability. *Rheumatol Internat*.
- [136] Pretorius, E., Bester, J. & Kell, D. B. (2016). A bacterial component to Alzheimer-type dementia seen via a systems biology approach that links iron dysregulation and inflammagen shedding to disease *J Alzheimers Dis*, in press.
- [137] Proal, A. D., Albert, P. J. & Marshall, T. G. (2013). The human microbiome and autoimmunity. *Curr Opin Rheumatol* **25**, 234-40.
- [138] Mangin, M., Sinha, R. & Fincher, K. (2014). Inflammation and vitamin D: the infection connection. *Inflamm Res* **63**, 803-19.
- [139] Proal, A. D., Albert, P. J. & Marshall, T. G. (2014). Inflammatory disease and the human microbiome. *Discov Med* 17, 257-65.
- [140] Itzhaki, R. F., Lathe, R., Balin, B. J., Ball, M. J., Braak, H., Bearer, E. L., Bullido, M. J., Carter, C., Clerici, M., Cosby, S. L., Del Tredici, K., Field, H., Fulop, T., Grassi, C., Griffin, W. S. T., Haas, J., Hudson, A. P., Kamer, A., Kell, D. B., Licastro, F., Letenneur, L., Lövheim, H., Mancuso, R., Miklossy, J., Otth, C., Palamara, A. T., Perry, G., Preston, C., Pretorius, E., Strandberg, T., Tabet, N., Taylor-Robinson, S. D. & Whittum-Hudson, J. A. (2016). Microbes and Alzheimer's Disease. J Alzheimers Dis 51, 979-984.
- [141] Morath, S., Geyer, A. & Hartung, T. (2001). Structure-function relationship of cytokine induction by lipoteichoic acid from *Staphylococcus aureus*. *J Exp Med* **193**, 393-7.
- [142] Postgate, J. R. (1967). Viability measurements and the survival of microbes under minimum stress. *Adv. Micr. Physiol.* **1,** 1-23.
- [143] Postgate, J. R. (1969). Viable counts and viability. Meth. Microbiol. 1, 611-628.
- [144] Postgate, J. R. (1976). Death in microbes and macrobes. In *In The Survival of Vegetative Microbes* (ed. T. R. G. Gray and J. R. Postgate), pp. 1-19. Cambridge University Press, Cambridge.
- [145] Staley, J. T. & Konopka, A. (1985). Measurement of *in situ* activities of nonphotosynthetic microorganisms in aquatic and terrestrial habitats. *Annu Rev Microbiol* **39**, 321-46.
- [146] Tanaka, T., Kawasaki, K., Daimon, S., Kitagawa, W., Yamamoto, K., Tamaki, H., Tanaka, M., Nakatsu, C. H. & Kamagata, Y. (2014). A hidden pitfall in the preparation of agar media undermines microorganism cultivability. *Appl Environ Microbiol* **80**, 7659-66.
- [147] Nichols, D., Cahoon, N., Trakhtenberg, E. M., Pham, L., Mehta, A., Belanger, A., Kanigan, T., Lewis, K. & Epstein, S. S. (2010). Use of ichip for high-throughput *in situ* cultivation of "uncultivable" microbial species. *Appl Environ Microbiol* **76**, 2445-50.
- [148] Browne, H. P., Forster, S. C., Anonye, B. O., Kumar, N., Neville, B. A., Stares, M. D., Goulding, D. & Lawley, T. D. (2016). Culturing of 'unculturable' human microbiota reveals novel taxa and extensive sporulation. *Nature* **533**, 543-6.
- [149] Kaprelyants, A. S., Gottschal, J. C. & Kell, D. B. (1993). Dormancy in non-sporulating bacteria. *FEMS Microbiol. Rev.* **10,** 271-286.
- [150] Kell, D. B., Kaprelyants, A. S., Weichart, D. H., Harwood, C. L. & Barer, M. R. (1998). Viability and activity in readily culturable bacteria: a review and discussion of the practical issues. *Antonie van Leeuwenhoek* **73**, 169-187.
- [151] Meyer, R. D. (1983). Legionella infections a review of 5 years of research. Rev Infect Dis 5, 258-278.
- [152] Barker, J., Farrell, I. D. & Hutchison, J. G. (1986). Factors affecting growth of *Legionella pneumophila* in liquid media. *J Med Microbiol* **22**, 97-100.
- [153] Molinari, J. (1997). *Legionella* and human disease: Part 1: A path of scientific and community discovery. *Compend Contin Educ Dent* **18,** 556-9.
- [154] Saito, A., Rolfe, R. D., Edelstein, P. H. & Finegold, S. M. (1981). Comparison of liquid growth media for *Legionella pneumophila*. *J Clin Microbiol* **14**, 623-7.
- [155] Maiwald, M., Schuhmacher, F., Ditton, H. J. & vonHerbay, A. (1998). Environmental occurrence of the Whipple's disease bacterium (*Tropheryma whippelii*). *Appl. Env. Microbiol.* **64,** 760-762.
- [156] Maiwald, M. & Relman, D. A. (2001). Whipple's disease and Tropheryma whippelii: Secrets slowly revealed. *Clin Infect Dis* **32**, 457-463.

- [157] Omsland, A., Cockrell, D. C., Howe, D., Fischer, E. R., Virtaneva, K., Sturdevant, D. E., Porcella, S. F. & Heinzen, R. A. (2009). Host cell-free growth of the Q fever bacterium *Coxiella burnetii*. *Proc Natl Acad Sci* **106**, 4430-4434.
- [158] Omsland, A. (2012). Axenic growth of Coxiella burnetii. Adv Exp Med Biol 984, 215-29.
- [159] Marshall, B. J. (2001). One Hundred Years of Discovery and Rediscovery of *Helicobacter pylori* and Its Association with Peptic Ulcer Disease. In *Helicobacter pylori: Physiology and Genetics* (ed. H. L. T. Mobley, G. L. Mendz and S. L. Hazell), pp. 19-24. ASM Press, Washington (DC).
- [160] Marshall, B. (2006). Helicobacter connections. ChemMedChem 1, 783-802.
- [161] Marshall, B. J. & Warren, J. R. (1984). Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration. *Lancet* **1**, 1311-1315.
- [162] Marshall, B. J., Armstrong, J. A., McGechie, D. B. & Glancy, R. J. (1985). Attempt to fulfil Koch's postulates for pyloric *Campylobacter*. *Med J Aust* **142**, 436-9.
- [163] Kaprelyants, A. S. & Kell, D. B. (1993). Dormancy in stationary-phase cultures of *Micrococcus luteus*: flow cytometric analysis of starvation and resuscitation. *Appl. Env. Microbiol.* **59,** 3187-3196.
- [164] Kaprelyants, A. S., Mukamolova, G. V. & Kell, D. B. (1994). Estimation of dormant *Micrococcus luteus* cells by penicillin lysis and by resuscitation in cell-free spent medium at high dilution. *FEMS Microbiol Lett* **115**, 347-352.
- [165] Alnimr, A. M. (2015). Dormancy models for *Mycobacterium tuberculosis*: A minireview. *Braz J Microbiol* **46**, 641-7.
- [166] Gengenbacher, M. & Kaufmann, S. H. E. (2012). *Mycobacterium tuberculosis*: success through dormancy. *FEMS Microbiol Rev* **36**, 514-532.
- [167] Kana, B. D., Gordhan, B. G., Downing, K. J., Sung, N., Vostroktunova, G., Machowski, E. E., Tsenova, L., Young, M., Kaprelyants, A., Kaplan, G. & Mizrahi, V. (2008). The resuscitation-promoting factors of *Mycobacterium tuberculosis* are required for virulence and resuscitation from dormancy but are collectively dispensable for growth *in vitro*. *Mol Microbiol* 67, 672-84.
- [168] Shleeva, M., Kondratieva, T., Rubakova, E., Vostroknutova, G., Kaprelyants, A. & Apt, A. (2015). Reactivation of dormant "non-culturable" *Mycobacterium tuberculosis* developed *in vitro* after injection in mice: Both the dormancy depth and host genetics influence the outcome. *Microb Pathog* **78**, 63-6.
- [169] Wayne, L. G. (1994). Dormancy of *Mycobacterium tuberculosis and* latency of disease. *Eur. J. Clin. Microbiol. Infect. Dis.* **13**, 908-914.
- [170] Mukamolova, G. V., Kaprelyants, A. S., Young, D. I., Young, M. & Kell, D. B. (1998). A bacterial cytokine. *Proc. Natl. Acad. Sci.* **95**, 8916-8921.
- [171] Mukamolova, G. V., Turapov, O. A., Young, D. I., Kaprelyants, A. S., Kell, D. B. & Young, M. (2002). A family of autocrine growth factors in *Mycobacterium tuberculosis*. *Mol Microbiol* **46**, 623-35.
- [172] Mukamolova, G. V., Turapov, O. A., Kazarian, K., Telkov, M., Kaprelyants, A. S., Kell, D. B. & Young, M. (2002). The rpf gene of Micrococcus luteus encodes an essential secreted growth factor. Mol Microbiol 46, 611-21.
- [173] Mukamolova, G. V., Murzin, A. G., Salina, E. G., Demina, G. R., Kell, D. B., Kaprelyants, A. S. & Young, M. (2006). Muralytic activity of *Micrococcus luteus* Rpf and its relationship to physiological activity in promoting bacterial growth and resuscitation. *Mol Microbiol* **59**, 84-98.
- [174] Mukamolova, G. V., Turapov, O., Malkin, J., Woltmann, G. & Barer, M. R. (2010). Resuscitation-promoting factors reveal an occult population of tubercle bacilli in sputum. *Am J Respir Crit Care Med* **181**, 174-80.
- [175] Yeremeev, V. V., Kondratieva, T. K., Rubakova, E. I., Petrovskaya, S. N., Kazarian, K. A., Telkov, M. V., Biketov, S. F., Kaprelyants, A. S. & Apt, A. S. (2003). Proteins of the Rpf family: immune cell reactivity and vaccination efficacy against tuberculosis in mice. *Infect Immun* 71, 4789-94.
- [176] Zvi, A., Ariel, N., Fulkerson, J., Sadoff, J. C. & Shafferman, A. (2008). Whole genome identification of *Mycobacterium tuberculosis* vaccine candidates by comprehensive data mining and bioinformatic analyses. *BMC Med Genomics* **1,** 18.
- [177] Dye, C., Scheele, S., Dolin, P., Pathania, V. & Raviglione, R. C. (1999). Global burden of tuberculosis Estimated incidence, prevalence, and mortality by country. *JAMA* **282**, 677-686.
- [178] Flynn, J. L. & Chan, J. (2001). Tuberculosis: Latency and reactivation. *Infect. Immun.* **69,** 4195-4201.

- [179] Wayne, L. G. & Sohaskey, C. D. (2001). Nonreplicating persistence of *Mycobacterium tuberculosis*. *Annu. Rev. Microbiol.* **55,** 139-63.
- [180] Gomez, J. E. & McKinney, J. D. (2004). *M. tuberculosis* persistence, latency, and drug tolerance. *Tuberculosis (Edinb)* **84,** 29-44.
- [181] Amieva, M. R. & El-Omar, E. M. (2008). Host-bacterial interactions in *Helicobacter pylori* infection. *Gastroenterology* **134**, 306-23.
- [182] Cover, T. L. & Blaser, M. J. (2009). *Helicobacter pylori* in health and disease. *Gastroenterology* **136**, 1863-73.
- [183] Domingue, G. J. & Woody, H. B. (1997). Bacterial persistence and expression of disease. *Clin Microbiol Rev* **10**, 320-344.
- [184] Young, D., Stark, J. & Kirschner, D. (2008). Systems biology of persistent infection: tuberculosis as a case study. *Nat Rev Microbiol* **6**, 520-8.
- [185] Lewis, K. (2010). Persister cells. Annu Rev Microbiol 64, 357-72.
- [186] Amato, S. M., Fazen, C. H., Henry, T. C., Mok, W. W. K., Orman, M. A., Sandvik, E. L., Volzing, K. G. & Brynildsen, M. P. (2014). The role of metabolism in bacterial persistence. *Front Microbiol* **5**.
- [187] Conlon, B. P. (2014). *Staphylococcus aureus* chronic and relapsing infections: Evidence of a role for persister cells: An investigation of persister cells, their formation and their role in *S. aureus* disease. *Bioessays* **36**, 991-6.
- [188] Kester, J. C. & Fortune, S. M. (2014). Persisters and beyond: mechanisms of phenotypic drug resistance and drug tolerance in bacteria. *Crit Rev Biochem Mol Biol* **49**, 91-101.
- [189] Levin, B. R., Concepción-Acevedo, J. & Udekwu, K. I. (2014). Persistence: a copacetic and parsimonious hypothesis for the existence of non-inherited resistance to antibiotics. *Curr Opin Microbiol* **21**, 18-21.
- [190] Maisonneuve, E. & Gerdes, K. (2014). Molecular mechanisms underlying bacterial persisters. *Cell* **157**, 539-48.
- [191] Rank, R. G. & Yeruva, L. (2014). Hidden in plain sight: chlamydial gastrointestinal infection and its relevance to persistence in human genital infection. *Infect Immun* **82**, 1362-71.
- [192] Holden, D. W. (2015). Persisters unmasked. Science 347, 30-2.
- [193] Orman, M. A., Mok, W. W. K. & Brynildsen, M. P. (2015). Aminoglycoside-enabled elucidation of bacterial persister metabolism. *Curr Protoc Microbiol* **36**, 17 9 1-17 9 14.
- [194] Stepanyan, K., Wenseleers, T., Duéñez-Guzmán, E. A., Muratori, F., Van den Bergh, B., Verstraeten, N., De Meester, L., Verstrepen, K. J., Fauvart, M. & Michiels, J. (2015). Fitness trade-offs explain low levels of persister cells in the opportunistic pathogen *Pseudomonas aeruginosa*. *Mol Ecol* **24**, 1572-83.
- [195] Pu, Y., Zhao, Z., Li, Y., Zou, J., Ma, Q., Zhao, Y., Ke, Y., Zhu, Y., Chen, H., Baker, M. A., Ge, H., Sun, Y., Xie, X. S. & Bai, F. (2016). Enhanced efflux activity facilitates drug tolerance in dormant bacterial cells. *Mol Cell* 62, 284-94.
- [196] Gerdes, K. & Semsey, S. (2016). Pumping persisters. *Nature* **534,** 41-2.
- [197] Dehio, C., Berry, C. & Bartenschlager, R. (2012). Persistent intracellular pathogens. *FEMS Microbiol Rev* **36**, 513.
- [198] Prideaux, B., Via, L. E., Zimmerman, M. D., Eum, S., Sarathy, J., O'Brien, P., Chen, C., Kaya, F., Weiner, D. M., Chen, P. Y., Song, T., Lee, M., Shim, T. S., Cho, J. S., Kim, W., Cho, S. N., Olivier, K. N., Barry, C. E., 3rd & Dartois, V. (2015). The association between sterilizing activity and drug distribution into tuberculosis lesions. *Nat Med* 21, 1223-7.
- [199] Dobson, P. D. & Kell, D. B. (2008). Carrier-mediated cellular uptake of pharmaceutical drugs: an exception or the rule? *Nat Rev Drug Disc* **7**, 205-220.
- [200] Kell, D. B., Dobson, P. D. & Oliver, S. G. (2011). Pharmaceutical drug transport: the issues and the implications that it is essentially carrier-mediated only. *Drug Disc Today* **16**, 704-714.
- [201] Kell, D. B., Dobson, P. D., Bilsland, E. & Oliver, S. G. (2013). The promiscuous binding of pharmaceutical drugs and their transporter-mediated uptake into cells: what we (need to) know and how we can do so. *Drug Disc Today* **18**, 218-239.

- [202] Kell, D. B. & Oliver, S. G. (2014). How drugs get into cells: tested and testable predictions to help discriminate between transporter-mediated uptake and lipoidal bilayer diffusion. *Front Pharmacol* **5,** 231.
- [203] Thwaites, G. E. & Gant, V. (2011). Are bloodstream leukocytes Trojan Horses for the metastasis of *Staphylococcus aureus? Nat Rev Microbiol* **9,** 215-22.
- [204] Seubert, A., Schulein, R. & Dehio, C. (2002). Bacterial persistence within erythrocytes: a unique pathogenic strategy of *Bartonella* spp. *Int J Med Microbiol* **291**, 555-60.
- [205] Blango, M. G. & Mulvey, M. A. (2010). Persistence of uropathogenic *Escherichia coli* in the face of multiple antibiotics. *Antimicrob Agents Chemother* **54,** 1855-63.
- [206] Mysorekar, I. U. & Hultgren, S. J. (2006). Mechanisms of uropathogenic *Escherichia coli* persistence and eradication from the urinary tract. *Proc Natl Acad Sci U S A* **103**, 14170-5.
- [207] Goneau, L. W., Hannan, T. J., MacPhee, R. A., Schwartz, D. J., Macklaim, J. M., Gloor, G. B., Razvi, H., Reid, G., Hultgren, S. J. & Burton, J. P. (2015). Subinhibitory antibiotic therapy alters recurrent urinary tract infection pathogenesis through modulation of bacterial virulence and host immunity. *mBio* **6**, e00356-15.
- [208] Chen, S. L., Wu, M., Henderson, J. P., Hooton, T. M., Hibbing, M. E., Hultgren, S. J. & Gordon, J. I. (2013). Genomic diversity and fitness of *E. coli* strains recovered from the intestinal and urinary tracts of women with recurrent urinary tract infection. *Sci Transl Med* **5**, 184ra60.
- [209] Sullivan, W. J., Jr. & Jeffers, V. (2012). Mechanisms of *Toxoplasma gondii* persistence and latency. *FEMS Microbiol Rev* **36**, 717-33.
- [210] Posey, J. E. & Gherardini, F. C. (2000). Lack of a role for iron in the Lyme disease pathogen. *Science* **288**, 1651-3.
- [211] Aguirre, J. D., Clark, H. M., McIlvin, M., Vazquez, C., Palmere, S. L., Grab, D. J., Seshu, J., Hart, P. J., Saito, M. & Culotta, V. C. (2013). A manganese-rich environment supports superoxide dismutase activity in a Lyme disease pathogen, *Borrelia burgdorferi*. *J Biol Chem* **288**, 8468-78.
- [212] Weinberg, E. D. (1975). Nutritional immunity. Host's attempt to withold iron from microbial invaders. *JAMA* 231, 39-41.
- [213] Weinberg, E. D. (1984). Iron withholding: a defense against infection and neoplasia. *Physiol Rev* **64**, 65-102
- [214] Marx, J. J. M. (2002). Iron and infection: competition between host and microbes for a precious element. *Best Pract Res Clin Haematol* **15**, 411-26.
- [215] Fischbach, M. A., Lin, H. N., Liu, D. R. & Walsh, C. T. (2006). How pathogenic bacteria evade mammalian sabotage in the battle for iron. *Nat Chem Biol* **2**, 132-138.
- [216] Weinberg, E. D. & Miklossy, J. (2008). Iron withholding: a defense against disease. *J Alzheimers Dis* **13**, 451-63.
- [217] Weinberg, E. D. (2009). Iron availability and infection. Biochim Biophys Acta 1790, 600-5.
- [218] Reid, D. W., Anderson, G. J. & Lamont, I. L. (2009). Role of lung iron in determining the bacterial and host struggle in cystic fibrosis. *Am J Physiol Lung Cell Mol Physiol* **297**, L795-802.
- [219] Chu, B. C., Garcia-Herrero, A., Johanson, T. H., Krewulak, K. D., Lau, C. K., Peacock, R. S., Slavinskaya, Z. & Vogel, H. J. (2010). Siderophore uptake in bacteria and the battle for iron with the host; a bird's eye view. *Biometals* **23**, 601-11.
- [220] Nairz, M., Schroll, A., Sonnweber, T. & Weiss, G. (2010). The struggle for iron a metal at the host-pathogen interface. *Cell Microbiol* **12**, 1691-702.
- [221] Skaar, E. P. (2010). The battle for iron between bacterial pathogens and their vertebrate hosts. *PLoS Pathog* **6**, e1000949.
- [222] Hood, M. I. & Skaar, E. P. (2012). Nutritional immunity: transition metals at the pathogen-host interface. *Nat Rev Microbiol* **10**, 525-37.
- [223] Cassat, J. E. & Skaar, E. P. (2013). Iron in infection and immunity. Cell Host Microbe 13, 509-19.
- [224] Deriu, E., Liu, J. Z., Pezeshki, M., Edwards, R. A., Ochoa, R. J., Contreras, H., Libby, S. J., Fang, F. C. & Raffatellu, M. (2013). Probiotic bacteria reduce *Salmonella typhimurium* intestinal colonization by competing for iron. *Cell Host Microbe* **14**, 26-37.
- [225] Leal, S. M., Jr., Roy, S., Vareechon, C., Carrion, S., Clark, H., Lopez-Berges, M. S., Di Pietro, A., Schrettl, M., Beckmann, N., Redl, B., Haas, H. & Pearlman, E. (2013). Targeting iron acquisition blocks

- infection with the fungal pathogens *Aspergillus fumigatus* and *Fusarium oxysporum*. *PLoS Pathog* **9**, e1003436.
- [226] Silva-Gomes, S., Vale-Costa, S., Appelberg, R. & Gomes, M. S. (2013). Iron in intracellular infection: to provide or to deprive? *Front Cell Infect Microbiol* **3**, 96.
- [227] Armitage, A. E. & Drakesmith, H. (2014). The battle for iron. Science 346, 1299-300.
- [228] Becker, K. W. & Skaar, E. P. (2014). Metal limitation and toxicity at the interface between host and pathogen. *FEMS Microbiol Rev* **38**, 1235-49.
- [229] Diaz-Ochoa, V. E., Jellbauer, S., Klaus, S. & Raffatellu, M. (2014). Transition metal ions at the crossroads of mucosal immunity and microbial pathogenesis. *Front Cell Infect Microbiol* **4,** 2.
- [230] Potrykus, J., Ballou, E. R., Childers, D. S. & Brown, A. J. P. (2014). Conflicting interests in the pathogen-host tug of war: fungal micronutrient scavenging versus mammalian nutritional immunity. *PLoS Pathog* **10**, e1003910.
- [231] Barber, M. F. & Elde, N. C. (2015). Buried Treasure: Evolutionary Perspectives on Microbial Iron Piracy. *Trends Genet* **31**, 627-36.
- [232] Nairz, M., Ferring-Appel, D., Casarrubea, D., Sonnweber, T., Viatte, L., Schroll, A., Haschka, D., Fang, F. C., Hentze, M. W., Weiss, G. & Galy, B. (2015). Iron Regulatory Proteins Mediate Host Resistance to *Salmonella* Infection. *Cell Host Microbe* **18**, 254-61.
- [233] Raymond, K. N., Allred, B. E. & Sia, A. K. (2015). Coordination Chemistry of Microbial Iron Transport. *Acc Chem Res* **48**, 2496-505.
- [234] Subashchandrabose, S. & Mobley, H. L. T. (2015). Back to the metal age: battle for metals at the host-pathogen interface during urinary tract infection. *Metallomics* **7**, 935-942.
- [235] Wessling-Resnick, M. (2015). Nramp1 and Other Transporters Involved in Metal Withholding during Infection. *J Biol Chem* **290**, 18984-90.
- [236] Xia, Y., Farah, N., Maxan, A., Zhou, J. & Lehmann, C. (2016). Therapeutic iron restriction in sepsis. *Med Hypotheses* **89,** 37-9.
- [237] Entman, S. S., Richardson, L. D. & Killam, A. P. (1982). Elevated serum ferritin in the altered ferrokinetics of toxemia of pregnancy. *AmJ Obs Gynecol* **144**, 418-422.
- [238] Samuels, P., Main, E. K., Mennuti, M. T. & Gabbe, S. G. (1987). The origin of increased serum iron in pregnancy-induced hypertension. *Am J Obstet Gynecol* **157**, 721-5.
- [239] Raman, L., Pawashe, A. B. & Yasodhara, P. (1992). Hyperferritinemia in pregnancy induced hypertension and eclampsia. *J Postgrad Med* **38**, 65-7.
- [240] Hubel, C. A., Kozlov, A. V., Kagan, V. E., Evans, R. W., Davidge, S. T., McLaughlin, M. K. & Roberts, J. M. (1996). Decreased transferrin and increased transferrin saturation in sera of women with preeclampsia: implications for oxidative stress. *Am J Obstet Gynecol* **175**, 692-700.
- [241] Lao, T. T., Tam, K. F. & Chan, L. Y. (2000). Third trimester iron status and pregnancy outcome in non-anaemic women; pregnancy unfavourably affected by maternal iron excess. *Hum Reprod* **15**, 1843-8.
- [242] Rayman, M. P., Barlis, J., Evans, R. W., Redman, C. W. & King, L. J. (2002). Abnormal iron parameters in the pregnancy syndrome preeclampsia. *Am J Obstet Gynecol* **187**, 412-8.
- [243] Serdar, Z., Gür, E. & Develioğlu, O. (2006). Serum iron and copper status and oxidative stress in severe and mild preeclampsia. *Cell Biochem Funct* **24**, 209-15.
- [244] Smith, T. G. & Robbins, P. A. (2007). Iron, pre-eclampsia and hypoxia-inducible factor. *BJOG* **114**, 1581-2.
- [245] Bhatla, N., Kaul, N., Lal, N., Kriplani, A., Agarwal, N., Saxena, R. & Gupta, S. K. (2009). Comparison of effect of daily versus weekly iron supplementation during pregnancy on lipid peroxidation. *J Obstet Gynaecol Res* **35**, 438-45.
- [246] Siddiqui, I. A., Jaleel, A., Kadri, H. M., Saeed, W. A. & Tamimi, W. (2011). Iron status parameters in preeclamptic women. *Arch Gynecol Obstet* **284**, 587-91.
- [247] Fatima, N., Islam, F., Noor, L., Das, S. R., Zeba, D. & Zesmin, F. (2013). Serum Ferritin in Preeclampsia and Eclampsia: A Case Control Study. *Faridpur Med Coll J* **8**, 18-21.
- [248] Kandi, S., Sudhakar, T., Ramadevi, C., Venugopal, B., Rajkumar, Rafi, M. & Ramana, K. V. (2014). Pre Eclampsia and Iron Status: A Review. *Am J Med Biol Res* **2**, 121-123.

- [249] Negi, R., Pande, D., Karki, K., Kumar, A., Khanna, R. S. & Khanna, H. D. (2014). Association of oxidative DNA damage, protein oxidation and antioxidant function with oxidative stress induced cellular injury in pre-eclamptic/eclamptic mothers during fetal circulation. *Chem Biol Interact* **208**, 77-83.
- [250] Bester, J., Soma, P., Kell, D. B. & Pretorius, E. (2015). Viscoelastic and ultrastructural characteristics of whole blood and plasma in Alzheimer-type dementia, and the possible role of bacterial lipopolysaccharides (LPS). *Oncotarget Gerontology* **6**, 35284-35303.
- [251] Pretorius, E., Mbotwe, S., Bester, J., Robinson, C. & Kell, D. B. (2016). Acute induction of anomalous blood clotting by highly substoichiometric levels of bacterial lipopolysaccharide (LPS). *bioRxiv*, 2016-053538v1.
- [252] Pretorius, E., Mbotwe, S., Bester, J., Robinson, C. & Kell, D. B. (2016). Acute induction of anomalous blood clotting by highly substoichiometric levels of bacterial lipopolysaccharide. *J R Soc Interface*, submitted.
- [253] Lin, I. H., Miller, D. S., Bertics, P. J., Murphy, C. J., de Pablo, J. J. & Abbott, N. L. (2011). Endotoxin-induced structural transformations in liquid crystalline droplets. *Science* **332**, 1297-300.
- [254] Miller, D. S. & Abbott, N. L. (2013). Influence of droplet size, pH and ionic strength on endotoxin-triggered ordering transitions in liquid crystalline droplets. *Soft Matter* **9**, 374-382.
- [255] Harris, C. M. & Kell, D. B. (1985). The estimation of microbial biomass. *Biosensors* 1, 17-84.
- [256] Kaprelyants, A. S. & Kell, D. B. (1996). Do bacteria need to communicate with each other for growth? *Trends Microbiol.* **4,** 237-242.
- [257] Domingue, G. J. (1995). Electron dense cytoplasmic particles and chronic infection a bacterial pleomorphy hypothesis. *Endocytobiosis Cell Res.* **11,** 19-40.
- [258] Domingue, G. J., Sr. (1998). Cryptic bacterial infection in chronic prostatitis: diagnostic and therapeutic implications. *Curr Opin Urol* **8**, 45-9.
- [259] Mattman, L. (2001). Cell wall deficient forms: stealth pathogens, 3rd Ed. CRC Press, Boca Raton.
- [260] Domingue, G. J. (2010). Demystifying pleomorphic forms in persistence and expression of disease: Are they bacteria, and is peptidoglycan the solution? *Discov Med* **10**, 234-46.
- [261] Davey, H. M. & Kell, D. B. (1996). Flow cytometry and cell sorting of heterogeneous microbial populations: the importance of single-cell analysis. *Microbiol. Rev.* **60**, 641-696.
- [262] Kaprelyants, A. S. & Kell, D. B. (1992). Rapid assessment of bacterial viability and vitality using rhodamine 123 and flow cytometry. *J. Appl. Bacteriol.* **72**, 410-422.
- [263] Gant, V. A., Warnes, G., Phillips, I. & Savidge, G. F. (1993). The application of flow cytometry to the study of bacterial responses to antibiotics. *J. Med. Microbiol.* **39,** 147-154.
- [264] Mason, D. J., Shanmuganathan, S., Mortimer, F. C. & Gant, V. A. (1998). A fluorescent Gram stain for flow cytometry and epifluorescence microscopy. *Appl Environ Microbiol* **64**, 2681-5.
- [265] Nebe-von-Caron, G., Stephens, P. J., Hewitt, C. J., Powell, J. R. & Badley, R. A. (2000). Analysis of bacterial function by multi-colour fluorescence flow cytometry and single cell sorting. *J Microbiol Methods* **42**, 97-114.
- [266] Shapiro, H. M. & Nebe-von-Caron, G. (2004). Multiparameter flow cytometry of bacteria. *Methods Mol Biol* **263**, 33-44.
- [267] Müller, S. & Nebe-von-Caron, G. (2010). Functional single-cell analyses: flow cytometry and cell sorting of microbial populations and communities. *FEMS Microbiol Rev* **34**, 554-87.
- [268] Davey, H. M. & Hexley, P. (2011). Red but not dead? Membranes of stressed *Saccharomyces cerevisiae* are permeable to propidium iodide. *Environ Microbiol* **13**, 163-71.
- [269] Davey, H. M. (2011). Life, death, and in-between: meanings and methods in microbiology. *Appl Environ Microbiol* **77**, 5571-6.
- [270] Kell, D. B., Markx, G. H., Davey, C. L. & Todd, R. W. (1990). Real-time monitoring of cellular biomass: Methods and applications. *Trends Anal. Chem.* **9**, 190-194.
- [271] Kell, D. B. & Sonnleitner, B. (1995). GMP Good Modelling Practice: an essential component of good manufacturing practice. *Trends Biotechnol.* **13**, 481-492.
- [272] Firstenberg-Eden, R. & Eden, G. (1984). *Impedance Microbiology*. Research Studies Press, Letchworth.
- [273] Harris, C. M., Todd, R. W., Bungard, S. J., Lovitt, R. W., Morris, J. G. & Kell, D. B. (1987). The dielectric permittivity of microbial suspensions at radio frequencies: a novel method for the estimation of microbial biomass. *Enzyme Microbial Technol.* **9**, 181-186.

- [274] Kell, D. B. & Davey, C. L. (1990). Conductimetric and impedimetric devices. In *Biosensors: a practical approach* (ed. A. E. G. Cass), pp. 125-154. IRL Press, Oxford.
- [275] Woese, C. R. & Fox, G. E. (1977). Phylogenetic structure of the prokaryotic domain: the primary kingdoms. *Proc Natl Acad Sci U S A* **74**, 5088-90.
- [276] Petti, C. A. (2007). Detection and identification of microorganisms by gene amplification and sequencing. *Clinical Infectious Diseases* **44**, 1108-1114.
- [277] Klouche, M. & Schröder, U. (2008). Rapid methods for diagnosis of bloodstream infections. *Clin Chem Lab Med* **46**, 888-908.
- [278] Woo, P. C. Y., Lau, S. K. P., Teng, J. L. L., Tse, H. & Yuen, K. Y. (2008). Then and now: use of 16S rDNA gene sequencing for bacterial identification and discovery of novel bacteria in clinical microbiology laboratories. *Clin Microbiol Infect* **14**, 908-934.
- [279] Cherkaoui, A., Emonet, S., Ceroni, D., Candolfi, B., Hibbs, J., Francois, P. & Schrenzel, J. (2009). Development and validation of a modified broad-range 16S rDNA PCR for diagnostic purposes in clinical microbiology. *J Microbiol Methods* **79**, 227-31.
- [280] Caporaso, J. G., Lauber, C. L., Walters, W. A., Berg-Lyons, D., Lozupone, C. A., Turnbaugh, P. J., Fierer, N. & Knight, R. (2011). Global patterns of 16S rRNA diversity at a depth of millions of sequences per sample. *Proc Natl Acad Sci U S A* 108 Suppl 1, 4516-22.
- [281] Kramski, M., Gaeguta, A. J., Lichtfuss, G. F., Rajasuriar, R., Crowe, S. M., French, M. A., Lewin, S. R., Center, R. J. & Purcell, D. F. (2011). Novel sensitive real-time PCR for quantification of bacterial 16S rRNA genes in plasma of HIV-infected patients as a marker for microbial translocation. *J Clin Microbiol* 49, 3691-3.
- [282] Kämpfer, P. (2012). Systematics of prokaryotes: the state of the art. *Antonie Van Leeuwenhoek* **101**, 3-11.
- [283] Ohlin, A., Bäckman, A., Ewald, U., Schollin, J. & Björkqvist, M. (2012). Diagnosis of neonatal sepsis by broad-range 16S real-time polymerase chain reaction. *Neonatology* **101**, 241-6.
- [284] Langille, M. G. I., Zaneveld, J., Caporaso, J. G., McDonald, D., Knights, D., Reyes, J. A., Clemente, J. C., Burkepile, D. E., Vega Thurber, R. L., Knight, R., Beiko, R. G. & Huttenhower, C. (2013). Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat Biotechnol* **31**, 814-21.
- [285] Mizrahi-Man, O., Davenport, E. R. & Gilad, Y. (2013). Taxonomic classification of bacterial 16S rRNA genes using short sequencing reads: evaluation of effective study designs. *PLoS One* **8**, e53608.
- [286] Valencia-Shelton, F. & Loeffelholz, M. (2014). Nonculture techniques for the detection of bacteremia and fungemia. *Future Microbiol* **9**, 543-59.
- [287] Yarza, P., Yilmaz, P., Pruesse, E., Glöckner, F. O., Ludwig, W., Schleifer, K. H., Whitman, W. B., Euzéby, J., Amann, R. & Rosselló-Móra, R. (2014). Uniting the classification of cultured and uncultured bacteria and archaea using 16S rRNA gene sequences. *Nat Rev Microbiol* **12**, 635-45.
- [288] Zumla, A., Al-Tawfiq, J. A., Enne, V. I., Kidd, M., Drosten, C., Breuer, J., Muller, M. A., Hui, D., Maeurer, M., Bates, M., Mwaba, P., Al-Hakeem, R., Gray, G., Gautret, P., Al-Rabeeah, A. A., Memish, Z. A. & Gant, V. (2014). Rapid point of care diagnostic tests for viral and bacterial respiratory tract infections--needs, advances, and future prospects. *Lancet Infect Dis* 14, 1123-35.
- [289] D'Amore, R., Ijaz, U. Z., Schirmer, M., Kenny, J. G., Gregory, R., Darby, A. C., Shakya, M., Podar, M., Quince, C. & Hall, N. (2016). A comprehensive benchmarking study of protocols and sequencing platforms for 16S rRNA community profiling. *BMC Genomics* **17**, 55.
- [290] Versalovic, J., Carroll, K. C., Funke, G., Jorgensen, J. H., Landry, M. L. & Warnock, D. W. (2011). *Manual of Clinical Microbiology, 10th Edition*. American Society of Microbiology, Washington.
- [291] Nikkari, S., McLaughlin, I. J., Bi, W., Dodge, D. E. & Relman, D. A. (2001). Does blood of healthy subjects contain bacterial ribosomal DNA? *J Clin Microbiol* **39**, 1956-9.
- [292] McLaughlin, R. W., Vali, H., Lau, P. C., Palfree, R. G. E., De Ciccio, A., Sirois, M., Ahmad, D., Villemur, R., Desrosiers, M. & Chan, E. C. S. (2002). Are there naturally occurring pleomorphic bacteria in the blood of healthy humans? *J Clin Microbiol* **40**, 4771-5.
- [293] Moriyama, K., Ando, C., Tashiro, K., Kuhara, S., Okamura, S., Nakano, S., Takagi, Y., Miki, T., Nakashima, Y. & Hirakawa, H. (2008). Polymerase chain reaction detection of bacterial 16S rRNA gene in human blood. *Microbiol Immunol* 52, 375-82.

- [294] Amar, J., Serino, M., Lange, C., Chabo, C., Iacovoni, J., Mondot, S., Lepage, P., Klopp, C., Mariette, J., Bouchez, O., Perez, L., Courtney, M., Marre, M., Klopp, P., Lantieri, O., Doré, J., Charles, M. A., Balkau, B., Burcelin, R. & Grp, D. S. (2011). Involvement of tissue bacteria in the onset of diabetes in humans: evidence for a concept. *Diabetologia* **54**, 3055-3061.
- [295] Gaibani, P., Mariconti, M., Bua, G., Bonora, S., Sassera, D., Landini, M. P., Mulatto, P., Novati, S., Bandi, C. & Sambri, V. (2013). Development of a broad-range 23S rDNA real-time PCR assay for the detection and quantification of pathogenic bacteria in human whole blood and plasma specimens. *Biomed Res Int* **2013**, 264651.
- [296] Dinakaran, V., Rathinavel, A., Pushpanathan, M., Sivakumar, R., Gunasekaran, P. & Rajendhran, J. (2014). Elevated levels of circulating DNA in cardiovascular disease patients: metagenomic profiling of microbiome in the circulation. *PLoS One* **9**, e105221.
- [297] Sato, J., Kanazawa, A., Ikeda, F., Yoshihara, T., Goto, H., Abe, H., Komiya, K., Kawaguchi, M., Shimizu, T., Ogihara, T., Tamura, Y., Sakurai, Y., Yamamoto, R., Mita, T., Fujitani, Y., Fukuda, H., Nomoto, K., Takahashi, T., Asahara, T., Hirose, T., Nagata, S., Yamashiro, Y. & Watada, H. (2014). Gut dysbiosis and detection of "live gut bacteria" in blood of Japanese patients with type 2 diabetes. *Diabetes Care* 37, 2343-50.
- [298] Damgaard, C., Magnussen, K., Enevold, C., Nilsson, M., Tolker-Nielsen, T., Holmstrup, P. & Nielsen, C. H. (2015). Viable bacteria associated with red blood cells and plasma in freshly drawn blood donations. *PLoS One* 10, e0120826.
- [299] Gyarmati, P., Kjellander, C., Aust, C., Kalin, M., Öhrmalm, L. & Giske, C. G. (2015). Bacterial Landscape of Bloodstream Infections in Neutropenic Patients via High Throughput Sequencing. *Plos One* **10**.
- [300] Gyarmati, P., Kjellander, C., Aust, C., Song, Y., Öhrmalm, L. & Giske, C. G. (2016). Metagenomic analysis of bloodstream infections in patients with acute leukemia and therapy-induced neutropenia. *Scientific Reports* 6.
- [301] Païssé, S., Valle, C., Servant, F., Courtney, M., Burcelin, R., Amar, J. & Lelouvier, B. (2016). Comprehensive description of blood microbiome from healthy donors assessed by 16S targeted metagenomic sequencing. *Transfusion*.
- [302] Domingue, G. J. & Schlegel, J. U. (1977). Novel bacterial structures in human blood: cultural isolation. *Infect Immun* **15**, 621-7.
- [303] Belstrøm, D., Holmstrup, P., Damgaard, C., Borch, T. S., Skjødt, M. O., Bendtzen, K. & Nielsen, C. H. (2011). The atherogenic bacterium *Porphyromonas gingivalis* evades circulating phagocytes by adhering to erythrocytes. *Infect Immun* **79**, 1559-65.
- [304] Billings, F. (1915). Focal infection. Appleton, New York.
- [305] Price, W. A. (1923). Dental infections oral and systemic, being a contribution to the pathology of dental infections, focal infections and the degenerative diseases, Parts I and II. Penton Press, Cleveland.
- [306] Miklossy, J., Kis, A., Radenovic, A., Miller, L., Forro, L., Martins, R., Reiss, K., Darbinian, N., Darekar, P., Mihaly, L. & Khalili, K. (2006). Beta-amyloid deposition and Alzheimer's type changes induced by Borrelia spirochetes. *Neurobiol Aging* **27**, 228-36.
- [307] Miklossy, J. (2008). Chronic inflammation and amyloidogenesis in Alzheimer's disease -- role of Spirochetes. *J Alzheimers Dis* **13,** 381-91.
- [308] Woolard, M. D. & Frelinger, J. A. (2008). Outsmarting the host: bacteria modulating the immune response. *Immunol Res* **41**, 188-202.
- [309] Nicolson, G. L. & Haier, J. (2009). Role of chronic bacterial and viral infections in neurodegenerative, neurobehavioural, psychiatric, autoimmune and fatiguing illnesses: part 1. *Br J Med Pract* **2**, 20-28.
- [310] Proal, A. D., Albert, P. J. & Marshall, T. (2009). Autoimmune disease in the era of the metagenome. *Autoimmun Rev* **8**, 677-81.
- [311] Nicolson, G. L. & Haier, J. (2010). Role of chronic bacterial and viral infections in neurodegenerative, neurobehavioural, psychiatric, autoimmune and fatiguing illnesses: part 2. *Br J Med Pract* **3,** 301-310.
- [312] Miklossy, J. (2011). Alzheimer's disease a neurospirochetosis. Analysis of the evidence following Koch's and Hill's criteria. *J Neuroinflammation* **8**, 90.

- [313] Proal, A. D., Albert, P. J., Blaney, G. P., Lindseth, I. A., Benediktsson, C. & Marshall, T. G. (2011). Immunostimulation in the era of the metagenome. *Cell Mol Immunol* **8,** 213-25.
- [314] Brecher, M. E. & Hay, S. N. (2005). Bacterial contamination of blood components. *Clin Microbiol Rev* **18**, 195-204.
- [315] Halliwell, B. & Gutteridge, J. M. C. (2006). *Free Radicals in Biology and Medicine, 4th Ed.* Oxford University Press, Oxford.
- [316] Bollmann, A., Lewis, K. & Epstein, S. S. (2007). Incubation of environmental samples in a diffusion chamber increases the diversity of recovered isolates. *Appl Environ Microbiol* **73**, 6386-90.
- [317] D'Onofrio, A., Crawford, J. M., Stewart, E. J., Witt, K., Gavrish, E., Epstein, S., Clardy, J. & Lewis, K. (2010). Siderophores from neighboring organisms promote the growth of uncultured bacteria. *Chem Biol* **17**, 254-64.
- [318] Kaeberlein, T., Lewis, K. & Epstein, S. S. (2002). Isolating "uncultivable" microorganisms in pure culture in a simulated natural environment. *Science* **296**, 1127-9.
- [319] Lewis, K., Epstein, S., D'Onofrio, A. & Ling, L. L. (2010). Uncultured microorganisms as a source of secondary metabolites. *J Antibiot (Tokyo)* **63**, 468-76.
- [320] Ling, L. L., Schneider, T., Peoples, A. J., Spoering, A. L., Engels, I., Conlon, B. P., Mueller, A., Schäberle, T. F., Hughes, D. E., Epstein, S., Jones, M., Lazarides, L., Steadman, V. A., Cohen, D. R., Felix, C. R., Fetterman, K. A., Millett, W. P., Nitti, A. G., Zullo, A. M., Chen, C. & Lewis, K. (2015). A new antibiotic kills pathogens without detectable resistance. *Nature* 517, 455-459.
- [321] Deschner, J., Eick, S., Damanaki, A. & Nokhbehsaim, M. (2014). The role of adipokines in periodontal infection and healing. *Mol Oral Microbiol* **29**, 258-69.
- [322] Herrera, J. A., Chaudhuri, G. & López-Jaramillo, P. (2001). Is infection a major risk factor for preeclampsia? *Med Hypotheses* **57**, 393-7.
- [323] López-Jaramillo, P., Casas, J. P. & Serrano, N. (2001). Preeclampsia: from epidemiological observations to molecular mechanisms. *Braz J Med Biol Res* **34**, 1227-35.
- [324] von Dadelszen, P. & Magee, L. A. (2002). Could an infectious trigger explain the differential maternal response to the shared placental pathology of preeclampsia and normotensive intrauterine growth restriction? *Acta Obstet Gynecol Scand* **81**, 642-8.
- [325] Marshall, B. (2002). Helicobacter pylori: 20 years on. Clin Med 2, 147-52.
- [326] Figura, N., Franceschi, F., Santucci, A., Bernardini, G., Gasbarrini, G. & Gasbarrini, A. (2010). Extragastric manifestations of *Helicobacter pylori* infection. *Helicobacter* **15 Suppl 1**, 60-8.
- [327] Franceschi, F., Tortora, A., Gasbarrini, G. & Gasbarrini, A. (2014). *Helicobacter pylori* and extragastric diseases. *Helicobacter* **19 Suppl 1,** 52-8.
- [328] Pellicano, R., Franceschi, F., Saracco, G., Fagoonee, S., Roccarina, D. & Gasbarrini, A. (2009). Helicobacters and extragastric diseases. *Helicobacter* **14 Suppl 1,** 58-68.
- [329] Prelipcean, C. C., Mihai, C., Gogalniceanu, P., Mitrica, D., Drug, V. L. & Stanciu, C. (2007). Extragastric manifestations of Helicobacter pylori infection. *Rev Med Chir Soc Med Nat Iasi* **111**, 575-83.
- [330] Banić, M., Franceschi, F., Babić, Z. & Gasbarrini, A. (2012). Extragastric manifestations of *Helicobacter pylori* infection. *Helicobacter* **17 Suppl 1,** 49-55.
- [331] Roubaud Baudron, C., Franceschi, F., Salles, N. & Gasbarrini, A. (2013). Extragastric diseases and *Helicobacter pylori. Helicobacter* **18 Suppl 1,** 44-51.
- [332] Suzuki, H., Marshall, B. J. & Hibi, T. (2006). Overview: *Helicobacter pylori* and extragastric disease. *Int J Hematol* **84**, 291-300.
- [333] Franceschi, F., Zuccala, G., Roccarina, D. & Gasbarrini, A. (2014). Clinical effects of *Helicobacter pylori* outside the stomach. *Nat Rev Gastroenterol Hepatol* **11**, 234-42.
- [334] Testerman, T. L. & Morris, J. (2014). Beyond the stomach: an updated view of *Helicobacter pylori* pathogenesis, diagnosis, and treatment. *World J Gastroenterol* **20**, 12781-808.
- [335] Ponzetto, A., Cardaropoli, S., Piccoli, E., Rolfo, A., Gennero, L., Kanduc, D. & Todros, T. (2006). Pre-eclampsia is associated with *Helicobacter pylori* seropositivity in Italy. *J Hypertens* **24**, 2445-9.
- [336] Franceschi, F., Di Simone, N., D'Ippolito, S., Castellani, R., Di Nicuolo, F., Gasbarrini, G., Yamaoka, Y., Todros, T., Scambia, G. & Gasbarrini, A. (2012). Antibodies Anti-CagA Cross-React with Trophoblast Cells: A Risk Factor for Pre-Eclampsia? *Helicobacter* 17, 426-434.

- [337] Shimoda, A., Ueda, K., Nishiumi, S., Murata-Kamiya, N., Mukai, S. A., Sawada, S., Azuma, T., Hatakeyama, M. & Akiyoshi, K. (2016). Exosomes as nanocarriers for systemic delivery of the *Helicobacter pylori* virulence factor CagA. *Sci Rep* **6**, 18346.
- [338] Raghupathy, R. (2013). Cytokines as Key Players in the Pathophysiology of Preeclampsia. *Med Princ Pract* **22**, 8-19.
- [339] Wurdinger, T., Gatson, N. N., Balaj, L., Kaur, B., Breakefield, X. O. & Pegtel, D. M. (2012). Extracellular vesicles and their convergence with viral pathways. *Adv Virol* **2012**, 767694.
- [340] van Dongen, H. M., Masoumi, N., Witwer, K. W. & Pegtel, D. M. (2016). Extracellular Vesicles Exploit Viral Entry Routes for Cargo Delivery. *Microbiol Mol Biol Rev* **80**, 369-86.
- [341] Xie, F., Hu, Y., Magee, L. A., Money, D. M., Patrick, D. M., Brunham, R. M., Thomas, E., von Dadelszen, P. & for the Toxaemia group. (2010). *Chlamydia pneumoniae* infection in preeclampsia. *Hypertens Pregnancy* **29**, 468-77.
- [342] Heine, R. P., Ness, R. B. & Roberts, J. M. (2003). Seroprevalence of antibodies to *Chlamydia pneumoniae* in women with preeclampsia. *Obstet Gynecol* **101**, 221-6.
- [343] El-Shourbagy, M. A. A., El-Refaie, T. A., Sayed, K. K. A., Wahba, K. A. H., El-Din, A. S. S. & Fathy, M. M. (2011). Impact of seroconversion and antichlamydial treatment on the rate of pre-eclampsia among Egyptian primigravidae. *Int J Gynaecol Obstet* **113**, 137-40.
- [344] Haggerty, C. L., Klebanoff, M. A., Panum, I., Uldum, S. A., Bass, D. C., Olsen, J., Roberts, J. M. & Ness, R. B. (2013). Prenatal *Chlamydia trachomatis* infection increases the risk of preeclampsia. *Pregnancy Hypertens* **3**, 151-154.
- [345] Haggerty, C. L., Panum, I., Uldum, S. A., Bass, D. C., Olsen, J., Darville, T., Eastman, J. M., Simhan, H. N., Roberts, J. M. & Ness, R. B. (2013). *Chlamydia trachomatis* infection may increase the risk of preeclampsia. *Pregnancy Hypertension-an International Journal of Womens Cardiovascular Health* 3, 28-33.
- [346] Xie, F., Hu, Y., Magee, L. A., Money, D. M., Patrick, D. M., Krajden, M., Thomas, E., von Dadelszen, P. & Toxemia Study, G. (2010). An association between cytomegalovirus infection and pre-eclampsia: a case-control study and data synthesis. *Acta Obstet Gynecol Scand* **89**, 1162-7.
- [347] Xie, F., von Dadelszen, P. & Nadeau, J. (2014). CMV infection, TLR-2 and -4 expression, and cytokine profiles in early-onset preeclampsia with HELLP syndrome. *Am J Reprod Immunol* **71**, 379-86.
- [348] Panarelli, M. & Sattar, N. (2006). Pre-eclampsia associated with *Helicobacter pylori* seropositivity. *J Hypertens* **24**, 2353-2354.
- [349] Mosbah, A. & Nabiel, Y. (2016). *Helicobacter pylori, Chlamydiae pneumoniae* and trachomatis as probable etiological agents of preeclampsia. *J Matern-Fetal Neonat Med* **29,** 1607-1612.
- [350] Tersigni, C., Franceschi, F., Todros, T., Cardaropoli, S., Scambia, G. & Di Simone, N. (2014). Insights into the Role of *Helicobacter pylori* Infection in Preeclampsia: From the Bench to the Bedside. *Front Immunol* **5**, 484.
- [351] Üstün, Y., Engin-Üstün, Y., Ozkaplan, E., Otlu, B. & Sait Tekerekoğlu, M. (2010). Association of *Helicobacter pylori* infection with systemic inflammation in preeclampsia. *J Matern Fetal Neonatal Med* 23, 311-4.
- [352] Aksoy, H., Ozkan, A., Aktas, F. & Borekci, B. (2009). *Helicobacter pylori* Seropositivity and its Relationship with Serum Malondialdehyde and Lipid Profile in Preeclampsia. *J Clin Lab Anal* 23, 219-222.
- [353] Cardaropoli, S., Rolfo, A. & Todros, T. (2014). *Helicobacter pylori* and pregnancy-related disorders. *World J Gastroenterol* **20**, 654-64.
- [354] Pugliese, A., Beltramo, T., Todros, T., Cardaropoli, S. & Ponzetto, A. (2008). Interleukin-18 and gestosis: correlation with *Helicobacter pylori* seropositivity. *Cell Biochem Funct* **26**, 817-9.
- [355] Cardaropoli, S., Giuffrida, D., Piazzese, A. & Todros, T. (2015). *Helicobacter pylori* seropositivity and pregnancy-related diseases: a prospective cohort study. *J Reprod Immunol* **109**, 41-7.
- [356] Cardaropoli, S., Rolfo, A., Piazzese, A., Ponzetto, A. & Todros, T. (2011). *Helicobacter pylori*'s virulence and infection persistence define pre-eclampsia complicated by fetal growth retardation. *World J Gastroenterol* **17**, 5156-65.

- [357] McDonnold, M., Dunn, H., Hester, A., Pacheco, L. D., Hankins, G. D., Saade, G. R. & Costantine, M. M. (2014). High risk human papillomavirus at entry to prenatal care and risk of preeclampsia. *Am J Obstet Gynecol* **210**, 138 e1-5.
- [358] Hill, J. A., Devoe, L. D. & Bryans, C. I., Jr. (1986). Frequency of asymptomatic bacteriuria in preeclampsia. *Obstet Gynecol* **67**, 529-32.
- [359] Hsu, C. D. & Witter, F. R. (1995). Urogenital infection in preeclampsia. Int J Gynaecol Obstet 49, 271-5.
- [360] Mittendorf, R., Lain, K. Y., Williams, M. A. & Walker, C. K. (1996). Preeclampsia. A nested, case-control study of risk factors and their interactions. *J Reprod Med* **41**, 491-6.
- [361] Easter, S. R., Cantonwine, D. E., Zera, C. A., Lim, K. H., Parry, S. I. & McElrath, T. F. (2016). Urinary tract infection during pregnancy, angiogenic factor profiles, and risk of preeclampsia. *Am J Obs Gynecol* **214**.
- [362] Mazor-Dray, E., Levy, A., Schlaeffer, F. & Sheiner, E. (2009). Maternal urinary tract infection: is it independently associated with adverse pregnancy outcome? *J Matern Fetal Neonatal Med* **22**, 124-8.
- [363] Minassian, C., Thomas, S. L., Williams, D. J., Campbell, O. & Smeeth, L. (2013). Acute maternal infection and risk of pre-eclampsia: a population-based case-control study. *PLoS One* **8**, e73047.
- [364] Rezavand, N., Veisi, F., Zangane, M., Amini, R. & Almasi, A. (2016). Association between Asymptomatic Bacteriuria and Pre-Eclampsia. *Glob J Health Sci* **8**, 235-239.
- [365] Karmon, A. & Sheiner, E. (2008). The relationship between urinary tract infection during pregnancy and preeclampsia: causal, confounded or spurious? *Arch Gynecol Obstet* **277**, 479-81.
- [366] Villar, J., Carroli, G., Wojdyla, D., Abalos, E., Giordano, D., Ba'aqeel, H., Farnot, U., Bergsjø, P., Bakketeig, L., Lumbiganon, P., Campodonico, L., Al-Mazrou, Y., Lindheimer, M., Kramer, M. & World Health Organization Antenatal Care Trial Research Group. (2006). Preeclampsia, gestational hypertension and intrauterine growth restriction, related or independent conditions? *Am J Obstet Gynecol* **194**, 921-31.
- [367] Bánhidy, F., Ács, N., Puhó, E. H. & Czeizel, A. E. (2007). Pregnancy complications and birth outcomes of pregnant women with urinary tract infections and related drug treatments. *Scand J Infect Dis* **39**, 390-7.
- [368] López-Jaramillo, P., Herrera, J. A., Arenas-Mantilla, M., Jauregui, I. E. & Mendoza, M. A. (2008). Subclinical infection as a cause of inflammation in preeclampsia. *Am J Ther* **15**, 373-6.
- [369] Ide, M. & Papapanou, P. N. (2013). Epidemiology of association between maternal periodontal disease and adverse pregnancy outcomes--systematic review. *J Periodontol* **84,** S181-94.
- [370] Dunlop, A. L., Mulle, J. G., Ferranti, E. P., Edwards, S., Dunn, A. B. & Corwin, E. J. (2015). Maternal Microbiome and Pregnancy Outcomes That Impact Infant Health: A Review. *Adv Neonatal Care* **15**, 377-85.
- [371] Hlimi, T. (2015). Association of anemia, pre-eclampsia and eclampsia with seasonality: A realist systematic review. *Health & Place* **31**, 180-192.
- [372] Brabin, B. J. & Johnson, P. M. (2005). Placental malaria and pre-eclampsia through the looking glass backwards? *J Reprod Immunol* **65**, 1-15.
- [373] Anya, S. E. (2004). Seasonal variation in the risk and causes of maternal death in the Gambia: malaria appears to be an important factor. *Am J Trop Med Hyg* **70**, 510-3.
- [374] Sartelet, H., Rogier, C., Milko-Sartelet, I., Angel, G. & Michel, G. (1996). Malaria associated preeclampsia in Senegal. *Lancet* **347**, 1121.
- [375] Romero, R., Mazor, M., Wu, Y. K., Sirtori, M., Oyarzun, E., Mitchell, M. D. & Hobbins, J. C. (1988). Infection in the pathogenesis of preterm labor. *Semin Perinatol* **12**, 262-79.
- [376] Toth, M., Witkin, S. S., Ledger, W. & Thaler, H. (1988). The Role of Infection in the Etiology of Preterm Birth. *Obstet Gynecol* **71**, 723-726.
- [377] Cassell, G. H., Waites, K. B., Watson, H. L., Crouse, D. T. & Harasawa, R. (1993). *Ureaplasma urealyticum* intrauterine infection: role in prematurity and disease in newborns. *Clin Microbiol Rev* **6**, 69-87.
- [378] McGregor, J. A., French, J. I., Jones, W., Milligan, K., McKinney, P. J., Patterson, E. & Parker, R. (1994). Bacterial vaginosis is associated with prematurity and vaginal fluid mucinase and sialidase: results

- of a controlled trial of topical clindamycin cream. *Am J Obstet Gynecol* **170,** 1048-59; discussion 1059-60.
- [379] Goldenberg, R. L., Hauth, J. C. & Andrews, W. W. (2000). Intrauterine infection and preterm delivery. *N Engl J Med* **342**, 1500-7.
- [380] Gonçalves, L. F., Chaiworapongsa, T. & Romero, R. (2002). Intrauterine infection and prematurity. *Ment Retard Dev Disabil Res Rev* **8,** 3-13.
- [381] Gerber, S., Vial, Y., Hohlfeld, P. & Witkin, S. S. (2003). Detection of *Ureaplasma urealyticum* in second-trimester amniotic fluid by polymerase chain reaction correlates with subsequent preterm labor and delivery. *J Infect Dis* **187**, 518-21.
- [382] Gardella, C., Riley, D. E., Hitti, J., Agnew, K., Krieger, J. N. & Eschenbach, D. (2004). Identification and sequencing of bacterial rDNAs in culture-negative amniotic fluid from women in premature labor. *Am J Perinatol* **21**, 319-23.
- [383] Check, J. H. (2010). A practical approach to the prevention of miscarriage Part 4-role of infection. *Clin Exp Obstet Gyn* **37**, 252-255.
- [384] Frey, H. A. & Klebanoff, M. A. (2016). The epidemiology, etiology, and costs of preterm birth. *Semin Fetal Neonatal Med* **21**, 68-73.
- [385] Nadeau, H. C. G., Subramaniam, A. & Andrews, W. W. (2016). Infection and preterm birth. *Semin Fetal Neonatal Med* **21,** 100-5.
- [386] Vinturache, A. E., Gyamfi-Bannerman, C., Hwang, J., Mysorekar, I. U., Jacobsson, B. & Preterm Birth International Collaborative (PREBIC). (2016). Maternal microbiome A pathway to preterm birth. Semin Fetal Neonatal Med 21, 94-9.
- [387] Espinoza, J., Erez, O. & Romero, R. (2006). Preconceptional antibiotic treatment to prevent preterm birth in women with a previous preterm delivery. *Am J Obstet Gynecol* **194,** 630-7.
- [388] Joergensen, J. S., Kjaer Weile, L. K. & Lamont, R. F. (2014). The early use of appropriate prophylactic antibiotics in susceptible women for the prevention of preterm birth of infectious etiology. *Expert Opin Pharmacother* **15**, 2173-91.
- [389] Goldenberg, R. L., Culhane, J. F., Iams, J. D. & Romero, R. (2008). Epidemiology and causes of preterm birth. *Lancet* **371**, 75-84.
- [390] Bastek, J. A., Gómez, L. M. & Elovitz, M. A. (2011). The role of inflammation and infection in preterm birth. *Clin Perinatol* **38**, 385-406.
- [391] Johnson, H. L., Ghanem, K. G., Zenilman, J. M. & Erbelding, E. J. (2011). Sexually transmitted infections and adverse pregnancy outcomes among women attending inner city public sexually transmitted diseases clinics. *Sex Transm Dis* **38**, 167-71.
- [392] Rours, G. I. J. G., Duijts, L., Moll, H. A., Arends, L. R., de Groot, R., Jaddoe, V. W., Hofman, A., Steegers, E. A. P., Mackenbach, J. P., Ott, A., Willemse, H. F. M., van der Zwaan, E. A. E., Verkooijen, R. P. & Verbrugh, H. A. (2011). *Chlamydia trachomatis* infection during pregnancy associated with preterm delivery: a population-based prospective cohort study. *Eur J Epidemiol* **26**, 493-502.
- [393] Jefferson, K. K. (2012). The bacterial etiology of preterm birth. Adv Appl Microbiol 80, 1-22.
- [394] Subramaniam, A., Abramovici, A., Andrews, W. W. & Tita, A. T. (2012). Antimicrobials for Preterm Birth Prevention: An Overview. *Infect Dis Obs Gynecol* **2012**.
- [395] Aagaard, K., Ma, J., Antony, K. M., Ganu, R., Petrosino, J. & Versalovic, J. (2014). The placenta harbors a unique microbiome. *Sci Transl Med* **6,** 237ra65.
- [396] Kacerovsky, M., Vrbacky, F., Kutova, R., Pliskova, L., Andrys, C., Musilova, I., Menon, R., Lamont, R. & Nekvindova, J. (2015). Cervical microbiota in women with preterm prelabor rupture of membranes. *PLoS One* **10**, e0126884.
- [397] Lamont, R. F. (2015). Advances in the Prevention of Infection-Related Preterm Birth. *Front Immunol* **6**, 566.
- [398] Lee, S. Y. R. & Leung, C. W. (2012). Histological chorioamnionitis implication for bacterial colonization, laboratory markers of infection, and early onset sepsis in very-low-birth-weight neonates. *J Matern Fetal Neonatal Med* **25**, 364-8.
- [399] Allen-Daniels, M. J., Serrano, M. G., Pflugner, L. P., Fettweis, J. M., Prestosa, M. A., Koparde, V. N., Brooks, J. P., Strauss, J. F., Romero, R., Chaiworapongsa, T., Eschenbach, D. A., Buck, G. A. &

- Jefferson, K. K. (2015). Identification of a gene in *Mycoplasma hominis* associated with preterm birth and microbial burden in intraamniotic infection. *Am J Obstet Gynecol* **212**.
- [400] de Andrade Ramos, B., Kanninen, T. T., Sisti, G. & Witkin, S. S. (2015). Microorganisms in the female genital tract during pregnancy: tolerance versus pathogenesis. *Am J Reprod Immunol* **73**, 383-9.
- [401] Ueno, T., Niimi, H., Yoneda, N., Yoneda, S., Mori, M., Tabata, H., Minami, H., Saito, S. & Kitajima, I. (2015). Eukaryote-Made Thermostable DNA Polymerase Enables Rapid PCR-Based Detection of Mycoplasma, Ureaplasma and Other Bacteria in the Amniotic Fluid of Preterm Labor Cases. *PLoS One* 10, e0129032.
- [402] Yoneda, S., Shiozaki, A., Yoneda, N., Ito, M., Shima, T., Fukuda, K., Ueno, T., Niimi, H., Kitajima, I., Kigawa, M. & Saito, S. (2016). Antibiotic Therapy Increases the Risk of Preterm Birth in Preterm Labor without Intra-Amniotic Microbes, but may Prolong the Gestation Period in Preterm Labor with Microbes, Evaluated by Rapid and High-Sensitive PCR System. Am J Reprod Immunol 75, 440-50.
- [403] Kacerovsky, M., Lenco, J., Musilova, I., Tambor, V., Lamont, R., Torloni, M. R., Menon, R. & Group, P. B. W. (2014). Proteomic biomarkers for spontaneous preterm birth: a systematic review of the literature. *Reprod Sci* 21, 283-95.
- [404] Oliver, R. S. & Lamont, R. F. (2013). Infection and antibiotics in the aetiology, prediction and prevention of preterm birth. *J Obstet Gynaecol* **33**, 768-75.
- [405] McClure, E. M. & Goldenberg, R. L. (2009). Infection and stillbirth. *Semin Fetal Neonat Med* **14,** 182-189.
- [406] Menezes, E. V., Yakoob, M. Y., Soomro, T., Haws, R. A., Darmstadt, G. L. & Bhutta, Z. A. (2009). Reducing stillbirths: prevention and management of medical disorders and infections during pregnancy. *BMC Pregnancy Childbirth* **9 Suppl 1,** S4.
- [407] Nigro, G., Mazzocco, M., Mattia, E., Di Renzo, G. C., Carta, G. & Anceschi, M. M. (2011). Role of the infections in recurrent spontaneous abortion. *J Matern-Fetal Neo Med* **24**, 983-989.
- [408] Giakoumelou, S., Wheelhouse, N., Cuschieri, K., Entrican, G., Howie, S. E. M. & Horne, A. W. (2015). The role of infection in miscarriage. *Hum Reprod Update*.
- [409] Toth, A., Lesser, M. L., Brooks-Toth, C. W. & Feiner, C. (1986). Outcome of subsequent pregnancies following antibiotic therapy after primary or multiple spontaneous abortions. *Surg Gynecol Obstet* **163**, 243-50.
- [410] Onderdonk, A. B., Delaney, M. L., DuBois, A. M., Allred, E. N., Leviton, A. & Extremely Low Gestational Age Newborns Study Investigators. (2008). Detection of bacteria in placental tissues obtained from extremely low gestational age neonates. *Am J Obstet Gynecol* **198**, 110 e1-7.
- [411] Satokari, R., Gronroos, T., Laitinen, K., Salminen, S. & Isolauri, E. (2009). *Bifidobacterium* and *Lactobacillus* DNA in the human placenta. *Lett Appl Microbiol* **48,** 8-12.
- [412] Stout, M. J., Conlon, B., Landeau, M., Lee, I., Bower, C., Zhao, Q., Roehl, K. A., Nelson, D. M., Macones, G. A. & Mysorekar, I. U. (2013). Identification of intracellular bacteria in the basal plate of the human placenta in term and preterm gestations. *Am J Obstet Gynecol* **208**, 226 e1-7.
- [413] Doyle, R. M., Alber, D. G., Jones, H. E., Harris, K., Fitzgerald, F., Peebles, D. & Klein, N. (2014). Term and preterm labour are associated with distinct microbial community structures in placental membranes which are independent of mode of delivery. *Placenta* **35**, 1099-101.
- [414] Cao, B., Stout, M. J., Lee, I. & Mysorekar, I. U. (2014). Placental Microbiome and Its Role in Preterm Birth. *Neoreviews* **15**, e537-e545.
- [415] Cao, B. & Mysorekar, I. U. (2014). Intracellular bacteria in placental basal plate localize to extravillous trophoblasts. *Placenta* **35**, 139-42.
- [416] Mysorekar, I. U. & Cao, B. (2014). Microbiome in parturition and preterm birth. *Semin Reprod Med* **32,** 50-5.
- [417] Zheng, J., Xiao, X., Zhang, Q., Mao, L., Yu, M. & Xu, J. (2015). The Placental Microbiome Varies in Association with Low Birth Weight in Full-Term Neonates. *Nutrients* **7**, 6924-37.
- [418] Abrahamsson, T. R., Wu, R. Y. & Jenmalm, M. C. (2015). Gut microbiota and allergy: the importance of the pregnancy period. *Pediatr Res* **77**, 214-9.

- [419] Antony, K. M., Ma, J., Mitchell, K. B., Racusin, D. A., Versalovic, J. & Aagaard, K. (2015). The preterm placental microbiome varies in association with excess maternal gestational weight gain. *Am J Obstet Gynecol* **212**, 653 e1-16.
- [420] Garmi, G., Okopnik, M., Keness, Y., Zafran, N., Berkowitz, E. & Salim, R. (2015). Correlation between Clinical, Placental Histology and Microbiological Findings in Spontaneous Preterm Births. *Fetal Diagn Ther*.
- [421] Prince, A. L., Ma, J., Kannan, P. S., Alvarez, M., Gisslen, T., Harris, R. A., Sweeney, E. L., Knox, C. L., Lambers, D. S., Jobe, A. H., Chougnet, C. A., Kallapur, S. G. & Aagaard, K. M. (2016). The placental microbiome is altered among subjects with spontaneous preterm birth with and without chorioamnionitis. *Am J Obstet Gynecol*.
- [422] Collado, M. C., Rautava, S., Aakko, J., Isolauri, E. & Salminen, S. (2016). Human gut colonisation may be initiated *in utero* by distinct microbial communities in the placenta and amniotic fluid. *Sci Rep* **6**, 23129.
- [423] Viniker, D. A. (1999). Hypothesis on the role of sub-clinical bacteria of the endometrium (bacteria endometrialis) in gynaecological and obstetric enigmas. *Hum Reprod Update* **5**, 373-85.
- [424] Svare, J. A., Schmidt, H., Hansen, B. B. & Lose, G. (2006). Bacterial vaginosis in a cohort of Danish pregnant women: prevalence and relationship with preterm delivery, low birthweight and perinatal infections. *BJOG* **113**, 1419-25.
- [425] Sheldon, I. M. & Bromfield, J. J. (2011). Innate immunity in the human endometrium and ovary. *Am J Reprod Immunol* **66 Suppl 1,** 63-71.
- [426] Aagaard, K., Riehle, K., Ma, J., Segata, N., Mistretta, T. A., Coarfa, C., Raza, S., Rosenbaum, S., Van den Veyver, I., Milosavljevic, A., Gevers, D., Huttenhower, C., Petrosino, J. & Versalovic, J. (2012). A metagenomic approach to characterization of the vaginal microbiome signature in pregnancy. *PLoS One* 7, e36466.
- [427] Walther-António, M. R., Jeraldo, P., Berg Miller, M. E., Yeoman, C. J., Nelson, K. E., Wilson, B. A., White, B. A., Chia, N. & Creedon, D. J. (2014). Pregnancy's stronghold on the vaginal microbiome. *PLoS One* **9**, e98514.
- [428] Huang, Y. E., Wang, Y., He, Y., Ji, Y., Wang, L. P., Sheng, H. F., Zhang, M., Huang, Q. T., Zhang, D. J., Wu, J. J., Zhong, M. & Zhou, H. W. (2015). Homogeneity of the vaginal microbiome at the cervix, posterior fornix, and vaginal canal in pregnant Chinese women. *Microb Ecol* **69**, 407-14.
- [429] Witkin, S. S. (2015). The vaginal microbiome, vaginal anti-microbial defence mechanisms and the clinical challenge of reducing infection-related preterm birth. *BJOG* **122**, 213-8.
- [430] Payne, M. S. & Bayatibojakhi, S. (2014). Exploring preterm birth as a polymicrobial disease: an overview of the uterine microbiome. *Front Immunol* **5**, 595.
- [431] Verstraelen, H., Vilchez-Vargas, R., Desimpel, F., Jauregui, R., Vankeirsbilck, N., Weyers, S., Verhelst, R., De Sutter, P., Pieper, D. H. & Van De Wiele, T. (2016). Characterisation of the human uterine microbiome in non-pregnant women through deep sequencing of the V1-2 region of the 16S rRNA gene. *PeerJ* 4, e1602.
- [432] Dong, Y., St Clair, P. J., Ramzy, I., Kagan-Hallet, K. S. & Gibbs, R. S. (1987). A microbiologic and clinical study of placental inflammation at term. *Obstet Gynecol* **70**, 175-82.
- [433] Bearfield, C., Davenport, E. S., Sivapathasundaram, V. & Allaker, R. P. (2002). Possible association between amniotic fluid micro-organism infection and microflora in the mouth. *BJOG* **109**, 527-533.
- [434] Wassenaar, T. M. & Panigrahi, P. (2014). Is a foetus developing in a sterile environment? *Lett Appl Microbiol* **59**, 572-9.
- [435] Combs, C. A., Gravett, M., Garite, T. J., Hickok, D. E., Lapidus, J., Porreco, R., Rael, J., Grove, T., Morgan, T. K., Clewell, W., Miller, H., Luthy, D., Pereira, L., Nageotte, M., Robilio, P. A., Fortunato, S., Simhan, H., Baxter, J. K., Amon, E., Franco, A., Trofatter, K., Heyborne, K. & ProteoGenix/Obstetrix Collaborative Research Network. (2014). Amniotic fluid infection, inflammation, and colonization in preterm labor with intact membranes. Am J Obstet Gynecol 210, 125 e1-125 e15.
- [436] Combs, C. A., Garite, T. J., Lapidus, J. A., Lapointe, J. P., Gravett, M., Rael, J., Amon, E., Baxter, J. K., Brady, K., Clewell, W., Eddleman, K. A., Fortunato, S., Franco, A., Haas, D. M., Heyborne, K., Hickok, D. E., How, H. Y., Luthy, D., Miller, H., Nageotte, M., Pereira, L., Porreco, R., Robilio, P. A., Simhan,

- H., Sullivan, S. A., Trofatter, K., Westover, T. & Obstetrix Collaborative Research Network. (2015). Detection of microbial invasion of the amniotic cavity by analysis of cervicovaginal proteins in women with preterm labor and intact membranes. *Am J Obstet Gynecol* **212**, 482 e1-482 e12.
- [437] Koleva, P. T., Kim, J. S., Scott, J. A. & Kozyrskyj, A. L. (2015). Microbial programming of health and disease starts during fetal life. *Birth Defects Res C Embryo Today* **105**, 265-77.
- [438] Pelzer, E. S., Allan, J. A., Cunningham, K., Mengersen, K., Allan, J. M., Launchbury, T., Beagley, K. & Knox, C. L. (2011). Microbial colonization of follicular fluid: alterations in cytokine expression and adverse assisted reproduction technology outcomes. *Hum Reprod* **26**, 1799-1812.
- [439] Pelzer, E. S., Allan, J. A., Waterhouse, M. A., Ross, T., Beagley, K. W. & Knox, C. L. (2013). Microorganisms within human follicular fluid: effects on IVF. *PLoS One* **8**, e59062.
- [440] Barak, S., Oettinger-Barak, O., Machtei, E. E., Sprecher, H. & Ohel, G. (2007). Evidence of periopathogenic microorganisms in placentas of women with preeclampsia. *J Periodontol* **78**, 670-6.
- [441] McDonagh, S., Maidji, E., Ma, W., Chang, H. T., Fisher, S. & Pereira, L. (2004). Viral and bacterial pathogens at the maternal-fetal interface. *J Infect Dis* **190**, 826-34.
- [442] Amarasekara, R., Jayasekara, R. W., Senanayake, H. & Dissanayake, V. H. (2015). Microbiome of the placenta in pre-eclampsia supports the role of bacteria in the multifactorial cause of pre-eclampsia. *J Obstet Gynaecol Res* **41**, 662-669.
- [443] Muehlenbachs, A., Mutabingwa, T. K., Edmonds, S., Fried, M. & Duffy, P. E. (2006). Hypertension and maternal-fetal conflict during placental malaria. *PLoS Med* **3**, e446.
- [444] Duffy, P. E. (2007). *Plasmodium* in the placenta: parasites, parity, protection, prevention and possibly preeclampsia. *Parasitology* **134**, 1877-81.
- [445] Vanterpool, S. F., Been, J. V., Houben, M. L., Nikkels, P. G., De Krijger, R. R., Zimmermann, L. J., Kramer, B. W., Progulske-Fox, A. & Reyes, L. (2016). *Porphyromonas gingivalis* within Placental Villous Mesenchyme and Umbilical Cord Stroma Is Associated with Adverse Pregnancy Outcome. *PLoS One* **11**, e0146157.
- [446] Chaparro, A., Blanlot, C., Ramirez, V., Sanz, A., Quintero, A., Inostroza, C., Bittner, M., Navarro, M. & Illanes, S. E. (2013). *Porphyromonas gingivalis, Treponema denticola* and toll-like receptor 2 are associated with hypertensive disorders in placental tissue: a case-control study. *J Periodontal Res* **48**, 802-9.
- [447] Ebringer, A., Rashid, T. & Wilson, C. (2010). Rheumatoid arthritis, Proteus, anti-CCP antibodies and Karl Popper. *Autoimmun Rev* **9**, 216-23.
- [448] Ebringer, A. (2012). *Rheumatoid arthritis and Proteus*. Springer, London.
- [449] Ebringer, A. & Rashid, T. (2014). Rheumatoid arthritis is caused by a *Proteus* urinary tract infection. *APMIS* **122**, 363-8.
- [450] Vatanen, T., Kostic, A. D., d'Hennezel, E., Siljander, H., Franzosa, E. A., Yassour, M., Kolde, R., Vlamakis, H., Arthur, T. D., Hamalainen, A. M., Peet, A., Tillmann, V., Uibo, R., Mokurov, S., Dorshakova, N., Ilonen, J., Virtanen, S. M., Szabo, S. J., Porter, J. A., Lahdesmaki, H., Huttenhower, C., Gevers, D., Cullen, T. W., Knip, M., Group, D. S. & Xavier, R. J. (2016). Variation in Microbiome LPS Immunogenicity Contributes to Autoimmunity in Humans. *Cell* **165**, 842-53.
- [451] Ferrier, L., Mazelin, L., Cenac, N., Desreumaux, P., Janin, A., Emilie, D., Colombel, J. F., Garcia-Villar, R., Fioramonti, J. & Bueno, L. (2003). Stress-induced disruption of colonic epithelial barrier: role of interferon-gamma and myosin light chain kinase in mice. *Gastroenterology* **125**, 795-804.
- [452] Honda, K. & Littman, D. R. (2012). The microbiome in infectious disease and inflammation. *Annu Rev Immunol* **30**, 759-95.
- [453] Klatt, N. R., Funderburg, N. T. & Brenchley, J. M. (2013). Microbial translocation, immune activation, and HIV disease. *Trends Microbiol* **21**, 6-13.
- [454] Sawchuck, D. J. & Wittmann, B. K. (2014). Pre-eclampsia renamed and reframed: Intra-abdominal hypertension in pregnancy. *Med Hypotheses* **83**, 619-32.
- [455] Perez, P. F., Dore, J., Leclerc, M., Levenez, F., Benyacoub, J., Serrant, P., Segura-Roggero, I., Schiffrin, E. J. & Donnet-Hughes, A. (2007). Bacterial imprinting of the neonatal immune system: lessons from maternal cells? *Pediatrics* **119**, e724-32.
- [456] Gensollen, T., Iyer, S. S., Kasper, D. L. & Blumberg, R. S. (2016). How colonization by microbiota in early life shapes the immune system. *Science* **352**, 539-44.

- [457] Newnham, J. P., Newnham, I. A., Ball, C. M., Wright, M., Pennell, C. E., Swain, J. & Doherty, D. A. (2009). Treatment of periodontal disease during pregnancy: a randomized controlled trial. *Obstet Gynecol* **114**, 1239-48.
- [458] Huang, X., Wang, J., Liu, J., Hua, L., Zhang, D., Hu, T. & Ge, Z. L. (2014). Maternal Periodontal Disease and Risk of Preeclampsia: A Meta-analysis. *Journal of Huazhong University of Science and Technology-Medical Sciences* **34**, 729-735.
- [459] Gilbert, G. L., Garland, S. M., Fairley, K. F. & McDowall, D. M. (1986). Bacteriuria due to ureaplasmas and other fastidious organisms during pregnancy: prevalence and significance. *Pediatr Infect Dis* 5, \$239-43.
- [460] Ha, J. E., Jun, J. K., Ko, H. J., Paik, D. I. & Bae, K. H. (2014). Association between periodontitis and preeclampsia in never-smokers: a prospective study. *J Clin Periodontol* **41**, 869-874.
- [461] Sgolastra, F., Petrucci, A., Severino, M., Gatto, R. & Monaco, A. (2013). Relationship between periodontitis and pre-eclampsia: a meta-analysis. *PLoS One* **8**, e71387.
- [462] Boggess, K. A. & Edelstein, B. L. (2006). Oral health in women during preconception and pregnancy: implications for birth outcomes and infant oral health. *Matern Child Health J* **10**, S169-74.
- [463] Boggess, K. A., Berggren, E. K., Koskenoja, V., Urlaub, D. & Lorenz, C. (2013). Severe Preeclampsia and Maternal Self-Report of Oral Health, Hygiene, and Dental Care. *J Periodontol* **84,** 143-151.
- [464] Armitage, G. C. (2013). Bi-directional relationship between pregnancy and periodontal disease. *Periodontol 2000* **61,** 160-76.
- [465] Moura da Silva, G., Coutinho, S. B., Piscoya, M. D. B. V., Ximenes, R. A. A. & Jamelli, S. R. (2012). Periodontitis as a Risk Factor for Preeclampsia. *J Periodontol* **83**, 1388-1396.
- [466] Jahromi, B. N., Adibi, R., Adibi, S. & Salarian, L. (2014). Periodontal Disease as a Risk Factor for Preeclampsia. *Womens Health Bull* 1, e18908.
- [467] Lachat, M. F., Solnik, A. L., Nana, A. D. & Citron, T. L. (2011). Periodontal disease in pregnancy: review of the evidence and prevention strategies. *J Perinat Neonatal Nurs* **25**, 312-9.
- [468] Politano, G. T., Passini, R., Nomura, M. L., Velloso, L., Morari, J. & Couto, E. (2011). Correlation between periodontal disease, inflammatory alterations and pre-eclampsia. *J Periodontal Res* **46**, 505-11.
- [469] Nabet, C., Lelong, N., Colombier, M. L., Sixou, M., Musset, A. M., Goffinet, F., Kaminski, M. & Epipap Group. (2010). Maternal periodontitis and the causes of preterm birth: the case-control Epipap study. *J Clin Periodontol* **37**, 37-45.
- [470] Madianos, P. N., Bobetsis, Y. A. & Offenbacher, S. (2013). Adverse pregnancy outcomes (APOs) and periodontal disease: pathogenic mechanisms. *J Periodontol* **84,** S170-80.
- [471] Bobetsis, Y. A., Barros, S. P. & Offenbacher, S. (2006). Exploring the relationship between periodontal disease and pregnancy complications. *J Am Dent Assoc* **137 Suppl,** 7S-13S.
- [472] Herrera, J. A., Parra, B., Herrera, E., Botero, J. E., Arce, R. M., Contreras, A. & López-Jaramillo, P. (2007). Periodontal disease severity is related to high levels of C-reactive protein in pre-eclampsia. *J Hypertens* **25**, 1459-64.
- [473] Ruma, M., Boggess, K., Moss, K., Jared, H., Murtha, A., Beck, J. & Offenbacher, S. (2008). Maternal periodontal disease, systemic inflammation, and risk for preeclampsia. *American Journal of Obstetrics and Gynecology* **198**.
- [474] Pralhad, S., Thomas, B. & Kushtagi, P. (2013). Periodontal disease and pregnancy hypertension: a clinical correlation. *J Periodontol* **84,** 1118-25.
- [475] Zi, M. Y., Longo, P. L., Bueno-Silva, B. & Mayer, M. P. (2014). Mechanisms Involved in the Association between Periodontitis and Complications in Pregnancy. *Front Public Health* **2**, 290.
- [476] Swati, P., Thomas, B., Vahab, S. A., Kapaettu, S. & Kushtagi, P. (2012). Simultaneous detection of periodontal pathogens in subgingival plaque and placenta of women with hypertension in pregnancy. *Arch Gynecol Obstet* **285**, 613-9.
- [477] Shub, A., Swain, J. R. & Newnham, J. P. (2006). Periodontal disease and adverse pregnancy outcomes. *J Matern Fetal Neonatal Med* **19,** 521-8.
- [478] Xiong, X., Buekens, P., Fraser, W. D., Beck, J. & Offenbacher, S. (2006). Periodontal disease and adverse pregnancy outcomes: a systematic review. *BJOG* **113**, 135-43.

- [479] Ha, J. E., Oh, K. J., Yang, H. J., Jun, J. K., Jin, B. H., Paik, D. I. & Bae, K. H. (2011). Oral health behaviors, periodontal disease, and pathogens in preeclampsia: a case-control study in Korea. *J Periodontol* 82, 1685-92.
- [480] Komine-Aizawa, S., Hirohata, N., Aizawa, S., Abiko, Y. & Hayakawa, S. (2015). *Porphyromonas gingivalis* lipopolysaccharide inhibits trophoblast invasion in the presence of nicotine. *Placenta* **36**, 27-33.
- [481] Contreras, A., Herrera, J. A., Soto, J. E., Arce, R. M., Jaramillo, A. & Botero, J. E. (2006). Periodontitis is associated with preeclampsia in pregnant women. *Journal of Periodontology* **77**, 182-188.
- [482] Oettinger-Barak, O., Barak, S., Ohel, G., Oettinger, M., Kreutzer, H., Peled, M. & Machtei, E. E. (2005). Severe pregnancy complication (preeclampsia) is associated with greater periodontal destruction. *J Periodontol* **76**, 134-7.
- [483] Foxman, B. (1990). Recurring urinary tract infection: incidence and risk factors. *Am J Public Health* **80**, 331-3.
- [484] Foxman, B. (2002). Epidemiology of urinary tract infections: incidence, morbidity, and economic costs. *Am J Med* **113 Suppl 1A,** 5S-13S.
- [485] Marrs, C. F., Zhang, L. & Foxman, B. (2005). *Escherichia coli* mediated urinary tract infections: are there distinct uropathogenic *E. coli* (UPEC) pathotypes? *FEMS Microbiol Lett* **252**, 183-90.
- [486] Hannan, T. J., Mysorekar, I. U., Hung, C. S., Isaacson-Schmid, M. L. & Hultgren, S. J. (2010). Early severe inflammatory responses to uropathogenic *E. coli* predispose to chronic and recurrent urinary tract infection. *PLoS Pathog* **6**, e1001042.
- [487] Hunstad, D. A. & Justice, S. S. (2010). Intracellular lifestyles and immune evasion strategies of uropathogenic *Escherichia coli*. *Annu Rev Microbiol* **64,** 203-21.
- [488] Ejrnæs, K. (2011). Bacterial characteristics of importance for recurrent urinary tract infections caused by *Escherichia coli*. *Dan Med Bull* **58**, B4187.
- [489] Hannan, T. J., Totsika, M., Mansfield, K. J., Moore, K. H., Schembri, M. A. & Hultgren, S. J. (2012). Host-pathogen checkpoints and population bottlenecks in persistent and intracellular uropathogenic *Escherichia coli* bladder infection. *FEMS Microbiol Rev* **36**, 616-48.
- [490] Foxman, B. (2014). Urinary tract infection syndromes: occurrence, recurrence, bacteriology, risk factors, and disease burden. *Infect Dis Clin North Am* **28**, 1-13.
- [491] Bower, J. M., Eto, D. S. & Mulvey, M. A. (2005). Covert operations of uropathogenic *Escherichia coli* within the urinary tract. *Traffic* **6**, 18-31.
- [492] Justice, S. S., Hung, C., Theriot, J. A., Fletcher, D. A., Anderson, G. G., Footer, M. J. & Hultgren, S. J. (2004). Differentiation and developmental pathways of uropathogenic *Escherichia coli* in urinary tract pathogenesis. *Proc Natl Acad Sci U S A* **101**, 1333-8.
- [493] Rosen, D. A., Hooton, T. M., Stamm, W. E., Humphrey, P. A. & Hultgren, S. J. (2007). Detection of intracellular bacterial communities in human urinary tract infection. *PLoS Med* **4**, e329.
- [494] Dhakal, B. K., Kulesus, R. R. & Mulvey, M. A. (2008). Mechanisms and consequences of bladder cell invasion by uropathogenic *Escherichia coli*. *Eur J Clin Invest* **38 Suppl 2,** 2-11.
- [495] Schwartz, D. J., Chen, S. L., Hultgren, S. J. & Seed, P. C. (2011). Population dynamics and niche distribution of uropathogenic *Escherichia coli* during acute and chronic urinary tract infection. *Infect Immun* 79, 4250-9.
- [496] Brauner, A., Jacobson, S. H. & Kuhn, I. (1992). Urinary *Escherichia coli* causing recurrent infections--a prospective follow-up of biochemical phenotypes. *Clin Nephrol* **38**, 318-23.
- [497] Russo, T. A., Stapleton, A., Wenderoth, S., Hooton, T. M. & Stamm, W. E. (1995). Chromosomal restriction fragment length polymorphism analysis of *Escherichia coli strains* causing recurrent urinary tract infections in young women. *J Infect Dis* **172**, 440-5.
- [498] Ikäheimo, R., Siitonen, A., Heiskanen, T., Kärkkäinen, U., Kuosmanen, P., Lipponen, P. & Mäkelä, P. H. (1996). Recurrence of urinary tract infection in a primary care setting: analysis of a 1-year follow-up of 179 women. *Clin Infect Dis* **22**, 91-9.
- [499] Rosen, D. A., Pinkner, J. S., Jones, J. M., Walker, J. N., Clegg, S. & Hultgren, S. J. (2008). Utilization of an intracellular bacterial community pathway in *Klebsiella pneumoniae* urinary tract infection and the effects of FimK on type 1 pilus expression. *Infect Immun* **76**, 3337-45.

- [500] Luo, Y., Ma, Y., Zhao, Q., Wang, L., Guo, L., Ye, L., Zhang, Y. & Yang, J. (2012). Similarity and divergence of phylogenies, antimicrobial susceptibilities, and virulence factor profiles of *Escherichia coli* isolates causing recurrent urinary tract infections that persist or result from reinfection. *J Clin Microbiol* **50**, 4002-7.
- [501] Özlü, T., Alçelik, A., Çalışkan, B. & Dönmez, M. E. (2012). Preeclampsia: is it because of the asymptomatic, unrecognized renal scars caused by urinary tract infections in childhood that become symptomatic with pregnancy? *Med Hypotheses* **79**, 653-5.
- [502] Kincaid-Smith, P. & Bullen, M. (1965). Bacteriuria in Pregnancy. Lancet 1, 395-9.
- [503] Loh, K. & Sivalingam, N. (2007). Urinary tract infections in pregnancy. Malays Fam Physician 2, 54-7.
- [504] Macejko, A. M. & Schaeffer, A. J. (2007). Asymptomatic bacteriuria and symptomatic urinary tract infections during pregnancy. *Urol Clin North Am* **34**, 35-42.
- [505] Schnarr, J. & Smaill, F. (2008). Asymptomatic bacteriuria and symptomatic urinary tract infections in pregnancy. *Eur J Clin Invest* **38 Suppl 2,** 50-7.
- [506] Imade, P. E., Izekor, P. E., Eghafona, N. O., Enabulele, O. I. & Ophori, E. (2010). Asymptomatic bacteriuria among pregnant women. *N Am J Med Sci* **2,** 263-6.
- [507] Gilbert, N. M., O'Brien, V. P., Hultgren, S., Macones, G., Lewis, W. G. & Lewis, A. L. (2013). Urinary tract infection as a preventable cause of pregnancy complications: opportunities, challenges, and a global call to action. *Glob Adv Health Med* **2**, 59-69.
- [508] Glaser, A. P. & Schaeffer, A. J. (2015). Urinary Tract Infection and Bacteriuria in Pregnancy. *Urol Clin North Am* **42**, 547-60.
- [509] Germain, S. J., Sacks, G. P., Sooranna, S. R., Sargent, I. L. & Redman, C. W. G. (2007). Systemic inflammatory priming in normal pregnancy and preeclampsia: the role of circulating syncytiotrophoblast microparticles. *J Immunology* **178**, 5949-56.
- [510] Redman, C. W. G. & Sargent, I. L. (2008). Circulating microparticles in normal pregnancy and preeclampsia. *Placenta* **29 Suppl A,** S73-7.
- [511] Redman, C. W. G., Tannetta, D. S., Dragovic, R. A., Gardiner, C., Southcombe, J. H., Collett, G. P. & Sargent, I. L. (2012). Review: Does size matter? Placental debris and the pathophysiology of preeclampsia. *Placenta* **33 Suppl**, S48-54.
- [512] Goulopoulou, S. & Davidge, S. T. (2015). Molecular mechanisms of maternal vascular dysfunction in preeclampsia. *Trends in Molecular Medicine* **21**, 88-97.
- [513] Niccolai, E., Emmi, G., Squatrito, D., Silvestri, E., Emmi, L., Amedei, A. & Prisco, D. (2015). Microparticles: Bridging the Gap between Autoimmunity and Thrombosis. *Semin Thromb Hemost* **41**, 413-422.
- [514] Souza, A. C. P., Yuen, P. S. T. & Star, R. A. (2015). Microparticles: markers and mediators of sepsis-induced microvascular dysfunction, immunosuppression, and AKI. *Kidney Int* **87**, 1100-8.
- [515] Mitchell, M. D., Peiris, H. N., Kobayashi, M., Koh, Y. Q., Duncombe, G., Illanes, S. E., Rice, G. E. & Salomon, C. (2015). Placental exosomes in normal and complicated pregnancy. *Am J Obstet Gynecol* **213**, S173-81.
- [516] Mawson, A. R. (2003). Effects of antiretroviral therapy on occurrence of pre-eclampsia. *Lancet* **361**, 347-8
- [517] Suy, A., Martínez, E., Coll, O., Lonca, M., Palacio, M., de Lazzari, E., Larrousse, M., Milinkovic, A., Hernández, S., Blanco, J. L., Mallolas, J., León, A., Vanrell, J. A. & Gatell, J. M. (2006). Increased risk of pre-eclampsia and fetal death in HIV-infected pregnant women receiving highly active antiretroviral therapy. *AIDS* **20**, 59-66.
- [518] Wimalasundera, R. C., Larbalestier, N., Smith, J. H., de Ruiter, A., Mc, G. T. S. A., Hughes, A. D., Poulter, N., Regan, L. & Taylor, G. P. (2002). Pre-eclampsia, antiretroviral therapy, and immune reconstitution. *Lancet* **360**, 1152-4.
- [519] Hall, D. R. (2007). Is pre-eclampsia less common in patients with HIV/AIDS? *J Reprod Immunol* **76,** 75-7.
- [520] Adams, J. W., Watts, D. H. & Phelps, B. R. (2016). A systematic review of the effect of HIV infection and antiretroviral therapy on the risk of pre-eclampsia. *Int J Gynaecol Obstet* **133**, 17-21.

- [521] Todros, T., Verdiglione, P., Oggè, G., Paladini, D., Vergani, P. & Cardaropoli, S. (2006). Low incidence of hypertensive disorders of pregnancy in women treated with spiramycin for toxoplasma infection. *Br J Clin Pharmacol* **61**, 336-40.
- [522] Alvarado-Esquivel, C., Vázquez-Alaníz, F., Sandoval-Carrillo, A. A., Salas-Pacheco, J. M., Hernández-Tinoco, J., Sanchez-Anguiano, L. F. & Liesenfeld, O. (2014). Lack of association between *Toxoplasma gondii* infection and hypertensive disorders in pregnancy: a case-control study in a Northern Mexican population. *Parasites & Vectors* 7.
- [523] Hall, D., Gebhardt, S., Theron, G. & Grové, D. (2014). Pre-eclampsia and gestational hypertension are less common in HIV infected women. *Pregnancy Hypertens* **4,** 91-6.
- [524] McCarthy, F. P., Kingdom, J. C., Kenny, L. C. & Walsh, S. K. (2011). Animal models of preeclampsia; uses and limitations. *Placenta* **32**, 413-9.
- [525] Kumasawa, K., Ikawa, M., Kidoya, H., Hasuwa, H., Saito-Fujita, T., Morioka, Y., Takakura, N., Kimura, T. & Okabe, M. (2011). Pravastatin induces placental growth factor (PGF) and ameliorates preeclampsia in a mouse model. *Proc Natl Acad Sci U S A* **108**, 1451-5.
- [526] Aubuchon, M., Schulz, L. C. & Schust, D. J. (2011). Preeclampsia: animal models for a human cure. *Proc Natl Acad Sci* **108**, 1197-8.
- [527] Faas, M. M., Schuiling, G. A., Baller, J. F., Visscher, C. A. & Bakker, W. W. (1994). A new animal model for human preeclampsia: ultra-low-dose endotoxin infusion in pregnant rats. *Am J Obstet Gynecol* **171**, 158-64.
- [528] Faas, M. M., Schuiling, G. A., Linton, E. A., Sargent, I. L. & Redman, C. W. G. (2000). Activation of peripheral leukocytes in rat pregnancy and experimental preeclampsia. *Am J Obstet Gynecol* 182, 351-357.
- [529] Sakawi, Y., Tarpey, M., Chen, Y. F., Calhoun, D. A., Connor, M. G., Chestnut, D. H. & Parks, D. A. (2000). Evaluation of low-dose endotoxin administration during pregnancy as a model of preeclampsia. *Anesthesiology* **93**, 1446-55.
- [530] Lin, F., Zeng, P., Xu, Z. Y., Ye, D. Y., Yu, X. F., Wang, N., Tang, J., Zhou, Y. & Huang, Y. P. (2012). Treatment of Lipoxin A<sub>4</sub> and its analogue on low-dose endotoxin induced preeclampsia in rat and possible mechanisms. *Reprod Toxicol* **34**, 677-685.
- [531] Cotechini, T., Komisarenko, M., Sperou, A., Macdonald-Goodfellow, S., Adams, M. A. & Graham, C. H. (2014). Inflammation in rat pregnancy inhibits spiral artery remodeling leading to fetal growth restriction and features of preeclampsia. *J Exp Med* **211**, 165-79.
- [532] Xue, P. P., Zheng, M. M., Gong, P., Lin, C. M., Zhou, J. J., Li, Y. J., Shen, L., Diao, Z. Y., Yan, G. J., Sun, H. X. & Hu, Y. L. (2015). Single Administration of Ultra-Low-Dose Lipopolysaccharide in Rat Early Pregnancy Induces TLR4 Activation in the Placenta Contributing to Preeclampsia. *Plos One* 10.
- [533] Kalkunte, S., Boij, R., Norris, W., Friedman, J., Lai, Z., Kurtis, J., Lim, K. H., Padbury, J. F., Matthiesen, L. & Sharma, S. (2010). Sera from preeclampsia patients elicit symptoms of human disease in mice and provide a basis for an in vitro predictive assay. *Am J Pathol* **177**, 2387-98.
- [534] Xie, F., Turvey, S. E., Williams, M. A., Mor, G. & von Dadelszen, P. (2010). Toll-like receptor signaling and pre-eclampsia. *Am J Reprod Immunol* **63,** 7-16.
- [535] Anton, L., Brown, A. G., Parry, S. & Elovitz, M. A. (2012). Lipopolysaccharide induces cytokine production and decreases extravillous trophoblast invasion through a mitogen-activated protein kinase-mediated pathway: possible mechanisms of first trimester placental dysfunction. *Hum Reprod* 27, 61-72.
- [536] Chatterjee, P., Weaver, L. E., Doersch, K. M., Kopriva, S. E., Chiasson, V. L., Allen, S. J., Narayanan, A. M., Young, K. J., Jones, K. A., Kuehl, T. J. & Mitchell, B. M. (2012). Placental Toll-like receptor 3 and Toll-like receptor 7/8 activation contributes to preeclampsia in humans and mice. *PLoS One* 7, e41884.
- [537] Pineda, A., Verdin-Terán, S. L., Camacho, A. & Moreno-Fierros, L. (2011). Expression of toll-like receptor TLR-2, TLR-3, TLR-4 and TLR-9 is increased in placentas from patients with preeclampsia. *Arch Med Res* **42**, 382-91.
- [538] Ahn, H., Park, J., Gilman-Sachs, A. & Kwak-Kim, J. (2011). Immunologic characteristics of preeclampsia, a comprehensive review. *Am J Reprod Immunol* **65,** 377-94.

- [539] Gallo, P. M., Rapsinski, G. J., Wilson, R. P., Oppong, G. O., Sriram, U., Goulian, M., Buttaro, B., Caricchio, R., Gallucci, S. & Tükel, Ç. (2015). Amyloid-DNA Composites of Bacterial Biofilms Stimulate Autoimmunity. *Immunity* **42**, 1171-84.
- [540] Spaulding, C. N., Dodson, K. W., Chapman, M. R. & Hultgren, S. J. (2015). Fueling the Fire with Fibers: Bacterial Amyloids Promote Inflammatory Disorders. *Cell Host Microbe* **18**, 1-2.
- [541] Rapsinski, G. J., Wynosky-Dolfi, M. A., Oppong, G. O., Tursi, S. A., Wilson, R. P., Brodsky, I. E. & Tükel, Ç. (2015). Toll-like receptor 2 and NLRP3 cooperate to recognize a functional bacterial amyloid, curli. *Infect Immun* **83**, 693-701.
- [542] Wucherpfennig, K. W. (2001). Mechanisms for the induction of autoimmunity by infectious agents. *J Clin Invest* **108**, 1097-104.
- [543] Wucherpfennig, K. W. (2001). Structural basis of molecular mimicry. J Autoimmun 16, 293-302.
- [544] Kohm, A. P., Fuller, K. G. & Miller, S. D. (2003). Mimicking the way to autoimmunity: an evolving theory of sequence and structural homology. *Trends Microbiol* **11**, 101-5.
- [545] Morris, J. A., Broughton, S. J. & Wessels, Q. (2016). Microbes, molecular mimicry and molecules of mood and motivation. *Med Hypotheses* **87**, 40-3.
- [546] Jain, D., Kaur, K. J., Goel, M. & Salunke, D. M. (2000). Structural basis of functional mimicry between carbohydrate and peptide ligands of con A. *Biochem Biophys Res Commun* **272**, 843-9.
- [547] Goel, M., Krishnan, L., Kaur, S., Kaur, K. J. & Salunke, D. M. (2004). Plasticity within the antigen-combining site may manifest as molecular mimicry in the humoral immune response. *J Immunol* **173,** 7358-67.
- [548] Uh, A., Nicholson, R. C., Gonzalez, G. V., Simmons, C. F., Gombart, A., Smith, R. & Equils, O. (2008). Lipopolysaccharide stimulation of trophoblasts induces corticotropin-releasing hormone expression through MyD88. *Am J Obstet Gynecol* **199**, 317 e1-6.
- [549] Chen, Q., Viall, C., Kang, Y., Liu, B., Stone, P. & Chamley, L. (2009). Anti-phospholipid antibodies increase non-apoptotic trophoblast shedding: a contribution to the pathogenesis of pre-eclampsia in affected women? *Placenta* **30**, 767-73.
- [550] Chen, Q., Guo, F., Hensby-Bennett, S., Stone, P. & Chamley, L. (2012). Antiphospholipid antibodies prolong the activation of endothelial cells induced by necrotic trophoblastic debris: implications for the pathogenesis of preeclampsia. *Placenta* **33**, 810-5.
- [551] Pantham, P., Rosario, R., Chen, Q., Print, C. G. & Chamley, L. W. (2012). Transcriptomic analysis of placenta affected by antiphospholipid antibodies: following the TRAIL of trophoblast death. *J Reprod Immunol* **94,** 151-4.
- [552] Tong, M., Viall, C. A. & Chamley, L. W. (2015). Antiphospholipid antibodies and the placenta: a systematic review of their in vitro effects and modulation by treatment. *Hum Reprod Update* **21**, 97-118.
- [553] Dechend, R., Homuth, V., Wallukat, G., Kreuzer, J., Park, J. K., Theuer, J., Juepner, A., Gulba, D. C., Mackman, N., Haller, H. & Luft, F. C. (2000). AT(1) receptor agonistic antibodies from preeclamptic patients cause vascular cells to express tissue factor. *Circulation* **101**, 2382-7.
- [554] Roberts, J. M. (2000). Angiotensin-1 receptor autoantibodies: A role in the pathogenesis of preeclampsia? *Circulation* **101**, 2335-7.
- [555] Wallukat, G., Neichel, D., Nissen, E., Homuth, V. & Luft, F. C. (2003). Agonistic autoantibodies directed against the angiotensin II AT1 receptor in patients with preeclampsia. *Can J Physiol Pharmacol* **81**, 79-83.
- [556] Dechend, R., Muller, D. N., Wallukat, G., Homuth, V., Krause, M., Dudenhausen, J. & Luft, F. C. (2004). AT1 receptor agonistic antibodies, hypertension, and preeclampsia. *Semin Nephrol* **24,** 571-9.
- [557] Dechend, R., Homuth, V., Wallukat, G., Muller, D. N., Krause, M., Dudenhausen, J., Haller, H. & Luft, F. C. (2006). Agonistic antibodies directed at the angiotensin II, AT1 receptor in preeclampsia. *J Soc Gynecol Investig* 13, 79-86.
- [558] Hubel, C. A., Wallukat, G., Wolf, M., Herse, F., Rajakumar, A., Roberts, J. M., Markovic, N., Thadhani, R., Luft, F. C. & Dechend, R. (2007). Agonistic angiotensin II type 1 receptor autoantibodies in postpartum women with a history of preeclampsia. *Hypertension* 49, 612-7.
- [559] Dechend, R. & Luft, F. C. (2008). Are we getting closer to a Nobel prize for unraveling preeclampsia? *Curr Cardiol Rep* **10**, 440-7.

- [560] Herse, F., Staff, A. C., Hering, L., Müller, D. N., Luft, F. C. & Dechend, R. (2008). AT1-receptor autoantibodies and uteroplacental RAS in pregnancy and pre-eclampsia. *J Mol Med (Berl)* **86,** 697-703.
- [561] Zhou, C. C., Zhang, Y., Irani, R. A., Zhang, H., Mi, T., Popek, E. J., Hicks, M. J., Ramin, S. M., Kellems, R. E. & Xia, Y. (2008). Angiotensin receptor agonistic autoantibodies induce pre-eclampsia in pregnant mice. *Nat Med* **14**, 855-62.
- [562] Lorquet, S., Pequeux, C., Munaut, C. & Foidart, J. M. (2010). Aetiology and physiopathology of preeclampsia and related forms. *Acta Clin Belg* **65**, 237-41.
- [563] Parrish, M. R., Murphy, S. R., Rutland, S., Wallace, K., Wenzel, K., Wallukat, G., Keiser, S., Ray, L. F., Dechend, R., Martin, J. N., Granger, J. P. & LaMarca, B. (2010). The effect of immune factors, tumor necrosis factor-alpha, and agonistic autoantibodies to the angiotensin II type I receptor on soluble fms-like tyrosine-1 and soluble endoglin production in response to hypertension during pregnancy. *Am J Hypertens* **23**, 911-6.
- [564] Carbillon, L. (2011). AT1-receptor autoantibody: a true causal factor of pre-eclampsia or only a marker of poor placentation? *Am J Hypertens* **24,** 375; author reply 376.
- [565] Herse, F. & LaMarca, B. (2013). Angiotensin II type 1 receptor autoantibody (AT1-AA)-mediated pregnancy hypertension. *Am J Reprod Immunol* **69,** 413-8.
- [566] Siddiqui, A. H., Irani, R. A., Zhang, W., Wang, W., Blackwell, S. C., Kellems, R. E. & Xia, Y. (2013). Angiotensin receptor agonistic autoantibody-mediated soluble fms-like tyrosine kinase-1 induction contributes to impaired adrenal vasculature and decreased aldosterone production in preeclampsia. *Hypertension* **61**, 472-9.
- [567] Pietarinen, I., Kivinen, S., Ylostalo, P., Makitalo, R. & Laakso, L. (1982). Smooth muscle antibodies in pre-eclampsia of pregnancy. *Gynecol Obstet Invest* **13**, 142-9.
- [568] Alanen, A. (1984). Serum IgE and smooth muscle antibodies in pre-eclampsia. *Acta Obstet Gynecol Scand* **63**, 581-2.
- [569] Dagenais, N. J. & Jamali, F. (2005). Protective effects of angiotensin II interruption: evidence for antiinflammatory actions. *Pharmacotherapy* **25**, 1213-29.
- [570] Fliser, D., Buchholz, K., Haller, H., Olmesartan, E. U. T. o., Pravastatin in, I. & Atherosclerosis, I. (2004). Antiinflammatory effects of angiotensin II subtype 1 receptor blockade in hypertensive patients with microinflammation. *Circulation* **110**, 1103-7.
- [571] Platten, M., Youssef, S., Hur, E. M., Ho, P. P., Han, M. H., Lanz, T. V., Phillips, L. K., Goldstein, M. J., Bhat, R., Raine, C. S., Sobel, R. A. & Steinman, L. (2009). Blocking angiotensin-converting enzyme induces potent regulatory T cells and modulates TH1- and TH17-mediated autoimmunity. *Proc Natl Acad Sci U S A* **106**, 14948-53.
- [572] Ma, G., Li, Y., Zhang, J., Liu, H., Hou, D., Zhu, L., Zhang, Z. & Zhang, L. (2013). Association between the presence of autoantibodies against adrenoreceptors and severe pre-eclampsia: a pilot study. *PLoS One* **8**, e57983.
- [573] Li, Y., Ma, G., Zhang, Z., Yue, Y., Yuan, Y., Wang, Y., Miao, G. & Zhang, L. (2013). Association of autoantibodies against the M2-muscarinic receptor with perinatal outcomes in women with severe preeclampsia. *J Transl Med* **11**, 285.
- [574] Cornelius, D. C. & Lamarca, B. (2014). TH17- and IL-17- mediated autoantibodies and placental oxidative stress play a role in the pathophysiology of pre-eclampsia. *Minerva Ginecol* **66**, 243-9.
- [575] Darmochwał-Kolarz, D., Kludka-Sternik, M., Tabarkiewicz, J., Kolarz, B., Rolinski, J., Leszczynska-Gorzelak, B. & Oleszczuk, J. (2012). The predominance of Th17 lymphocytes and decreased number and function of Treg cells in preeclampsia. *J Reprod Immunol* **93,** 75-81.
- [576] Wenzel, K., Rajakumar, A., Haase, H., Geusens, N., Hubner, N., Schulz, H., Brewer, J., Roberts, L., Hubel, C. A., Herse, F., Hering, L., Qadri, F., Lindschau, C., Wallukat, G., Pijnenborg, R., Heidecke, H., Riemekasten, G., Luft, F. C., Muller, D. N., Lamarca, B. & Dechend, R. (2011). Angiotensin II type 1 receptor antibodies and increased angiotensin II sensitivity in pregnant rats. *Hypertension* 58, 77-84.
- [577] Stepan, H., Wallukat, G., Schultheiss, H. P., Faber, R. & Walther, T. (2007). Is parvovirus B19 the cause for autoimmunity against the angiotensin II type receptor? *J Reprod Immunol* **73**, 130-4.

- [578] Irani, R. A., Zhang, Y., Zhou, C. C., Blackwell, S. C., Hicks, M. J., Ramin, S. M., Kellems, R. E. & Xia, Y. (2010). Autoantibody-mediated angiotensin receptor activation contributes to preeclampsia through tumor necrosis factor-alpha signaling. *Hypertension* **55**, 1246-53.
- [579] Irani, R. A. & Xia, Y. (2011). Renin angiotensin signaling in normal pregnancy and preeclampsia. *Semin Nephrol* **31**, 47-58.
- [580] Mistry, H. D., Kurlak, L. O. & Broughton Pipkin, F. (2013). The placental renin-angiotensin system and oxidative stress in pre-eclampsia. *Placenta* **34**, 182-6.
- [581] Verdonk, K., Visser, W., Van Den Meiracker, A. H. & Danser, A. H. (2014). The renin-angiotensin-aldosterone system in pre-eclampsia: the delicate balance between good and bad. *Clin Sci (Lond)* **126,** 537-44.
- [582] Diveu, C., McGeachy, M. J. & Cua, D. J. (2008). Cytokines that regulate autoimmunity. *Curr Opin Immunol* **20**, 663-8.
- [583] Dong, C. (2008). Regulation and pro-inflammatory function of interleukin-17 family cytokines. *Immunol Rev* **226**, 80-6.
- [584] Miossec, P., Korn, T. & Kuchroo, V. K. (2009). Interleukin-17 and type 17 helper T cells. *N Engl J Med* **361**, 888-98.
- [585] Miossec, P. & Kolls, J. K. (2012). Targeting IL-17 and TH17 cells in chronic inflammation. *Nat Rev Drug Discov* **11**, 763-76.
- [586] Benedetti, G. & Miossec, P. (2014). Interleukin 17 contributes to the chronicity of inflammatory diseases such as rheumatoid arthritis. *Eur J Immunol* **44**, 339-47.
- [587] Annunziato, F., Cosmi, L., Liotta, F., Maggi, E. & Romagnani, S. (2008). The phenotype of human T<sub>h</sub>17 cells and their precursors, the cytokines that mediate their differentiation and the role of T<sub>h</sub>17 cells in inflammation. *Int Immunol* **20**, 1361-8.
- [588] Aujla, S. J., Dubin, P. J. & Kolls, J. K. (2007). Th17 cells and mucosal host defense. *Semin Immunol* **19**, 377-82.
- [589] Stockinger, B., Veldhoen, M. & Martin, B. (2007). Th17 T cells: linking innate and adaptive immunity. *Semin Immunol* **19**, 353-61.
- [590] Ouyang, W., Kolls, J. K. & Zheng, Y. (2008). The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity* **28**, 454-67.
- [591] Curtis, M. M. & Way, S. S. (2009). Interleukin-17 in host defence against bacterial, mycobacterial and fungal pathogens. *Immunology* **126**, 177-85.
- [592] Eyerich, K., Pennino, D., Scarponi, C., Foerster, S., Nasorri, F., Behrendt, H., Ring, J., Traidl-Hoffmann, C., Albanesi, C. & Cavani, A. (2009). IL-17 in atopic eczema: linking allergen-specific adaptive and microbial-triggered innate immune response. *J Allergy Clin Immunol* 123, 59-66 e4.
- [593] Khader, S. A., Gaffen, S. L. & Kolls, J. K. (2009). Th17 cells at the crossroads of innate and adaptive immunity against infectious diseases at the mucosa. *Mucosal Immunol* **2**, 403-11.
- [594] Ivanov, II, Zhou, L. & Littman, D. R. (2007). Transcriptional regulation of Th17 cell differentiation. Semin Immunol 19, 409-17.
- [595] Cypowyj, S., Picard, C., Marodi, L., Casanova, J. L. & Puel, A. (2012). Immunity to infection in IL-17-deficient mice and humans. *Eur J Immunol* **42**, 2246-54.
- [596] Rubino, S. J., Geddes, K. & Girardin, S. E. (2012). Innate IL-17 and IL-22 responses to enteric bacterial pathogens. *Trends Immunol* **33**, 112-8.
- [597] Weber, A., Zimmermann, C., Kieseier, B. C., Hartung, H. P. & Hofstetter, H. H. (2014). Bacteria and their cell wall components uniformly co-activate interleukin-17-producing thymocytes. *Clin Exp Immunol* **178**, 504-15.
- [598] Beringer, A., Noack, M. & Miossec, P. (2016). IL-17 in Chronic Inflammation: From Discovery to Targeting. *Trends Mol Med* **22**, 230-41.
- [599] Kumar, P., Monin, L., Castillo, P., Elsegeiny, W., Horne, W., Eddens, T., Vikram, A., Good, M., Schoenborn, A. A., Bibby, K., Montelaro, R. C., Metzger, D. W., Gulati, A. S. & Kolls, J. K. (2016). Intestinal Interleukin-17 Receptor Signaling Mediates Reciprocal Control of the Gut Microbiota and Autoimmune Inflammation. *Immunity* 44, 659-71.
- [600] Saito, S., Nakashima, A., Ito, M. & Shima, T. (2011). Clinical implication of recent advances in our understanding of IL-17 and reproductive immunology. *Expert Rev Clin Immunol* **7**, 649-57.

- [601] Fu, B., Tian, Z. & Wei, H. (2014). TH17 cells in human recurrent pregnancy loss and pre-eclampsia. *Cell Mol Immunol* **11**, 564-70.
- [602] Ozkan, Z. S., Simsek, M., Ilhan, F., Deveci, D., Godekmerdan, A. & Sapmaz, E. (2014). Plasma IL-17, IL-35, interferon-gamma, SOCS3 and TGF-beta levels in pregnant women with preeclampsia, and their relation with severity of disease. *J Matern Fetal Neonatal Med* 27, 1513-7.
- [603] Santner-Nanan, B., Peek, M. J., Khanam, R., Richarts, L., Zhu, E., Fazekas de St Groth, B. & Nanan, R. (2009). Systemic increase in the ratio between Foxp3+ and IL-17-producing CD4+ T cells in healthy pregnancy but not in preeclampsia. *J Immunol* **183**, 7023-30.
- [604] Jianjun, Z., Yali, H., Zhiqun, W., Mingming, Z. & Xia, Z. (2010). Imbalance of T-cell transcription factors contributes to the Th1 type immunity predominant in pre-eclampsia. *Am J Reprod Immunol* **63,** 38-45.
- [605] Saito, S. (2010). Th17 cells and regulatory T cells: new light on pathophysiology of preeclampsia. Immunol Cell Biol 88, 615-7.
- [606] Toldi, G., Rigó, J., Stenczer, B., Vásárhelyi, B. & Molvarec, A. (2011). Increased Prevalence of IL-17-Producing Peripheral Blood Lymphocytes in Pre-eclampsia. *Am J Reprod Immunol* **66**, 223-229.
- [607] Cornelius, D. C., Hogg, J. P., Scott, J., Wallace, K., Herse, F., Moseley, J., Wallukat, G., Dechend, R. & LaMarca, B. (2013). Administration of interleukin-17 soluble receptor C suppresses TH17 cells, oxidative stress, and hypertension in response to placental ischemia during pregnancy. *Hypertension* **62**, 1068-73.
- [608] Laresgoiti-Servitje, E. (2013). A leading role for the immune system in the pathophysiology of preeclampsia. *J Leukoc Biol* **94,** 247-57.
- [609] Perez-Sepulveda, A., Torres, M. J., Khoury, M. & Illanes, S. E. (2014). Innate immune system and preeclampsia. *Frontiers in Immunology* **5**.
- [610] Cao, W., Wang, X., Chen, T., Zhu, H., Xu, W., Zhao, S., Cheng, X. & Xia, L. (2015). The Expression of Notch/Notch Ligand, IL-35, IL-17, and Th17/Treg in Preeclampsia. *Dis Markers* **2015**, 316182.
- [611] Molvarec, A., Czegle, I., Szijártó, J. & Rigó, J. (2015). Increased circulating interleukin-17 levels in preeclampsia. *J Reprod Immunol* **112**, 53-57.
- [612] Vargas-Rojas, M. I., Solleiro-Villavicencio, H. & Soto-Vega, E. (2016). Th1, Th2, Th17 and Treg levels in umbilical cord blood in preeclampsia. *J Matern-Fetal Neo M* **29**, 1642-1645.
- [613] Wang, H., Guo, M., Liu, F., Wang, J., Zhou, Z., Ji, J., Ye, Y., Song, W., Liu, S. & Sun, B. (2015). Role of IL-17 Variants in Preeclampsia in Chinese Han Women. *PLoS One* **10**, e0140118.
- [614] Darmochwał-Kolarz, D. & Oleszczuk, J. (2014). The critical role of Th17 cells, T<sub>reg</sub> cells and costimulatory molecules in the development of pre-eclampsia. *Dev Period Med* **18**, 141-7.
- [615] Suntharalingam, G., Perry, M. R., Ward, S., Brett, S. J., Castello-Cortes, A., Brunner, M. D. & Panoskaltsis, N. (2006). Cytokine storm in a phase 1 trial of the anti-CD28 monoclonal antibody TGN1412. N Engl J Med 355, 1018-28.
- [616] Wang, H. & Ma, S. (2008). The cytokine storm and factors determining the sequence and severity of organ dysfunction in multiple organ dysfunction syndrome. *Am J Emerg Med* **26**, 711-5.
- [617] Tscherne, D. M. & García-Sastre, A. (2011). Virulence determinants of pandemic influenza viruses. *J Clin Invest* **121,** 6-13.
- [618] D'Elia, R. V., Harrison, K., Oyston, P. C., Lukaszewski, R. A. & Clark, G. C. (2013). Targeting the "cytokine storm" for therapeutic benefit. *Clin Vaccine Immunol* **20**, 319-27.
- [619] Ranieri, V. M., Thompson, B. T., Barie, P. S., Dhainaut, J. F., Douglas, I. S., Finfer, S., Gardlund, B., Marshall, J. C., Rhodes, A., Artigas, A., Payen, D., Tenhunen, J., Al-Khalidi, H. R., Thompson, V., Janes, J., Macias, W. L., Vangerow, B., Williams, M. D. & Group, P.-S. S. (2012). Drotrecogin alfa (activated) in adults with septic shock. N Engl J Med 366, 2055-64.
- [620] Lai, P. S., Matteau, A., Iddriss, A., Hawes, J. C., Ranieri, V. & Thompson, B. T. (2013). An updated metaanalysis to understand the variable efficacy of drotrecogin alfa (activated) in severe sepsis and septic shock. *Minerva Anestesiol* **79**, 33-43.
- [621] Vulcano, M., Meiss, R. P. & Isturiz, M. A. (2000). Deferoxamine reduces tissue injury and lethality in LPS-treated mice. *Int J Immunopharmacol* **22**, 635-44.

- [622] Messaris, E., Antonakis, P. T., Memos, N., Chatzigianni, E., Leandros, E. & Konstadoulakis, M. M. (2004). Deferoxamine administration in septic animals: improved survival and altered apoptotic gene expression. *Int Immunopharmacol* **4,** 455-9.
- [623] Doron, M. W., Makhlouf, R. A., Katz, V. L., Lawson, E. E. & Stiles, A. D. (1994). Increased Incidence of Sepsis at Birth in Neutropenic Infants of Mothers with Preeclampsia. *Journal of Pediatrics* **125**, 452-458.
- [624] Kocherlakota, P. & La Gamma, E. F. (1998). Preliminary report: rhG-CSF may reduce the incidence of neonatal sepsis in prolonged preeclampsia-associated neutropenia. *Pediatrics* **102**, 1107-11.
- [625] Bhaumik, S., Ghosh, S., Haldar, K. K., Mitra, P. K. & Manna, B. (2000). Risk of early onset neonatal septicemia in babies born to mothers with pre-eclampsia. *Indian Pediatr* **37**, 775-9.
- [626] Procianoy, R. S., Silveira, R. C., Mussi-Pinhata, M. M., Rugolo, L. M. S. S., Leone, C. R., Lopes, J. M. D., de Almeida, M. F. B. & Res, B. N. N. (2010). Sepsis and Neutropenia in Very Low Birth Weight Infants Delivered of Mothers with Preeclampsia. *Journal of Pediatrics* **157**, 434-U118.
- [627] Namdev, S., Bhat, V., Adhisivam, B. & Zachariah, B. (2014). Oxidative stress and antioxidant status among neonates born to mothers with pre-eclampsia and their early outcome. *J Matern Fetal Neonatal Med* 27, 1481-4.
- [628] Kenny, L. C., Dunn, W. B., Ellis, D. I., Myers, J., Baker, P. N., The GOPEC Consortium & Kell, D. B. (2005). Novel biomarkers for pre-eclampsia detected using metabolomics and machine learning. *Metabolomics* 1, 227-234 - online DOI: 10.1007/s11306-005-0003-1.
- [629] Kenny, L. C., Broadhurst, D., Brown, M., Dunn, W. B., Redman, C. W. G., Kell, D. B. & Baker, P. N. (2008). Detection and identification of novel metabolomic biomarkers in preeclampsia. *Reprod Sci* 15, 591-7.
- [630] Bahado-Singh, R. O., Akolekar, R., Mandal, R., Dong, E., Xia, J., Kruger, M., Wishart, D. S. & Nicolaides, K. (2012). Metabolomics and first-trimester prediction of early-onset preeclampsia. *J Matern Fetal Neonatal Med* 25, 1840-7.
- [631] Dunn, W. B., Brown, M., Worton, S. A., Davies, K., Jones, R. L., Kell, D. B. & Heazell, A. E. P. (2012). The metabolome of human placental tissue: investigation of first trimester tissue and changes related to preeclampsia in late pregnancy. *Metabolomics* **8**, 579-597.
- [632] Koster, M. P. H., Vreeken, R. J., Harms, A. C., Dane, A. D., Kuc, S., Schielen, P. C. J. I., Hankemeier, T., Berger, R., Visser, G. H. A. & Pennings, J. L. A. (2015). First-Trimester Serum Acylcarnitine Levels to Predict Preeclampsia: A Metabolomics Approach. *Dis Markers* **2015**, 857108.
- [633] Kuc, S., Koster, M. P. H., Pennings, J. L. A., Hankemeier, T., Berger, R., Harms, A. C., Dane, A. D., Schielen, P. C. J. I., Visser, G. H. A. & Vreeken, R. J. (2014). Metabolomics profiling for identification of novel potential markers in early prediction of preeclampsia. *PloS one* **9**, e98540.
- [634] Odibo, A. O., Goetzinger, K. R., Odibo, L., Cahill, A. G., Macones, G. A., Nelson, D. M. & Dietzen, D. J. (2011). First-trimester prediction of preeclampsia using metabolomic biomarkers: a discovery phase study. *Prenat Diagn* **31**, 990-4.
- [635] Woodham, P. C., O'Connell, T., Grimes, J., Haeri, S., Eichelberger, K., Baker, A. & Boggess, K. (2012). Metabolomics to predict severe preeclampsia in early pregnancy. *Am J Obstet Gynecol* **206**, S348-S348.
- [636] Myers, J. E., Hart, S., Armstrong, S., Mires, G. J., Beynon, R., Gaskell, S. J. & Baker, P. N. (2007). Evidence for multiple circulating factors in preeclampsia. *Am J Obstet Gynecol* **196**, 266 e1-6.
- [637] Myers, J. E., Kenny, L. C., McCowan, L. M., Chan, E. H., Dekker, G. A., Poston, L., Simpson, N. A. & North, R. A. (2013). Angiogenic factors combined with clinical risk factors to predict preterm preeclampsia in nulliparous women: a predictive test accuracy study. *BJOG* **120**, 1215-23.
- [638] Myers, J. E., Tuytten, R., Thomas, G., Laroy, W., Kas, K., Vanpoucke, G., Roberts, C. T., Kenny, L. C., Simpson, N. A., Baker, P. N. & North, R. A. (2013). Integrated proteomics pipeline yields novel biomarkers for predicting preeclampsia. *Hypertension* **61**, 1281-8.
- [639] Khan, G. H., Galazis, N., Docheva, N., Layfield, R. & Atiomo, W. (2015). Overlap of proteomics biomarkers between women with pre-eclampsia and PCOS: a systematic review and biomarker database integration. *Hum Reprod* **30**, 133-48.
- [640] Paulus, P., Jennewein, C. & Zacharowski, K. (2011). Biomarkers of endothelial dysfunction: can they help us deciphering systemic inflammation and sepsis? *Biomarkers* **16 Suppl 1,** S11-21.

- [641] Netea, M. G., Joosten, L. A. B., Latz, E., Mills, K. H. G., Natoli, G., Stunnenberg, H. G., O'Neill, L. A. J. & Xavier, R. J. (2016). Trained immunity: A program of innate immune memory in health and disease. *Science* **352**, aaf1098.
- [642] Savvidou, M. D., Lees, C. C., Parra, M., Hingorani, A. D. & Nicolaides, K. H. (2002). Levels of C-reactive protein in pregnant women who subsequently develop pre-eclampsia. *BJOG* **109**, 297-301.
- [643] Kashanian, M., Aghbali, F. & Mahali, N. (2013). Evaluation of the diagnostic value of the first-trimester maternal serum high-sensitivity C-reactive protein level for prediction of pre-eclampsia. *J Obstet Gynaecol Res* **39**, 1549-54.
- [644] Saito, S., Shiozaki, A., Nakashima, A., Sakai, M. & Sasaki, Y. (2007). The role of the immune system in preeclampsia. *Mol Aspects Med* **28**, 192-209.
- [645] Rinehart, B. K., Terrone, D. A., Lagoo-Deenadayalan, S., Barber, W. H., Hale, E. A., Martin, J. N., Jr. & Bennett, W. A. (1999). Expression of the placental cytokines tumor necrosis factor alpha, interleukin 1beta, and interleukin 10 is increased in preeclampsia. Am J Obstet Gynecol 181, 915-20.
- [646] Serin, Ý. S., Özçelik, B., Bapbuð, M., Kýlýç, H., Okur, D. & Erez, R. (2002). Predictive value of tumor necrosis factor alpha (TNF-alpha) in preeclampsia. *Eur J Obstet Gynecol Reprod Biol* **100**, 143-5.
- [647] Todros, T., Bontempo, S., Piccoli, E., Ietta, F., Romagnoli, R., Biolcati, M., Castellucci, M. & Paulesu, L. (2005). Increased levels of macrophage migration inhibitory factor (MIF) in preeclampsia. Eur J Obstet Gynecol Reprod Biol 123, 162-6.
- [648] Mehr, S. & Doyle, L. W. (2000). Cytokines as markers of bacterial sepsis in newborn infants: a review. *Pediatr Infect Dis J* **19,** 879-87.
- [649] Roger, T., Glauser, M. P. & Calandra, T. (2001). Macrophage migration inhibitory factor (MIF) modulates innate immune responses induced by endotoxin and Gram-negative bacteria. *J Endotoxin Res* **7**, 456-60.
- [650] Takala, A., Nupponen, I., Kylanpää-Bäck, M. L. & Repo, H. (2002). Markers of inflammation in sepsis. Ann Med **34**, 614-23.
- [651] Su, H., Chang, S. S., Han, C. M., Wu, K. Y., Li, M. C., Huang, C. Y., Lee, C. L., Wu, J. Y. & Lee, C. C. (2014). Inflammatory markers in cord blood or maternal serum for early detection of neonatal sepsis-a systemic review and meta-analysis. *J Perinatol* **34**, 268-74.
- [652] Briassoulis, G. & Galani, A. (2014). Prognostic markers of pediatric meningococcal sepsis. *Expert Rev Anti Infect Ther* **12**, 1017-20.
- [653] Palsson, B. Ø. (2006). Systems biology: properties of reconstructed networks. Cambridge University Press, Cambridge.
- [654] Kell, D. B. & Knowles, J. D. (2006). The role of modeling in systems biology. In *System modeling in cellular biology: from concepts to nuts and bolts* (ed. Z. Szallasi, J. Stelling and V. Periwal), pp. 3-18. MIT Press, Cambridge.
- [655] Palsson, B. Ø. (2015). Systems biology: constraint-based reconstruction and analysis. Cambridge University Press, Cambridge.
- [656] Hucka, M., Finney, A., Sauro, H. M., Bolouri, H., Doyle, J. C., Kitano, H., Arkin, A. P., Bornstein, B. J., Bray, D., Cornish-Bowden, A., Cuellar, A. A., Dronov, S., Gilles, E. D., Ginkel, M., Gor, V., Goryanin, I. I., Hedley, W. J., Hodgman, T. C., Hofmeyr, J. H., Hunter, P. J., Juty, N. S., Kasberger, J. L., Kremling, A., Kummer, U., Le Novere, N., Loew, L. M., Lucio, D., Mendes, P., Minch, E., Mjolsness, E. D., Nakayama, Y., Nelson, M. R., Nielsen, P. F., Sakurada, T., Schaff, J. C., Shapiro, B. E., Shimizu, T. S., Spence, H. D., Stelling, J., Takahashi, K., Tomita, M., Wagner, J. & Wang, J. (2003). The systems biology markup language (SBML): a medium for representation and exchange of biochemical network models. *Bioinformatics* 19, 524-31.
- [657] Funahashi, A., Matsuoka, Y., Jouraku, A., Morohashi, M., Kikuchi, N. & Kitano, H. (2008). CellDesigner 3.5: A versatile modeling tool for biochemical networks. *Proc IEEE* **96**, 1254-1265.
- [658] Hoops, S., Sahle, S., Gauges, R., Lee, C., Pahle, J., Simus, N., Singhal, M., Xu, L., Mendes, P. & Kummer, U. (2006). COPASI: a COmplex PAthway Simulator. *Bioinformatics* **22**, 3067-74.
- [659] Smoot, M. E., Ono, K., Ruscheinski, J., Wang, P. L. & Ideker, T. (2011). Cytoscape 2.8: New Features for Data Integration and Network Visualization. *Bioinformatics* 27, 431-432.

- [660] Reddy, A., Suri, S., Sargent, I. L., Redman, C. W. & Muttukrishna, S. (2009). Maternal circulating levels of activin A, inhibin A, sFlt-1 and endoglin at parturition in normal pregnancy and pre-eclampsia. *PLoS One* **4**, e4453.
- [661] Phillips, D. J., Jones, K. L., Scheerlinck, J. Y., Hedger, M. P. & de Kretser, D. M. (2001). Evidence for activin A and follistatin involvement in the systemic inflammatory response. *Mol Cell Endocrinol* **180**, 155-62.
- [662] Hodges, R., Salvador, L., D'Antona, D., Georgiou, H. M. & Wallace, E. M. (2010). Activin A as a marker of intrauterine infection in women with preterm prelabour rupture of membranes. *J Perinatol* **30**, 22-6.
- [663] Rosenberg, V. A., Buhimschi, I. A., Dulay, A. T., Abdel-Razeq, S. S., Oliver, E. A., Duzyj, C. M., Lipkind, H., Pettker, C. M. & Buhimschi, C. S. (2012). Modulation of amniotic fluid activin-A and inhibin-A in women with preterm premature rupture of the membranes and infection-induced preterm birth. *Am J Reprod Immunol* **67**, 122-31.
- [664] Petrakou, E., Fotopoulos, S., Anagnostakou, M., Anatolitou, F., Samitas, K., Semitekolou, M., Xanthou, G. & Xanthou, M. (2013). Activin-A exerts a crucial anti-inflammatory role in neonatal infections. Pediatr Res 74, 675-81.
- [665] Shu, C., Liu, Z., Cui, L., Wei, C., Wang, S., Tang, J. J., Cui, M., Lian, G., Li, W., Liu, X., Xu, H., Jiang, J., Lee, P., Zhang, D. Y., He, J. & Ye, F. (2014). Protein profiling of preeclampsia placental tissues. *PLoS One* **9**, e112890.
- [666] Lugli, A., Forster, Y., Haas, P., Nocito, A., Bucher, C., Bissig, H., Mirlacher, M., Storz, M., Mihatsch, M. J. & Sauter, G. (2003). Calretinin expression in human normal and neoplastic tissues: a tissue microarray analysis on 5233 tissue samples. *Hum Pathol* 34, 994-1000.
- [667] Rizzo, A., Carratelli, C. R., De Filippis, A., Bevilacqua, N., Tufano, M. A. & Buommino, E. (2014). Transforming activities of *Chlamydia pneumoniae* in human mesothelial cells. *Int Microbiol* 17, 185-193.
- [668] Xu, Q. L., Zhu, M., Jin, Y., Wang, N., Xu, H. X., Quan, L. M., Wang, S. S. & Li, S. S. (2014). The predictive value of the first-trimester maternal serum chemerin level for pre-eclampsia. *Peptides* **62**, 150-4.
- [669] Stepan, H., Philipp, A., Roth, I., Kralisch, S., Jank, A., Schaarschmidt, W., Lössner, U., Kratzsch, J., Blüher, M., Stumvoll, M. & Fasshauer, M. (2011). Serum levels of the adipokine chemerin are increased in preeclampsia during and 6 months after pregnancy. *Regulatory Peptides* **168**, 69-72.
- [670] AL-Refai, A. A. (2012). Evaluation of Serum Levels of the Adipokines Chemerin and Resistin in Preeclampsia. *Life Sci J* **9**, 5143-5151.
- [671] Duan, D. M., Niu, J. M., Lei, Q., Lin, X. H. & Chen, X. (2012). Serum levels of the adipokine chemerin in preeclampsia. *J Perinat Med* **40**, 121-127.
- [672] Wang, L. Q., Yang, T. L., Ding, Y. L., Zhong, Y., Yu, L. & Peng, M. (2015). Chemerin plays a protective role by regulating human umbilical vein endothelial cell-induced nitric oxide signaling in preeclampsia. *Endocrine* **48**, 299-308.
- [673] Broadhurst, D. & Kell, D. B. (2006). Statistical strategies for avoiding false discoveries in metabolomics and related experiments. *Metabolomics* **2**, 171-196.
- [674] Kukla, M., Zwirska-Korczala, K., Gabriel, A., Waluga, M., Warakomska, I., Szczygiel, B., Berdowska, A., Mazur, W., Wozniak-Grygiel, E. & Kryczka, W. (2010). Chemerin, vaspin and insulin resistance in chronic hepatitis C. *J Viral Hepat* 17, 661-667.
- [675] Kukla, M., Mazur, W., Buldak, R. J. & Żwirska-Korczala, K. (2011). Potential Role of Leptin, Adiponectin and Three Novel Adipokines-Visfatin, Chemerin and Vaspin-in Chronic Hepatitis. *Mol Med* 17, 1397-1410.
- [676] Kulig, P., Kantyka, T., Zabel, B. A., Banas, M., Chyra, A., Stefanska, A., Tu, H., Allen, S. J., Handel, T. M., Kozik, A., Potempa, J., Butcher, E. C. & Cichy, J. (2011). Regulation of Chemerin Chemoattractant and Antibacterial Activity by Human Cysteine Cathepsins. *J Immunol* **187**, 1403-1410.
- [677] Banas, M., Zabieglo, K., Kasetty, G., Kapinska-Mrowiecka, M., Borowczyk, J., Drukala, J., Murzyn, K., Zabel, B. A., Butcher, E. C., Schroeder, J. M., Schmidtchen, A. & Cichy, J. (2013). Chemerin is an antimicrobial agent in human epidermis. *PLoS One* **8**, e58709.
- [678] Zabel, B. A., Kwitniewski, M., Banas, M., Zabieglo, K., Murzyn, K. & Cichy, J. (2014). Chemerin regulation and role in host defense. *Am J Clin Exp Immunol* **3**, 1-19.

- [679] Banas, M., Zegar, A., Kwitniewski, M., Zabieglo, K., Marczynska, J., Kapinska-Mrowiecka, M., LaJevic, M., Zabel, B. A. & Cichy, J. (2015). The expression and regulation of chemerin in the epidermis. *Plos One* 10.
- [680] Horn, P., Metzing, U. B., Steidl, R., Romeike, B., Rauchfuß, F., Sponholz, C., Thomas-Rüddel, D., Ludewig, K., Birkenfeld, A. L., Settmacher, U., Bauer, M., Claus, R. A. & von Loeffelholz, C. (2016). Chemerin in peritoneal sepsis and its associations with glucose metabolism and prognosis: a translational cross-sectional study. *Crit Care* 20.
- [681] Strevens, H., Wide-Swensson, D., Grubb, A., Hansen, A., Horn, T., Ingemarsson, I., Larsen, S., Nyengaard, J. R., Torffvit, O., Willner, J. & Olsen, S. (2003). Serum cystatin C reflects glomerular endotheliosis in normal, hypertensive and pre-eclamptic pregnancies. *BJOG* **110**, 825-30.
- [682] Yang, X., Wang, H., Wang, Z. & Dong, M. (2006). Alteration and significance of serum cardiac troponin I and cystatin C in preeclampsia. *Clin Chim Acta* **374**, 168-9.
- [683] Kristensen, K., Wide-Swensson, D., Schmidt, C., Blirup-Jensen, S., Lindström, V., Strevens, H. & Grubb, A. (2007). Cystatin C, beta-2-microglobulin and beta-trace protein in pre-eclampsia. Acta Obstet Gynecol Scand 86, 921-6.
- [684] Guo, H. X., Wang, C. H., Li, Z. Q., Gong, S. P., Zhou, Z. Q., Leng, L. Z. & Zhong, M. (2012). The application of serum cystatin C in estimating the renal function in women with preeclampsia. *Reprod Sci* **19**, 712-7.
- [685] Novakov Mikic, A., Cabarkapa, V., Nikolic, A., Maric, D., Brkic, S., Mitic, G., Ristic, M. & Stosic, Z. (2012). Cystatin C in pre-eclampsia. *J Matern Fetal Neonatal Med* **25**, 961-5.
- [686] Xiao, J., Niu, J., Ye, X., Yu, Q. & Gu, Y. (2013). Combined biomarkers evaluation for diagnosing kidney injury in preeclampsia. *Hypertens Pregnancy*.
- [687] Yalcin, S., Ulas, T., Eren, M. A., Aydogan, H., Camuzcuoglu, A., Kucuk, A., Yuce, H. H., Demir, M. E., Vural, M. & Aksoy, N. (2013). Relationship between oxidative stress parameters and cystatin C levels in patients with severe preeclampsia. *Medicina (Kaunas)* 49, 118-23.
- [688] Odden, M. C., Scherzer, R., Bacchetti, P., Szczech, L. A., Sidney, S., Grunfeld, C. & Shlipak, M. G. (2007). Cystatin C level as a marker of kidney function in human immunodeficiency virus infection: the FRAM study. *Arch Intern Med* **167**, 2213-9.
- [689] Randers, E., Kornerup, K., Erlandsen, E. J., Hasling, C. & Danielsen, H. (2001). Cystatin C levels in sera of patients with acute infectious diseases with high C-reactive protein levels. *Scand J Clin Lab Invest* **61**, 333-5.
- [690] Gupta, S. K., Kitch, D., Tierney, C., Melbourne, K., Ha, B., McComsey, G. A. & Team, A. C. T. G. S. A. (2015). Markers of renal disease and function are associated with systemic inflammation in HIV infection. HIV Med 16, 591-8.
- [691] Walker, J. B. & Nesheim, M. E. (1999). The molecular weights, mass distribution, chain composition, and structure of soluble fibrin degradation products released from a fibrin clot perfused with plasmin. *J Biol Chem* **274**, 5201-12.
- [692] Bellart, J., Gilabert, R., Anglès, A., Piera, V., Miralles, R. M., Monasterio, J. & Cabero, L. (1999). Tissue factor levels and high ratio of fibrinopeptide A:D-dimer as a measure of endothelial procoagulant disorder in pre-eclampsia. *Br J Obstet Gynaecol* **106**, 594-7.
- [693] Gulec, U. K., Ozgunen, F. T., Guzel, A. B., Buyukkurt, S., Seydaoglu, G., Urunsak, I. F. & Evruke, I. C. (2012). An analysis of C-reactive protein, procalcitonin, and D-dimer in pre-eclamptic patients. Am J Reprod Immunol 68, 331-7.
- [694] Pinheiro, M. d. B., Junqueira, D. R. G., Coelho, F. F., Freitas, L. G., Carvalho, M. G., Gomes, K. B. & Dusse, L. M. S. (2012). D-dimer in preeclampsia: systematic review and meta-analysis. *Clin Chim Acta* **414**, 166-70.
- [695] Bozkurt, M., Yumru, A. E., Sahin, L. & Salman, S. (2015). Troponin I and D-Dimer levels in preeclampsia and eclampsia: prospective study. *Clin Exp Obstet Gynecol* **42**, 26-31.
- [696] Rahman, R., Begum, K., Khondker, L., Majumder, N. I., Nahar, K., Sultana, R. & Siddika, A. (2015). Role of D-dimer in determining coagulability status in pre-eclamptic and normotensive pregnant women. *Mymensingh Med J* **24**, 115-20.
- [697] Di Castelnuovo, A., de Curtis, A., Costanzo, S., Persichillo, M., Olivieri, M., Zito, F., Donati, M. B., de Gaetano, G., Iacoviello, L. & MOLI-SANI Project Investigators. (2013). Association of D-dimer levels

- with all-cause mortality in a healthy adult population: findings from the MOLI-SANI study. *Haematologica* **98**, 1476-80.
- [698] Jennings, I., Woods, T. A. L., Kitchen, D. P., Kitchen, S. & Walker, I. D. (2007). Laboratory D-dimer measurement: improved agreement between methods through calibration. *Thromb Haemost* 98, 1127-35.
- [699] Khalafallah, A. A., Morse, M., Al-Barzan, A. M., Adams, M., Dennis, A., Bates, G., Robertson, I., Seaton, D. & Brain, T. (2012). D-Dimer levels at different stages of pregnancy in Australian women: a single centre study using two different immunoturbidimetric assays. *Thromb Res* **130**, e171-7.
- [700] Rodelo, J. R., De la Rosa, G., Valencia, M. L., Ospina, S., Arango, C. M., Gómez, C. I., García, A., Nuñez, E. & Jaimes, F. A. (2012). D-dimer is a significant prognostic factor in patients with suspected infection and sepsis. *Am J Emerg Med* **30**, 1991-9.
- [701] Khalafallah, A., Jarvis, C., Morse, M., Albarzan, A. M., Stewart, P., Bates, G., Hayes, R., Robertson, I., Seaton, D. & Brain, T. (2014). Evaluation of the innovance d-dimer assay for the diagnosis of disseminated intravascular coagulopathy in different clinical settings. *Clin Appl Thromb Hemost* 20, 91-7.
- [702] Shovlin, C. L., Hughes, J. M. B., Scott, J., Seidman, C. E. & Seidman, J. G. (1997). Characterization of endoglin and identification of novel mutations in hereditary hemorrhagic telangiectasia. *American Journal of Human Genetics* **61**, 68-79.
- [703] Shovlin, C. L. (2010). Hereditary haemorrhagic telangiectasia: pathophysiology, diagnosis and treatment. *Blood Rev* **24**, 203-19.
- [704] Gregory, A. L., Xu, G., Sotov, V. & Letarte, M. (2014). Review: the enigmatic role of endoglin in the placenta. *Placenta* **35 Suppl**, S93-9.
- [705] Bell, M. J., Roberts, J. M., Founds, S. A., Jeyabalan, A., Terhorst, L. & Conley, Y. P. (2013). Variation in endoglin pathway genes is associated with preeclampsia: a case-control candidate gene association study. *BMC Pregnancy and Childbirth* **13**.
- [706] Venkatesha, S., Toporsian, M., Lam, C., Hanai, J., Mammoto, T., Kim, Y. M., Bdolah, Y., Lim, K. H., Yuan, H. T., Libermann, T. A., Stillman, I. E., Roberts, D., D'Amore, P. A., Epstein, F. H., Sellke, F. W., Romero, R., Sukhatme, V. P., Letarte, M. & Karumanchi, S. A. (2006). Soluble endoglin contributes to the pathogenesis of preeclampsia. *Nature Medicine* 12, 642-649.
- [707] Luft, F. C. (2006). Soluble endoglin (sEng) joins the soluble fms-like tyrosine kinase (sFlt) receptor as a pre-eclampsia molecule. *Nephrol Dial Transplant* **21**, 3052-4.
- [708] Govender, N., Moodley, J., Gathiram, P. & Naicker, T. (2014). Soluble fms-like tyrosine kinase-1 in HIV infected pre-eclamptic South African Black women. *Placenta* **35**, 618-624.
- [709] Pratt, A., Da Silva Costa, F., Borg, A. J., Kalionis, B., Keogh, R. & Murthi, P. (2015). Placenta-derived angiogenic proteins and their contribution to the pathogenesis of preeclampsia. *Angiogenesis* **18**, 115-23.
- [710] Kleinrouweler, C. E., Wiegerinck, M. M. J., Ris-Stalpers, C., Bossuyt, P. M. M., van der Post, J. A. M., von Dadelszen, P., Mol, B. W. J., Pajkrt, E. & EBM Connect Collaboration. (2012). Accuracy of circulating placental growth factor, vascular endothelial growth factor, soluble fms-like tyrosine kinase 1 and soluble endoglin in the prediction of pre-eclampsia: a systematic review and meta-analysis. *BJOG* **119**, 778-87.
- [711] Muenzner, P., Rohde, M., Kneitz, S. & Hauck, C. R. (2005). CEACAM engagement by human pathogens enhances cell adhesion and counteracts bacteria-induced detachment of epithelial cells. *J Cell Biol* **170**, 825-36.
- [712] Clemente, M., Núñez, O., Lorente, R., Rincón, D., Matilla, A., Salcedo, M., Catalina, M. V., Ripoll, C., Iacono, O. L., Bañares, R., Clemente, G. & García-Monzón, C. (2006). Increased intrahepatic and circulating levels of endoglin, a TGF-beta 1 co-receptor, in patients with chronic hepatitis C virus infection: relationship to histological and serum markers of hepatic fibrosis. *J Viral Hepat* 13, 625-632.
- [713] Silver, K. L., Conroy, A. L., Leke, R. G., Leke, R. J., Gwanmesia, P., Molyneux, M. E., Taylor, D. W., Rogerson, S. J. & Kain, K. C. (2011). Circulating soluble endoglin levels in pregnant women in Cameroon and Malawi--associations with placental malaria and fetal growth restriction. *PLoS One* **6**, e24985.

- [714] Sasmito, S. D., Ulfiati, A., Wardana, A., Nugraheni, F., Pradiptasari, N. F., Zulaifa, Z., Norahmawati, E., Sardjono, T. W. & Fitri, L. E. (2015). Endoglin Expression and The Level of TGF-β are Increased in The Placental Tissue and Correlated with Low Fetal Weight in Malaria Infected Mice. *J Trop Life Sci* **5**, 1-7.
- [715] Jansen, P., Mumme, T., Randau, T., Gravius, S. & Hermanns-Sachweh, B. (2014). Endoglin (CD105) expression differentiates between aseptic loosening and periprosthetic joint infection after total joint arthroplasty. *SpringerPlus* **3**.
- [716] Valerio, L. G. (2007). Mammalian iron metabolism. *Toxicology Mechanisms and Methods* **17**, 497-517.
- [717] Andrews, N. C. (2008). Forging a field: the golden age of iron biology. Blood 112, 219-30.
- [718] Chifman, J., Kniss, A., Neupane, P., Williams, I., Leung, B., Deng, Z., Mendes, P., Hower, V., Torti, F. M., Akman, S. A., Torti, S. V. & Laubenbacher, R. (2012). The core control system of intracellular iron homeostasis: A mathematical model. *J Theor Biol* **300**, 91-9.
- [719] Hower, V., Mendes, P., Torti, F. M., Laubenbacher, R., Akman, S., Shulaev, V. & Torti, S. V. (2009). A general map of iron metabolism and tissue-specific subnetworks. . *Mol Biosyst* **5**, 422-443.
- [720] Mitchell, S. & Mendes, P. (2013). A Computational Model of Liver Iron Metabolism. *PLoS Comp Biol* **9**, e1003299.
- [721] Oliveira, F., Rocha, S. & Fernandes, R. (2014). Iron metabolism: from health to disease. *J Clin Lab Anal* **28,** 210-8.
- [722] Kell, D. B. & Pretorius, E. (2015). Interpreting raised serum ferritin levels. BMJ 2015, 351.
- [723] Nielsen, P., Günther, U., Dürken, M., Fischer, R. & Düllmann, J. (2000). Serum ferritin iron in iron overload and liver damage: Correlation to body iron stores and diagnostic relevance. *J Lab Clin Med* **135,** 413-418.
- [724] Watanabe, K., Yamashita, Y., Ohgawara, K., Sekiguchi, M., Satake, N., Orino, K. & Yamamoto, S. (2001). Iron content of rat serum ferritin. *J Vet Med Sci* **63**, 587-589.
- [725] Yamanishi, H., Iyama, S., Yamaguchi, Y., Kanakura, Y. & Iwatani, Y. (2002). Relation between iron content of serum ferritin and clinical status factors extracted by factor analysis in patients with hyperferritinemia. *Clin Biochem* **35**, 523-9.
- [726] Konz, T., Añón Alvarez, E., Montes-Bayon, M. & Sanz-Medel, A. (2013). Antibody labeling and elemental mass spectrometry (inductively coupled plasma-mass spectrometry) using isotope dilution for highly sensitive ferritin determination and iron-ferritin ratio measurements. *Anal Chem* **85**, 8334-40.
- [727] Entman, S. S., Richardson, L. D. & Killam, A. P. (1983). Altered ferrokinetics in toxemia of pregnancy a possible indicator of decreased red cell survival. *Clin Exp Hypertens B* **2**, 171-178.
- [728] Hubel, C. A., Bodnar, L. M., Many, A., Harger, G., Ness, R. B. & Roberts, J. M. (2004). Nonglycosylated ferritin predominates in the circulation of women with preeclampsia but not intrauterine growth restriction. *Clin Chem* **50**, 948-951.
- [729] Visser, A. & van de Vyver, A. (2011). Severe hyperferritinemia in *Mycobacteria tuberculosis* infection. *Clin Infect Dis* **52**, 273-4.
- [730] Ishida, J. H. & Johansen, K. L. (2014). Iron and infection in hemodialysis patients. Semin Dial 27, 26-36.
- [731] Esteller, M. (2011). Non-coding RNAs in human disease. Nat Rev Genet 12, 861-74.
- [732] Kozomara, A. & Griffiths-Jones, S. (2014). miRBase: annotating high confidence microRNAs using deep sequencing data. *Nucleic Acids Res* **42**, D68-73.
- [733] Goulart, L. F., Bettella, F., Sønderby, I. E., Schork, A. J., Thompson, W. K., Mattingsdal, M., Steen, V. M., Zuber, V., Wang, Y., Dale, A. M., PRACTICAL/ELLIPSE consortium, Andreassen, O. A. & Djurovic, S. (2015). MicroRNAs enrichment in GWAS of complex human phenotypes. BMC Genomics 16, 304.
- [734] Londin, E., Loher, P., Telonis, A. G., Quann, K., Clark, P., Jing, Y., Hatzimichael, E., Kirino, Y., Honda, S., Lally, M., Ramratnam, B., Comstock, C. E., Knudsen, K. E., Gomella, L., Spaeth, G. L., Hark, L., Katz, L. J., Witkiewicz, A., Rostami, A., Jimenez, S. A., Hollingsworth, M. A., Yeh, J. J., Shaw, C. A., McKenzie, S. E., Bray, P., Nelson, P. T., Zupo, S., Van Roosbroeck, K., Keating, M. J., Calin, G. A., Yeo, C., Jimbo, M., Cozzitorto, J., Brody, J. R., Delgrosso, K., Mattick, J. S., Fortina, P. & Rigoutsos, I. (2015). Analysis of 13 cell types reveals evidence for the expression of numerous novel primate- and tissue-specific microRNAs. *Proc Natl Acad Sci U S A* 112, E1106-15.

- [735] Luo, S. S., Ishibashi, O., Ishikawa, G., Ishikawa, T., Katayama, A., Mishima, T., Takizawa, T., Shigihara, T., Goto, T., Izumi, A., Ohkuchi, A., Matsubara, S., Takeshita, T. & Takizawa, T. (2009). Human villous trophoblasts express and secrete placenta-specific microRNAs into maternal circulation via exosomes. *Biol Reprod* 81, 717-29.
- [736] Ouyang, Y., Mouillet, J. F., Coyne, C. B. & Sadovsky, Y. (2014). Review: placenta-specific microRNAs in exosomes good things come in nano-packages. *Placenta* **35 Suppl**, S69-73.
- [737] Record, M. (2014). Intercellular communication by exosomes in placenta: a possible role in cell fusion? *Placenta* **35**, 297-302.
- [738] Miura, K., Miura, S., Yamasaki, K., Higashijima, A., Kinoshita, A., Yoshiura, K. & Masuzaki, H. (2010). Identification of pregnancy-associated microRNAs in maternal plasma. *Clin Chem* **56**, 1767-71.
- [739] Morales Prieto, D. M. & Markert, U. R. (2011). MicroRNAs in pregnancy. J Reprod Immunol 88, 106-11.
- [740] Mouillet, J. F., Ouyang, Y., Coyne, C. B. & Sadovsky, Y. (2015). MicroRNAs in placental health and disease. *Am J Obstet Gynecol* **213**, S163-72.
- [741] Jairajpuri, D. S. & Almawi, W. Y. (2016). MicroRNA expression pattern in pre-eclampsia (Review). *Molecular Medicine Reports* **13**, 2351-2358.
- [742] Kotlabova, K., Doucha, J. & Hromadnikova, I. (2011). Placental-specific microRNA in maternal circulation--identification of appropriate pregnancy-associated microRNAs with diagnostic potential. *J Reprod Immunol* **89**, 185-91.
- [743] Zhao, Z., Moley, K. H. & Gronowski, A. M. (2013). Diagnostic potential for miRNAs as biomarkers for pregnancy-specific diseases. *Clin Biochem* **46**, 953-60.
- [744] Tsochandaridis, M., Nasca, L., Toga, C. & Levy-Mozziconacci, A. (2015). Circulating microRNAs as clinical biomarkers in the predictions of pregnancy complications. *Biomed Res Int* **2015**, 294954.
- [745] Zhu, X. M., Han, T., Sargent, I. L., Yin, G. W. & Yao, Y. Q. (2009). Differential expression profile of microRNAs in human placentas from preeclamptic pregnancies vs normal pregnancies. *Am J Obstet Gynecol* **200**, 661 e1-7.
- [746] Wang, D., Song, W. & Na, Q. (2012). The emerging roles of placenta-specific microRNAs in regulating trophoblast proliferation during the first trimester. *Aust N Z J Obstet Gynaecol* **52**, 565-70.
- [747] Chen, D. B. & Wang, W. (2013). Human placental microRNAs and preeclampsia. Biol Reprod 88, 130.
- [748] Dong, F., Zhang, Y., Xia, F., Yang, Y., Xiong, S., Jin, L. & Zhang, J. (2014). Genome-wide miRNA profiling of villus and decidua of recurrent spontaneous abortion patients. *Reproduction* **148**, 33-41.
- [749] Hromadnikova, I., Kotlabova, K., Hympanova, L., Doucha, J. & Krofta, L. (2014). First trimester screening of circulating C19MC microRNAs can predict subsequent onset of gestational hypertension. *PLoS One* **9**, e113735.
- [750] Anton, L., Olarerin-George, A. O., Hogenesch, J. B. & Elovitz, M. A. (2015). Placental expression of miR-517a/b and miR-517c contributes to trophoblast dysfunction and preeclampsia. *PLoS One* **10**, e0122707.
- [751] Yang, S., Li, H., Ge, Q., Guo, L. & Chen, F. (2015). Deregulated microRNA species in the plasma and placenta of patients with preeclampsia. *Mol Med Rep* **12**, 527-34.
- [752] Ura, B., Feriotto, G., Monasta, L., Bilel, S., Zweyer, M. & Celeghini, C. (2014). Potential role of circulating microRNAs as early markers of preeclampsia. *Taiwan J Obstet Gynecol* **53**, 232-4.
- [753] Bortolin-Cavaillé, M. L., Dance, M., Weber, M. & Cavaillé, J. (2009). C19MC microRNAs are processed from introns of large Pol-II, non-protein-coding transcripts. *Nucleic Acids Res* **37**, 3464-73.
- [754] Morales-Prieto, D. M., Ospina-Prieto, S., Chaiwangyen, W., Schoenleben, M. & Markert, U. R. (2013). Pregnancy-associated miRNA-clusters. *J Reprod Immunol* **97**, 51-61.
- [755] Delorme-Axford, E., Donker, R. B., Mouillet, J. F., Chu, T., Bayer, A., Ouyang, Y., Wang, T., Stolz, D. B., Sarkar, S. N., Morelli, A. E., Sadovsky, Y. & Coyne, C. B. (2013). Human placental trophoblasts confer viral resistance to recipient cells. *Proc Natl Acad Sci U S A* 110, 12048-53.
- [756] Bayer, A., Delorme-Axford, E., Sleigher, C., Frey, T. K., Trobaugh, D. W., Klimstra, W. B., Emert-Sedlak, L. A., Smithgall, T. E., Kinchington, P. R., Vadia, S., Seveau, S., Boyle, J. P., Coyne, C. B. & Sadovsky, Y. (2015). Human trophoblasts confer resistance to viruses implicated in perinatal infection. *Am J Obstet Gynecol* **212**, 71 e1-8.
- [757] Olarerin-George, A. O., Anton, L., Hwang, Y. C., Elovitz, M. A. & Hogenesch, J. B. (2013). A functional genomics screen for microRNA regulators of NF-kappaB signaling. *BMC Biol* **11**, 19.

- [758] Song, G. Y., Song, W. W., Han, Y., Wang, D. & Na, Q. (2013). Characterization of the role of microRNA-517a expression in low birth weight infants. *J Dev Orig Health Dis* **4,** 522-6.
- [759] Mouillet, J. F., Ouyang, Y., Bayer, A., Coyne, C. B. & Sadovsky, Y. (2014). The role of trophoblastic microRNAs in placental viral infection. *Int J Dev Biol* **58**, 281-9.
- [760] Moro, L., Bardaji, A., Macete, E., Barrios, D., Morales-Prieto, D. M., Espana, C., Mandomando, I., Sigauque, B., Dobano, C., Markert, U. R., Benitez-Ribas, D., Alonso, P. L., Menendez, C. & Mayor, A. (2016). Placental Microparticles and MicroRNAs in Pregnant Women with *Plasmodium falciparum* or HIV Infection. *PLoS One* 11, e0146361.
- [761] Khatun, S., Kanayama, N., Belayet, H. M., Bhuiyan, A. B., Jahan, S., Begum, A., Kobayashi, T. & Terao, T. (2000). Increased concentrations of plasma neuropeptide Y in patients with eclampsia and preeclampsia. *Am J Obstet Gynecol* **182**, 896-900.
- [762] Hauser, G. J., Dayao, E. K. & Zukowska-Grojec, Z. (1995). Effect of neuropeptide Y on endotoxin-induced suppression of the response to various agonists in conscious rats. *Life Sci* **57**, 235-44.
- [763] Kuncová, J., Sýkora, R., Chvojka, J., Švíglerová, J., Štengl, M., Kroužecký, A., Nalos, L. & Matějovic, M. (2011). Plasma and tissue levels of neuropeptide y in experimental septic shock: relation to hemodynamics, inflammation, oxidative stress, and hemofiltration. *Artif Organs* **35**, 625-33.
- [764] Goetz, D. H., Holmes, M. A., Borregaard, N., Bluhm, M. E., Raymond, K. N. & Strong, R. K. (2002). The neutrophil lipocalin NGAL is a bacteriostatic agent that interferes with siderophore-mediated iron acquisition. *Mol Cell* **10**, 1033-43.
- [765] Bachman, M. A., Lenio, S., Schmidt, L., Oyler, J. E. & Weiser, J. N. (2012). Interaction of lipocalin 2, transferrin, and siderophores determines the replicative niche of *Klebsiella pneumoniae* during pneumonia. *mBio* **3**.
- [766] Flo, T. H., Smith, K. D., Sato, S., Rodriguez, D. J., Holmes, M. A., Strong, R. K., Akira, S. & Aderem, A. (2004). Lipocalin 2 mediates an innate immune response to bacterial infection by sequestrating iron. *Nature* **432**, 917-21.
- [767] Chakraborty, S., Kaur, S., Guha, S. & Batra, S. K. (2012). The multifaceted roles of neutrophil gelatinase associated lipocalin (NGAL) in inflammation and cancer. *Biochim Biophys Acta* **1826,** 129-169
- [768] Cemgil Arikan, D., Ozkaya, M., Adali, E., Kilinc, M., Coskun, A., Ozer, A. & Bilge, F. (2010). Plasma lipocalin-2 levels in pregnant women with pre-eclampsia, and their relation with severity of disease. *J Matern Fetal Neonatal Med*.
- [769] D'Anna, R., Baviera, G., Giordano, D., Todarello, G., Corrado, F. & Buemi, M. (2008). Second trimester neutrophil gelatinase-associated lipocalin as a potential prediagnostic marker of preeclampsia. *Acta Obstet Gynecol Scand*, 1-4.
- [770] D'Anna, R., Baviera, G., Giordano, D., Russo, S., Dugo, N., Santamaria, A. & Corrado, F. (2009). First trimester serum PAPP-A and NGAL in the prediction of late-onset pre-eclampsia. *Prenat Diagn* **29**, 1066-8.
- [771] D'Anna, R., Baviera, G., Giordano, D., Todarello, G., Russo, S., Recupero, S., Bolignano, D. & Corrado, F. (2010). Neutrophil gelatinase-associated lipocalin serum evaluation through normal pregnancy and in pregnancies complicated by preeclampsia. *Acta Obstet Gynecol Scand* **89**, 275-8.
- [772] Stepan, H., Philipp, A., Reiche, M., Klostermann, K., Schrey, S., Reisenbuchler, C., Lossner, U., Kratzsch, J., Bluher, M., Stumvoll, M. & Fasshauer, M. (2010). Serum levels of the adipokine lipocalin-2 are increased in preeclampsia. *J Endocrinol Invest* **33**, 629-32.
- [773] Kim, S. M., Park, J. S., Norwitz, E. R., Jung, H. J., Kim, B. J., Park, C. W. & Jun, J. K. (2013). Circulating Levels of Neutrophil Gelatinase-Associated Lipocalin (NGAL) Correlate With the Presence and Severity of Preeclampsia. *Reprod Sci* **20**, 1083-1089.
- [774] Scazzochio, E., Munmany, M., Garcia, L., Meler, E., Crispi, F., Gratacos, E. & Figueras, F. (2013). Prognostic Role of Maternal Neutrophil Gelatinase-Associated Lipocalin in Women with Severe Early-Onset Preeclampsia. *Fetal Diagn Ther* **35**, 127-132.
- [775] Karampas, G., Eleftheriades, M., Panoulis, K., Rizou, M., Haliassos, A., Hassiakos, D., Vitoratos, N. & Rizos, D. (2014). Maternal serum levels of Neutrophil Gelatinase-Associated Lipocalin (NGAL), Matrix Metalloproteinase-9 (MMP-9) and their complex MMP-9/NGAL in pregnancies with

- preeclampsia and those with a small for gestational age neonate. A longitudinal study. *Prenat Diagn* **34**, 726-733.
- [776] Sachan, R., Patel, M., Gaurav, A., Gangwar, R. & Sachan, P. (2014). Correlation of serum neutrophil gelatinase associated lipocalin with disease severity in hypertensive disorders of pregnancy. *Adv Biomed Res* **3**, 223.
- [777] Simonazzi, G., Capelli, I., Curti, A., Comai, G., Rizzo, N. & G, L. A. M. (2015). Serum and Urinary Neutrophil Gelatinase-associated Lipocalin Monitoring in Normal Pregnancy Versus Pregnancies Complicated by Pre-eclampsia. *In Vivo* **29**, 117-121.
- [778] Mishra, J., Dent, C., Tarabishi, R., Mitsnefes, M. M., Ma, Q., Kelly, C., Ruff, S. M., Zahedi, K., Shao, M., Bean, J., Mori, K., Barasch, J. & Devarajan, P. (2005). Neutrophil gelatinase-associated lipocalin (NGAL) as a biomarker for acute renal injury after cardiac surgery. *Lancet* **365**, 1231-8.
- [779] Soni, S. S., Cruz, D., Bobek, I., Chionh, C. Y., Nalesso, F., Lentini, P., de Cal, M., Corradi, V., Virzi, G. & Ronco, C. (2010). NGAL: a biomarker of acute kidney injury and other systemic conditions. *Int Urol Nephrol* **42**, 141-50.
- [780] Clerico, A., Galli, C., Fortunato, A. & Ronco, C. (2012). Neutrophil gelatinase-associated lipocalin (NGAL) as biomarker of acute kidney injury: a review of the laboratory characteristics and clinical evidences. *Clin Chem Lab Med* **50**, 1505-17.
- [781] Patel, M., Sachan, R., Gangwar, R., Sachan, P. & Natu, S. (2013). Correlation of serum neutrophil gelatinase-associated lipocalin with acute kidney injury in hypertensive disorders of pregnancy. *Int J Nephrol Renovasc Dis* **6**, 181-6.
- [782] Aydoğdu, M., Gürsel, G., Sancak, B., Yeni, S., Sari, G., Taşyürek, S., Türk, M., Yüksel, S., Senes, M. & Özis, T. N. (2013). The use of plasma and urine neutrophil gelatinase associated lipocalin (NGAL) and Cystatin C in early diagnosis of septic acute kidney injury in critically ill patients. *Dis Markers* 34, 237-46
- [783] Otto, G. P., Busch, M., Sossdorf, M. & Claus, R. A. (2013). Impact of sepsis-associated cytokine storm on plasma NGAL during acute kidney injury in a model of polymicrobial sepsis. *Crit care* **17**, 419.
- [784] Mårtensson, J., Bell, M., Xu, S., Bottai, M., Ravn, B., Venge, P. & Martling, C. R. (2013). Association of plasma neutrophil gelatinase-associated lipocalin (NGAL) with sepsis and acute kidney dysfunction. *Biomarkers*.
- [785] Zughaier, S. M., Tangpricha, V., Leong, T., Stecenko, A. A. & McCarty, N. A. (2013). Peripheral monocytes derived from patients with cystic fibrosis and healthy donors secrete NGAL in response to *Pseudomonas aeruginosa* infection. *J Investig Med* **61**, 1018-25.
- [786] Smertka, M., Wroblewska, J., Suchojad, A., Majcherczyk, M., Jadamus-Niebroj, D., Owsianka-Podlesny, T., Brzozowska, A. & Maruniak-Chudek, I. (2014). Serum and Urinary NGAL in Septic Newborns. *BioMed Res Internat* **2014**, 717318.
- [787] Macdonald, S. P. J., Stone, S. F., Neil, C. L., van Eeden, P. E., Fatovich, D. M., Arendts, G. & Brown, S. G. (2014). Sustained Elevation of Resistin, NGAL and IL-8 Are Associated with Severe Sepsis/Septic Shock in the Emergency Department. *PLoS One* **9**, e110678.
- [788] Hjortrup, P. B., Haase, N., Treschow, F., Møller, M. H. & Perner, A. (2015). Predictive value of NGAL for use of renal replacement therapy in patients with severe sepsis. *Acta Anaesthesiol Scand* **59**, 25-34.
- [789] Suchojad, A., Tarko, A., Smertka, M., Majcherczyk, M., Brzozowska, A., Wroblewska, J. & Maruniak-Chudek, I. (2015). Factors limiting usefulness of serum and urinary NGAL as a marker of acute kidney injury in preterm newborns. *Ren Fail* 37, 439-45.
- [790] Nga, H. S., Medeiros, P., Menezes, P., Bridi, R., Balbi, A. & Ponce, D. (2015). Sepsis and AKI in Clinical Emergency Room Patients: The Role of Urinary NGAL. *Biomed Res Int* **2015**, 413751.
- [791] Kanda, J., Mori, K., Kawabata, H., Kuwabara, T., Mori, K. P., Imamaki, H., Kasahara, M., Yokoi, H., Mizumoto, C., Thoennissen, N. H., Koeffler, H. P., Barasch, J., Takaori-Kondo, A., Mukoyama, M. & Nakao, K. (2014). An AKI biomarker lipocalin 2 in the blood derives from the kidney in renal injury but from neutrophils in normal and infected conditions. *Clin Exp Nephrol*.
- [792] Mårtensson, J. & Bellomo, R. (2014). The Rise and Fall of NGAL in Acute Kidney Injury. *Blood Purif* **37**, 304-310.
- [793] Srisawat, N., Praditpornsilpa, K., Patarakul, K., Techapornrung, M., Daraswang, T., Sukmark, T., Khositrangsikun, K., Fakthongyoo, A., Oranrigsupak, P., Praderm, L., Suwattanasilpa, U.,

- Peerapornratana, S., Loahaveeravat, P., Suwachittanont, N., Wirotwan, T. O., Phonork, C., Kumpunya, S., Tiranathanagul, K., Chirathaworn, C., Eiam-ong, S., Tungsanga, K., Sitprija, V., Kellum, J. A., Townamchai, N. & Thai Lepto A. K. I. study group. (2015). Neutrophil Gelatinase Associated Lipocalin (NGAL) in Leptospirosis Acute Kidney Injury: A Multicenter Study in Thailand. *PLoS One* **10**, e0143367.
- [794] Leelahavanichkul, A., Somparn, P., Issara-Amphorn, J., Eiam-Ong, S., Avihingsanon, Y., Hirankarn, N. & Srisawat, N. (2016). Serum Neutrophil Gelatinase Associated Lipocalin (NGAL) Outperforms Serum Creatinine in Detecting Sepsis-Induced Acute Kidney Injury, Experiments on Bilateral Nephrectomy and Bilateral Ureter Obstruction Mouse Models. *Shock* **45**, 570-6.
- [795] Dewerchin, M. & Carmeliet, P. (2012). PIGF: a multitasking cytokine with disease-restricted activity. Cold Spring Harb Perspect Med 2.
- [796] De Vivo, A., Baviera, G., Giordano, D., Todarello, G., Corrado, F. & D'Anna, R. (2008). Endoglin, PIGF and sFlt-1 as markers for predicting pre-eclampsia. *Acta Obstet Gynecol Scand* **87**, 837-42.
- [797] Gómez-Arriaga, P. I., Herraiz, I., López-Jiménez, E. A., Gómez-Montes, E., Denk, B. & Galindo, A. (2013). Uterine artery Doppler and sFlt-1/PIGF ratio: usefulness in diagnosis of pre-eclampsia. *Ultrasound Obstet Gynecol* **41**, 530-7.
- [798] Ohkuchi, A., Hirashima, C., Takahashi, K., Suzuki, H., Matsubara, S. & Suzuki, M. (2013). Onset threshold of the plasma levels of soluble fms-like tyrosine kinase 1/placental growth factor ratio for predicting the imminent onset of preeclampsia within 4 weeks after blood sampling at 19-31 weeks of gestation. *Hypertens Res* **36**, 1073-80.
- [799] Hirashima, C., Ohkuchi, A., Takahashi, K., Suzuki, H., Matsubara, S. & Suzuki, M. (2014). A novel three-step approach for predicting the imminent onset of preeclampsia within 4 weeks after blood sampling at 19-31 weeks of gestation. *Hypertens Res* **37**, 519-525.
- [800] Lai, J., Garcia-Tizon Larroca, S., Peeva, G., Poon, L. C., Wright, D. & Nicolaides, K. H. (2014). Competing risks model in screening for preeclampsia by serum placental growth factor and soluble fms-like tyrosine kinase-1 at 30-33 weeks' gestation. *Fetal Diagn Ther* **35**, 240-8.
- [801] Masoura, S., Kalogiannidis, I., Makedou, K., Theodoridis, T., Koiou, K., Gerou, S., Athanasiadis, A. & Agorastos, T. (2014). Biomarkers of endothelial dysfunction in preeclampsia and neonatal morbidity: a case-control study. *European Journal of Obstetrics & Gynecology and Reproductive Biology* **175**, 119-123.
- [802] Ohkuchi, A., Hirashima, C., Takahashi, K., Shirasuna, K., Suzuki, H., Ariga, H., Kobayashi, M., Hirose, N., Matsubara, S. & Suzuki, M. (2014). A trio of risk factors for the onset of preeclampsia in the second and early third trimesters. *Pregnancy Hypertension-an International Journal of Womens Cardiovascular Health* **4,** 224-230.
- [803] Dröge, L., Herraìz, I., Zeisler, H., Schlembach, D., Stepan, H., Küssel, L., Henrich, W., Galindo, A. & Verlohren, S. (2015). Maternal serum sFlt-1/PIGF ratio in twin pregnancies with and without preeclampsia in comparison with singleton pregnancies. *Ultrasound Obstet Gynecol* **45**, 286-93.
- [804] Stepan, H., Herraiz, I., Schlembach, D., Verlohren, S., Brennecke, S., Chantraine, F., Klein, E., Lapaire, O., Llurba, E., Ramoni, A., Vatish, M., Wertaschnigg, D. & Galindo, A. (2015). Implementation of the sFlt-1/PIGF ratio for prediction and diagnosis of pre-eclampsia in singleton pregnancy: implications for clinical practice. *Ultrasound Obstet Gynecol* **45**, 241-6.
- [805] Rolfo, A., Attini, R., Tavassoli, E., Neve, F. V., Nigra, M., Cicilano, M., Nuzzo, A. M., Giuffrida, D., Biolcati, M., Nichelatti, M., Gaglioti, P., Todros, T. & Piccoli, G. B. (2015). Is It Possible to Differentiate Chronic Kidney Disease and Preeclampsia by means of New and Old Biomarkers? A Prospective Study. *Dis Markers* 2015, 127083.
- [806] Khalil, A., Maiz, N., Garcia-Mandujano, R., Penco, J. M. & Nicolaides, K. H. (2016). Longitudinal changes in maternal serum placental growth factor and soluble fms-like tyrosine kinase-1 in women at increased risk of pre-eclampsia. *Ultrasound Obstet Gynecol* **47**, 324-31.
- [807] Zeisler, H., Llurba, E., Chantraine, F., Vatish, M., Staff, A. C., Sennstrom, M., Olovsson, M., Brennecke, S. P., Stepan, H., Allegranza, D., Dilba, P., Schoedl, M., Hund, M. & Verlohren, S. (2016). Predictive Value of the sFlt-1:PIGF Ratio in Women with Suspected Preeclampsia. N Engl J Med 374, 13-22.

- [808] Levine, R. J., Thadhani, R., Qian, C., Lam, C., Lim, K. H., Yu, K. F., Blink, A. L., Sachs, B. P., Epstein, F. H., Sibai, B. M., Sukhatme, V. P. & Karumanchi, S. A. (2005). Urinary placental growth factor and risk of preeclampsia. *JAMA* **293**, 77-85.
- [809] Odibo, A. O., Patel, K. R., Spitalnik, A., Odibo, L. & Huettner, P. (2014). Placental pathology, first-trimester biomarkers and adverse pregnancy outcomes. *J Perinatol* **34**, 186-191.
- [810] Bramham, K., Seed, P. T., Lightstone, L., Nelson-Piercy, C., Gill, C., Webster, P., Poston, L. & Chappell, L. C. (2016). Diagnostic and predictive biomarkers for pre-eclampsia in patients with established hypertension and chronic kidney disease. *Kidney Int* 89, 874-885.
- [811] Yano, K., Liaw, P. C., Mullington, J. M., Shih, S. C., Okada, H., Bodyak, N., Kang, P. M., Toltl, L., Belikoff, B., Buras, J., Simms, B. T., Mizgerd, J. P., Carmeliet, P., Karumanchi, S. A. & Aird, W. C. (2006). Vascular endothelial growth factor is an important determinant of sepsis morbidity and mortality. *J Exp Med* **203**, 1447-58.
- [812] Yano, K., Okada, Y., Beldi, G., Shih, S. C., Bodyak, N., Okada, H., Kang, P. M., Luscinskas, W., Robson, S. C., Carmeliet, P., Karumanchi, S. A. & Aird, W. C. (2008). Elevated levels of placental growth factor represent an adaptive host response in sepsis. J Exp Med 205, 2623-31.
- [813] Foidart, J. M., Schaaps, J. P., Chantraine, F., Munaut, C. & Lorquet, S. (2009). Dysregulation of antiangiogenic agents (sFlt-1, PLGF, and sEndoglin) in preeclampsia--a step forward but not the definitive answer. *J Reprod Immunol* **82**, 106-11.
- [814] Furuya, M., Kurasawa, K., Nagahama, K., Kawachi, K., Nozawa, A., Takahashi, T. & Aoki, I. (2011). Disrupted balance of angiogenic and antiangiogenic signalings in preeclampsia. *J Pregnancy* **2011**, 123717.
- [815] De Oliveira, L., Sass, N., Boute, T. & Moron, A. F. (2011). sFlt-1 and PIGF levels in a patient with mirror syndrome related to cytomegalovirus infection. *Eur J Obstet Gynecol Reprod Biol* **158**, 366-7.
- [816] Benton, S. J., Hu, Y., Xie, F., Kupfer, K., Lee, S. W., Magee, L. A. & von Dadelszen, P. (2012). Can placental growth factor in maternal circulation identify fetuses with placental intrauterine growth restriction? *Am J Obstet Gynecol* **206**, 163 e1-7.
- [817] Acharya, A. (2016). Promising biomarkers for superimposed pre-eclampsia in pregnant women with established hypertension and chronic kidney disease. *Kidney Int* **89**, 743-6.
- [818] Montagnana, M., Lippi, G., Albiero, A., Scevarolli, S., Salvagno, G. L., Franchi, M. & Guidi, G. C. (2008). Procalcitonin values in preeclamptic women are related to severity of disease. *Clin Chem Lab Med* **46**, 1050-1.
- [819] Can, M., Sancar, E., Harma, M., Guven, B., Mungan, G. & Acikgoz, S. (2011). Inflammatory markers in preeclamptic patients. *Clin Chem Lab Med* **49**, 1469-72.
- [820] Birdir, C., Janssen, K., Stanescu, A. D., Enekwe, A., Kasimir-Bauer, S., Gellhaus, A., Kimmig, R. & Köninger, A. (2015). Maternal serum copeptin, MR-proANP and procalcitonin levels at 11-13 weeks gestation in the prediction of preeclampsia. *Arch Gynecol Obstet* **292**, 1033-42.
- [821] Simon, L., Gauvin, F., Amre, D. K., Saint-Louis, P. & Lacroix, J. (2004). Serum procalcitonin and Creactive protein levels as markers of bacterial infection: a systematic review and meta-analysis. *Clin Infect Dis* **39**, 206-17.
- [822] Ucar, B., Yildiz, B., Aksit, M. A., Yarar, C., Colak, O., Akbay, Y. & Colak, E. (2008). Serum amyloid A, procalcitonin, tumor necrosis factor-alpha, and interleukin-1beta levels in neonatal late-onset sepsis. *Mediators Inflamm* **2008**, 737141.
- [823] Riedel, S., Melendez, J. H., An, A. T., Rosenbaum, J. E. & Zenilman, J. M. (2011). Procalcitonin as a marker for the detection of bacteremia and sepsis in the emergency department. *Am J Clin Pathol* **135**, 182-9.
- [824] Riedel, S. (2012). Procalcitonin and the role of biomarkers in the diagnosis and management of sepsis. *Diagn Microbiol Infect Dis* **73**, 221-7.
- [825] Schuetz, P., Muller, B., Christ-Crain, M., Stolz, D., Tamm, M., Bouadma, L., Luyt, C. E., Wolff, M., Chastre, J., Tubach, F., Kristoffersen, K. B., Burkhardt, O., Welte, T., Schroeder, S., Nobre, V., Wei, L., Bhatnagar, N., Bucher, H. C. & Briel, M. (2012). Procalcitonin to initiate or discontinue antibiotics in acute respiratory tract infections. *Cochrane Database Syst Rev* **9**, CD007498.
- [826] Ulla, M., Pizzolato, E., Lucchiari, M., Loiacono, M., Soardo, F., Forno, D., Morello, F., Lupia, E., Moiraghi, C., Mengozzi, G. & Battista, S. (2013). Diagnostic and prognostic value of presepsin in the

- management of sepsis in the emergency department: a multicenter prospective study. *Critical Care*
- [827] Henriquez-Camacho, C. & Losa, J. (2014). Biomarkers for Sepsis. Biomed Research International.
- [828] Tian, G., Pan, S. Y., Ma, G., Liao, W., Su, Q. G., Gu, B. C. & Qin, K. (2014). Serum levels of procalcitonin as a biomarker for differentiating between sepsis and systemic inflammatory response syndrome in the neurological intensive care unit. *J Clin Neurosci* 21, 1153-8.
- [829] Westwood, M., Ramaekers, B., Whiting, P., Tomini, F., Joore, M., Armstrong, N., Ryder, S., Stirk, L., Severens, J. & Kleijnen, J. (2015). Procalcitonin testing to guide antibiotic therapy for the treatment of sepsis in intensive care settings and for suspected bacterial infection in emergency department settings: a systematic review and cost-effectiveness analysis. *Health Technol Assess* 19, v-xxv, 1-236.
- [830] Engin-Üstün, Y., Üstün, Y., Karabulut, A. B., Özkaplan, E., Meydanli, M. M. & Kafkaslı, A. (2007). Serum amyloid A levels are increased in pre-eclampsia. *Gynecol Obstet Invest* **64,** 117-20.
- [831] Kristensen, K., Wide-Swensson, D., Lindström, V., Schmidt, C., Grubb, A. & Strevens, H. (2009). Serum amyloid A protein and C-reactive protein in normal pregnancy and preeclampsia. *Gynecol Obstet Invest* **67**, 275-80.
- [832] Malle, E. & De Beer, F. C. (1996). Human serum amyloid A (SAA) protein: a prominent acute-phase reactant for clinical practice. *Eur J Clin Invest* **26,** 427-35.
- [833] Pizzini, C., Mussap, M., Plebani, M. & Fanos, V. (2000). C-reactive protein and serum amyloid A protein in neonatal infections. *Scand J Infect Dis* **32**, 229-35.
- [834] Sipe, J. D. (2000). Serum amyloid A: from fibril to function. Current status. Amyloid 7, 10-2.
- [835] Urieli-Shoval, S., Linke, R. P. & Matzner, Y. (2000). Expression and function of serum amyloid A, a major acute-phase protein, in normal and disease states. *Curr Opin Hematol* **7**, 64-9.
- [836] Falsey, A. R., Walsh, E. E., Francis, C. W., Looney, R. J., Kolassa, J. E., Hall, W. J. & Abraham, G. N. (2001). Response of C-reactive protein and serum amyloid A to influenza A infection in older adults. *J Infect Dis* **183**, 995-9.
- [837] Arnon, S., Litmanovitz, I., Regev, R. H., Bauer, S., Shainkin-Kestenbaum, R. & Dolfin, T. (2007). Serum amyloid A: an early and accurate marker of neonatal early-onset sepsis. *J Perinatol* **27**, 297-302.
- [838] Bozinovski, S., Hutchinson, A., Thompson, M., Macgregor, L., Black, J., Giannakis, E., Karlsson, A. S., Silvestrini, R., Smallwood, D., Vlahos, R., Irving, L. B. & Anderson, G. P. (2008). Serum amyloid a is a biomarker of acute exacerbations of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* **177**, 269-78.
- [839] Cicarelli, D. D., Vieira, J. E. & Bensenor, F. E. (2008). Comparison of C-reactive protein and serum amyloid a protein in septic shock patients. *Mediators Inflamm* **2008**, 631414.
- [840] Ebert, E. C. & Nagar, M. (2008). Gastrointestinal manifestations of amyloidosis. *Am J Gastroenterol* **103,** 776-87.
- [841] Lannergård, A., Larsson, A., Friman, G. & Ewald, U. (2008). Human serum amyloid A (SAA) and high sensitive C-reactive protein (hsCRP) in preterm newborn infants with nosocomial infections. *Acta Paediatr* **97**, 1061-5.
- [842] Çetinkaya, M., Özkan, H., Köksal, N., Çelebi, S. & Hacımustafaoğlu, M. (2009). Comparison of serum amyloid A concentrations with those of C-reactive protein and procalcitonin in diagnosis and follow-up of neonatal sepsis in premature infants. *J Perinatol* **29**, 225-31.
- [843] Lannergård, A., Viberg, A., Cars, O., Karlsson, M. O., Sandström, M. & Larsson, A. (2009). The time course of body temperature, serum amyloid A protein, C-reactive protein and interleukin-6 in patients with bacterial infection during the initial 3 days of antibiotic therapy. *Scand J Infect Dis* **41**, 663-71.
- [844] Yuan, H., Huang, J., Lv, B., Yan, W., Hu, G., Wang, J. & Shen, B. (2013). Diagnosis value of the serum amyloid A test in neonatal sepsis: a meta-analysis. *Biomed Res Int* **2013**, 520294.
- [845] Derebe, M. G., Zlatkov, C. M., Gattu, S., Ruhn, K. A., Vaishnava, S., Diehl, G. E., MacMillan, J. B., Williams, N. S. & Hooper, L. V. (2014). Serum amyloid A is a retinol binding protein that transports retinol during bacterial infection. *Elife* **3**, e03206.

- [846] Buhimschi, I. A., Nayeri, U. A., Zhao, G., Shook, L. L., Pensalfini, A., Funai, E. F., Bernstein, I. M., Glabe, C. G. & Buhimschi, C. S. (2014). Protein misfolding, congophilia, oligomerization, and defective amyloid processing in preeclampsia. Sci Transl Med 6, 245ra92.
- [847] Little, C. S., Hammond, C. J., MacIntyre, A., Balin, B. J. & Appelt, D. M. (2004). *Chlamydia pneumoniae* induces Alzheimer-like amyloid plaques in brains of BALB/c mice. *Neurobiol Aging* **25**, 419-29.
- [848] Wozniak, M. A., Itzhaki, R. F., Shipley, S. J. & Dobson, C. B. (2007). Herpes simplex virus infection causes cellular beta-amyloid accumulation and secretase upregulation. *Neurosci Lett* **429**, 95-100.
- [849] Veas, C. J., Aguilera, V. C., Munoz, I. J., Gallardo, V. I., Miguel, P. L., Gonzalez, M. A., Lamperti, L. I., Escudero, C. A. & Aguayo, C. R. (2011). Fetal endothelium dysfunction is associated with circulating maternal levels of sE-selectin, sVCAM1, and sFlt-1 during pre-eclampsia. *J Matern Fetal Neonatal Med* 24, 1371-7.
- [850] Whitehead, C. L., Palmer, K. R., Nilsson, U., Gao, Y., Saglam, B., Lappas, M. & Tong, S. (2011). Placental expression of a novel primate-specific splice variant of sFlt-1 is upregulated in pregnancies complicated by severe early onset pre-eclampsia. *BJOG* 118, 1268-71.
- [851] Higgins, L. E., Rey de Castro, N., Addo, N., Wareing, M., Greenwood, S. L., Jones, R. L., Sibley, C. P., Johnstone, E. D. & Heazell, A. E. P. (2015). Placental Features of Late-Onset Adverse Pregnancy Outcome. *PLoS One* 10, e0129117.
- [852] Hod, T., Cerdeira, A. S. & Karumanchi, S. A. (2015). Molecular Mechanisms of Preeclampsia. *Cold Spring Harb Perspect Med* **5**.
- [853] Li, F., Hagaman, J. R., Kim, H. S., Maeda, N., Jennette, J. C., Faber, J. E., Karumanchi, S. A., Smithies, O. & Takahashi, N. (2012). eNOS deficiency acts through endothelin to aggravate sFlt-1-induced preeclampsia-like phenotype. *J Am Soc Nephrol* **23**, 652-60.
- [854] Thadhani, R., Kisner, T., Hagmann, H., Bossung, V., Noack, S., Schaarschmidt, W., Jank, A., Kribs, A., Cornely, O. A., Kreyssig, C., Hemphill, L., Rigby, A. C., Khedkar, S., Lindner, T. H., Mallmann, P., Stepan, H., Karumanchi, S. A. & Benzing, T. (2011). Pilot study of extracorporeal removal of soluble fms-like tyrosine kinase 1 in preeclampsia. *Circulation* **124**, 940-50.
- [855] Carney, E. F. (2015). Hypertension: sFlt-1 removal seems to be beneficial in women with pre-eclampsia. *Nat Rev Nephrol* **11,** 690.
- [856] Thadhani, R., Hagmann, H., Schaarschmidt, W., Roth, B., Cingoez, T., Karumanchi, S. A., Wenger, J., Lucchesi, K. J., Tamez, H., Lindner, T., Fridman, A., Thome, U., Kribs, A., Danner, M., Hamacher, S., Mallmann, P., Stepan, H. & Benzing, T. (2016). Removal of Soluble Fms-Like Tyrosine Kinase-1 by Dextran Sulfate Apheresis in Preeclampsia. J Am Soc Nephrol 27, 903-13.
- [857] Shapiro, N. I., Yano, K., Okada, H., Fischer, C., Howell, M., Spokes, K. C., Ngo, L., Angus, D. C. & Aird, W. C. (2008). A prospective, observational study of soluble FLT-1 and vascular endothelial growth factor in sepsis. Shock 29, 452-7.
- [858] Schuetz, P., Jones, A. E., Aird, W. C. & Shapiro, N. I. (2011). Endothelial cell activation in emergency department patients with sepsis-related and non-sepsis-related hypotension. *Shock* **36**, 104-8.
- [859] Shapiro, N. I., Schuetz, P., Yano, K., Sorasaki, M., Parikh, S. M., Jones, A. E., Trzeciak, S., Ngo, L. & Aird, W. C. (2010). The association of endothelial cell signaling, severity of illness, and organ dysfunction in sepsis. *Crit Care* **14**, R182.
- [860] Alves, B. E., Montalvao, S. A., Aranha, F. J., Lorand-Metze, I., De Souza, C. A., Annichino-Bizzacchi, J. M. & De Paula, E. V. (2011). Time-course of sFlt-1 and VEGF-A release in neutropenic patients with sepsis and septic shock: a prospective study. *J Transl Med* 9, 23.
- [861] Xing, K., Murthy, S., Liles, W. C. & Singh, J. M. (2012). Clinical utility of biomarkers of endothelial activation in sepsis--a systematic review. *Crit Care* **16**, R7.
- [862] Minakami, H., Takahashi, T., Izumi, A. & Tamada, T. (1993). Increased levels of plasma thrombomodulin in preeclampsia. *Gynecol Obstet Invest* **36**, 208-10.
- [863] Hsu, C. D., Iriye, B., Johnson, T. R., Witter, F. R., Hong, S. F. & Chan, D. W. (1993). Elevated circulating thrombomodulin in severe preeclampsia. *Am J Obstet Gynecol* **169**, 148-9.
- [864] Bontis, J., Vavilis, D., Agorastos, T., Zournatzi, V., Konstantinidis, T. & Tagou, K. (1995). Maternal plasma level of thrombomodulin is increased in mild preeclampsia. Eur J Obstet Gynecol Reprod Biol 60, 139-41.

- [865] Hsu, C. D., Copel, J. A., Hong, S. F. & Chan, D. W. (1995). Thrombomodulin levels in preeclampsia, gestational hypertension, and chronic hypertension. *Obstet Gynecol* **86**, 897-9.
- [866] Shaarawy, M. & Didy, H. E. (1996). Thrombomodulin, plasminogen activator inhibitor type 1 (PAI-1) and fibronectin as biomarkers of endothelial damage in preeclampsia and eclampsia. *Int J Gynaecol Obstet* **55**, 135-9.
- [867] Boffa, M. C., Valsecchi, L., Fausto, A., Gozin, D., Vigano' D'Angelo, S., Safa, O., Castiglioni, M. T., Amiral, J. & D'Angelo, A. (1998). Predictive value of plasma thrombomodulin in preeclampsia and gestational hypertension. *Thromb Haemost* **79**, 1092-5.
- [868] Kobayashi, H., Sadakata, H., Suzuki, K., She, M. Y., Shibata, S. & Terao, T. (1998). Thrombomodulin release from umbilical endothelial cells initiated by preeclampsia plasma-induced neutrophil activation. *Obstet Gynecol* **92**, 425-30.
- [869] Brenner, B. (2004). Haemostatic changes in pregnancy. Thromb Res 114, 409-14.
- [870] Bosco, C., Parra, M., Barja, P., Rodrigo, R., Fernández, V., Suarez, M. & Muñoz, H. (2005). Increased immunohistochemical expression of thrombomodulin at placental perivascular myofibroblast in severe preeclampsia (PE). *Histol Histopathol* **20**, 1045-55.
- [871] Wiwanitkit, V. (2008). Correlation between thrombomodulin and severe preeclampsia: a summary. *Clin Appl Thromb Hemost* **14**, 99-101.
- [872] Turner, R. J., Bloemenkamp, K. W. M., Bruijn, J. A. & Baelde, H. J. (2016). Loss of Thrombomodulin in Placental Dysfunction in Preeclampsia. *Arterioscler Thromb Vasc Biol* **36**, 728-35.
- [873] Saito, H., Maruyama, I., Shimazaki, S., Yamamoto, Y., Aikawa, N., Ohno, R., Hirayama, A., Matsuda, T., Asakura, H., Nakashima, M. & Aoki, N. (2007). Efficacy and safety of recombinant human soluble thrombomodulin (ART-123) in disseminated intravascular coagulation: results of a phase III, randomized, double-blind clinical trial. *J Thromb Haemost* **5**, 31-41.
- [874] Levi, M. & Van Der Poll, T. (2013). Thrombomodulin in sepsis. Minerva Anestesiol 79, 294-8.
- [875] Mimuro, J., Takahashi, H., Kitajima, I., Tsuji, H., Eguchi, Y., Matsushita, T., Kuroda, T. & Sakata, Y. (2013). Impact of recombinant soluble thrombomodulin (thrombomodulin alfa) on disseminated intravascular coagulation. *Thromb Res* **131**, 436-43.
- [876] Vincent, J. L., Ramesh, M. K., Ernest, D., LaRosa, S. P., Pachl, J., Aikawa, N., Hoste, E., Levy, H., Hirman, J., Levi, M., Daga, M., Kutsogiannis, D. J., Crowther, M., Bernard, G. R., Devriendt, J., Puigserver, J. V., Blanzaco, D. U., Esmon, C. T., Parrillo, J. E., Guzzi, L., Henderson, S. J., Pothirat, C., Mehta, P., Fareed, J., Talwar, D., Tsuruta, K., Gorelick, K. J., Osawa, Y. & Kaul, I. (2013). A randomized, double-blind, placebo-controlled, Phase 2b study to evaluate the safety and efficacy of recombinant human soluble thrombomodulin, ART-123, in patients with sepsis and suspected disseminated intravascular coagulation. *Crit Care Med* 41, 2069-79.
- [877] Shirahata, A., Mimuro, J., Takahashi, H., Kitajima, I., Tsuji, H., Eguchi, Y., Matsushita, T., Kajiki, M., Honda, G. & Sakata, Y. (2014). Recombinant soluble human thrombomodulin (thrombomodulin alfa) in the treatment of neonatal disseminated intravascular coagulation. *Eur J Pediatr* 173, 303-11.
- [878] Yamakawa, K., Aihara, M., Ogura, H., Yuhara, H., Hamasaki, T. & Shimazu, T. (2015). Recombinant human soluble thrombomodulin in severe sepsis: a systematic review and meta-analysis. *J Thromb Haemost* **13**, 508-19.
- [879] Yoshimura, J., Yamakawa, K., Ogura, H., Umemura, Y., Takahashi, H., Morikawa, M., Inoue, Y., Fujimi, S., Tanaka, H., Hamasaki, T. & Shimazu, T. (2015). Benefit profile of recombinant human soluble thrombomodulin in sepsis-induced disseminated intravascular coagulation: a multicenter propensity score analysis. *Crit Care* **19**, 78.
- [880] Levi, M. (2015). Recombinant soluble thrombomodulin: coagulation takes another chance to reduce sepsis mortality. *J Thromb Haemost* **13,** 505-7.
- [881] Hayakawa, M., Yamakawa, K., Saito, S., Uchino, S., Kudo, D., Iizuka, Y., Sanui, M., Takimoto, K., Mayumi, T., Ono, K. & Japan Septic Disseminated Intravascular Coagulation study, g. (2016). Recombinant human soluble thrombomodulin and mortality in sepsis-induced disseminated intravascular coagulation. A multicentre retrospective study. *Thromb Haemost* 115.

- [882] Kim, Y. M., Romero, R., Oh, S. Y., Kim, C. J., Kilburn, B. A., Armant, D. R., Nien, J. K., Gomez, R., Mazor, M., Saito, S., Abrahams, V. M. & Mor, G. (2005). Toll-like receptor 4: A potential link between "danger signals," the innate immune system and preeclampsia? *Am J Obstet Gynecol* **193**, 921-927.
- [883] Bonney, E. A. (2007). Preeclampsia: a view through the danger model. *Journal of Reproductive Immunology* **76**, 68-74.
- [884] Fasshauer, M., Waldeyer, T., Seeger, J., Schrey, S., Ebert, T., Kratzsch, J., Lossner, U., Bluher, M., Stumvoll, M., Faber, R. & Stepan, H. (2008). Serum levels of the adipokine visfatin are increased in pre-eclampsia. *Clin Endocrinol (Oxf)* **69,** 69-73.
- [885] Adali, E., Yildizhan, R., Kolusari, A., Kurdoglu, M., Bugdayci, G., Sahin, H. G. & Kamaci, M. (2009). Increased visfatin and leptin in pregnancies complicated by pre-eclampsia. *J Matern Fetal Neonatal Med* **22**, 873-9.
- [886] Mazaki-Tovi, S., Vaisbuch, E., Romero, R., Kusanovic, J. P., Chaiworapongsa, T., Kim, S. K., Nhan-Chang, C. L., Gomez, R., Alpay Savasan, Z., Madan, I., Yoon, B. H., Yeo, L., Mittal, P., Ogge, G., Gonzalez, J. M. & Hassan, S. S. (2010). Maternal and neonatal circulating visfatin concentrations in patients with pre-eclampsia and a small-for-gestational age neonate. *J Matern Fetal Neonatal Med* 23, 1119-28.
- [887] Hu, W., Wang, Z., Wang, H., Huang, H. & Dong, M. (2008). Serum visfatin levels in late pregnancy and pre-eclampsia. *Acta Obstet Gynecol Scand* **87**, 413-8.
- [888] Zonneveld, R., Martinelli, R., Shapiro, N. I., Kuijpers, T. W., Plotz, F. B. & Carman, C. V. (2014). Soluble adhesion molecules as markers for sepsis and the potential pathophysiological discrepancy in neonates, children and adults. *Crit Care* 18, 204.
- [889] de Pablo, R., Monserrat, J., Reyes, E., Díaz, D., Rodríguez-Zapata, M., de la Hera, A., Prieto, A. & Álvarez-Mon, M. (2013). Circulating sICAM-1 and sE-Selectin as biomarker of infection and prognosis in patients with systemic inflammatory response syndrome. *Eur J Intern Med* **24**, 132-8.
- [890] Endo, S., Suzuki, Y., Takahashi, G., Shozushima, T., Ishikura, H., Murai, A., Nishida, T., Irie, Y., Miura, M., Iguchi, H., Fukui, Y., Tanaka, K., Nojima, T. & Okamura, Y. (2012). Usefulness of presepsin in the diagnosis of sepsis in a multicenter prospective study. *J Infect Chemother* **18**, 891-7.
- [891] Endo, S., Suzuki, Y., Takahashi, G., Shozushima, T., Ishikura, H., Murai, A., Nishida, T., Irie, Y., Miura, M., Iguchi, H., Fukui, Y., Tanaka, K., Nojima, T. & Okamura, Y. (2014). Presepsin as a powerful monitoring tool for the prognosis and treatment of sepsis: a multicenter prospective study. *J Infect Chemother* **20**, 30-4.
- [892] Mussap, M., Noto, A., Fravega, M. & Fanos, V. (2011). Soluble CD14 subtype presepsin (sCD14-ST) and lipopolysaccharide binding protein (LBP) in neonatal sepsis: new clinical and analytical perspectives for two old biomarkers. *J Matern Fetal Neonatal Med* **24 Suppl 2,** 12-4.
- [893] Nakamura, Y., Ishikura, H., Nishida, T., Kawano, Y., Yuge, R., Ichiki, R. & Murai, A. (2014). Usefulness of presepsin in the diagnosis of sepsis in patients with or without acute kidney injury. *BMC Anesthesiol* **14**, 88.
- [894] Okamura, Y. & Yokoi, H. (2011). Development of a point-of-care assay system for measurement of presepsin (sCD14-ST). *Clin Chim Acta* **412**, 2157-61.
- [895] Wu, J., Hu, L., Zhang, G., Wu, F. & He, T. (2015). Accuracy of Presepsin in Sepsis Diagnosis: A Systematic Review and Meta-Analysis. *PLoS One* **10**, e0133057.
- [896] Kell, D. B. & Westerhoff, H. V. (1986). Metabolic control theory: its role in microbiology and biotechnology. *FEMS Microbiol. Rev.* **39**, 305-320.
- [897] Kell, D. B. (1987). Forces, fluxes and the control of microbial growth and metabolism. The twelfth Fleming lecture. *J. Gen. Microbiol.* **133**, 1651-1665.
- [898] Fell, D. A. (1992). Metabolic Control Analysis a survey of its theoretical and experimental development. *Biochem. J.* **286**, 313-330.
- [899] Heinrich, R. & Schuster, S. (1996). The regulation of cellular systems. Chapman & Hall, New York.
- [900] Oliver, S. G., Winson, M. K., Kell, D. B. & Baganz, F. (1998). Systematic functional analysis of the yeast genome. *Trends Biotechnol.* **16**, 373-378.
- [901] Raamsdonk, L. M., Teusink, B., Broadhurst, D., Zhang, N., Hayes, A., Walsh, M., Berden, J. A., Brindle, K. M., Kell, D. B., Rowland, J. J., Westerhoff, H. V., van Dam, K. & Oliver, S. G. (2001). A functional genomics strategy that uses metabolome data to reveal the phenotype of silent mutations. *Nature Biotechnol.* **19**, 45-50.

- [902] Cornish-Bowden, A. & Cárdenas, M. L. (2001). Silent genes given voice. Nature 409, 571-572.
- [903] Goodacre, R., Vaidyanathan, S., Dunn, W. B., Harrigan, G. G. & Kell, D. B. (2004). Metabolomics by numbers: acquiring and understanding global metabolite data. *Trends Biotechnol.* **22**, 245-252.
- [904] Kell, D. B. (2004). Metabolomics and systems biology: making sense of the soup. *Curr. Op. Microbiol.* **7,** 296-307.
- [905] Kitano, H., Oda, K., Kimura, T., Matsuoka, Y., Csete, M., Doyle, J. & Muramatsu, M. (2004). Metabolic syndrome and robustness tradeoffs. *Diabetes* **53 Suppl 3**, S6-S15.
- [906] Kitano, H. (2004). Biological robustness. Nat Rev Genet 5, 826-37.
- [907] Wilhelm, T., Behre, J. & Schuster, S. (2004). Analysis of structural robustness of metabolic networks. *Syst Biol (Stevenage)* **1,** 114-20.
- [908] Grimbs, S., Selbig, J., Bulik, S., Holzhutter, H. G. & Steuer, R. (2007). The stability and robustness of metabolic states: identifying stabilizing sites in metabolic networks. *Mol Syst Biol* **3**, 146.
- [909] Kell, D. B. (2013). Finding novel pharmaceuticals in the systems biology era using multiple effective drug targets, phenotypic screening, and knowledge of transporters: where drug discovery went wrong and how to fix it. *FEBS J* **280**, 5957-5980.
- [910] Quinton-Tulloch, M. J., Bruggeman, F. J., Snoep, J. L. & Westerhoff, H. V. (2013). Trade-off of dynamic fragility but not of robustness in metabolic pathways in silico. *FEBS J* **280**, 160-73.
- [911] Kell, D. B. & Goodacre, R. (2014). Metabolomics and systems pharmacology: why and how to model the human metabolic network for drug discovery. *Drug Disc Today* **19**, 171-182.
- [912] Begley, P., Francis-McIntyre, S., Dunn, W. B., Broadhurst, D. I., Halsall, A., Tseng, A., Knowles, J., HUSERMET consortium, Goodacre, R. & Kell, D. B. (2009). Development and performance of a gas chromatography-time-of-flight mass spectrometry analysis for large-scale non-targeted metabolomic studies of human serum. *Anal Chem* **81**, 7038-7046.
- [913] Zelena, E., Dunn, W. B., Broadhurst, D., Francis-McIntyre, S., Carroll, K. M., Begley, P., O'Hagan, S., Knowles, J. D., Halsall, A., HUSERMET Consortium, Wilson, I. D. & Kell, D. B. (2009). Development of a robust and repeatable UPLC-MS method for the long-term metabolomic study of human serum. *Anal Chem* **81**, 1357-1364.
- [914] Dunn, W. B., Broadhurst, D., Begley, P., Zelena, E., Francis-McIntyre, S., Anderson, N., Brown, N., Knowles, J., Halsall, A., Haselden, J. N., Nicholls, A., Wilson, I. D., Kell, D. B., Goodacre, R. & The Husermet consortium. (2011). Procedures for large-scale metabolic profiling of serum and plasma using gas chromatography and liquid chromatography coupled to mass spectrometry. *Nat Protoc* 6, 1060-1083.
- [915] Dunn, W. B., Lin, W., Broadhurst, D., Begley, P., Brown, M., Zelena, E., Vaughan, A. A., Halsall, A., Harding, N., Knowles, J. D., Francis-McIntyre, S., Tseng, A., Ellis, D. I., O'Hagan, S., Aarons, G., Benjamin, B., Chew-Graham, S., Moseley, C., Potter, P., Winder, C. L., Potts, C., Thornton, P., McWhirter, C., Zubair, M., Burns, A., Cruickshank, J. K., Jayson, G. C., Purandare, N., Wu, F. W., Finn, J. D., Haselden, J. N., Nicholls, A. W., Wilson, I. D., Goodacre, R. & Kell, D. B. (2015). Molecular phenotyping of a UK population: defining the human serum metabolome. *Metabolomics* 11, 9-26.
- [916] Thiele, I., Swainston, N., Fleming, R. M. T., Hoppe, A., Sahoo, S., Aurich, M. K., Haraldsdottír, H., Mo, M. L., Rolfsson, O., Stobbe, M. D., Thorleifsson, S. G., Agren, R., Bölling, C., Bordel, S., Chavali, A. K., Dobson, P., Dunn, W. B., Endler, L., Goryanin, I., Hala, D., Hucka, M., Hull, D., Jameson, D., Jamshidi, N., Jones, J., Jonsson, J. J., Juty, N., Keating, S., Nookaew, I., Le Novère, N., Malys, N., Mazein, A., Papin, J. A., Patel, Y., Price, N. D., Selkov Sr., E., Sigurdsson, M. I., Simeonidis, E., Sonnenschein, N., Smallbone, K., Sorokin, A., Beek, H. V., Weichart, D., Nielsen, J. B., Westerhoff, H. V., Kell, D. B., Mendes, P. & Palsson, B. Ø. (2013). A community-driven global reconstruction of human metabolism. Nat Biotechnol. 31, 419-425.
- [917] Swainston, N., Mendes, P. & Kell, D. B. (2013). An analysis of a 'community-driven' reconstruction of the human metabolic network. *Metabolomics* **9**, 757-764.
- [918] Swainston, N., Smallbone, K., Hefzi, H., Dobson, P. D., Brewer, J., Hanscho, M., Zielinski, D. C., Ang, K. S., Gardiner, N. J., Gutierrez, J. M., Kyriakopoulos, S., Lakshmanan, M., Li, S., Liu, J. K., Martínez, V. S., Orellana, C. A., Quek, L.-E., Thomas, A., Zanghellini, J., Borth, N., Lee, D.-Y., Nielsen, L. K., Kell, D. B., Lewis, N. E. & Mendes, P. (2016). Recon 2.2: from reconstruction to model of human metabolism. *Metabolomics*, in press.

- [919] Herrgård, M. J., Swainston, N., Dobson, P., Dunn, W. B., Arga, K. Y., Arvas, M., Blüthgen, N., Borger, S., Costenoble, R., Heinemann, M., Hucka, M., Le Novère, N., Li, P., Liebermeister, W., Mo, M. L., Oliveira, A. P., Petranovic, D., Pettifer, S., Simeonidis, E., Smallbone, K., Spasić, I., Weichart, D., Brent, R., Broomhead, D. S., Westerhoff, H. V., Kırdar, B., Penttilä, M., Klipp, E., Palsson, B. Ø., Sauer, U., Oliver, S. G., Mendes, P., Nielsen, J. & Kell, D. B. (2008). A consensus yeast metabolic network obtained from a community approach to systems biology. *Nat Biotechnol.* **26**, 1155-1160.
- [920] Bolte, A. C., van Geijn, H. P. & Dekker, G. A. (2001). Pathophysiology of preeclampsia and the role of serotonin. *European Journal of Obstetrics Gynecology and Reproductive Biology* **95**, 12-21.
- [921] Kell, D. B., Kaprelyants, A. S. & Grafen, A. (1995). On pheromones, social behaviour and the functions of secondary metabolism in bacteria. *Trends Ecol. Evolution* **10**, 126-129.
- [922] Takano, E. (2006). Gamma-butyrolactones: *Streptomyces* signalling molecules regulating antibiotic production and differentiation. *Curr Opin Microbiol* **9**, 287-94.
- [923] Hyppönen, E., Cavadino, A., Williams, D., Fraser, A., Vereczkey, A., Fraser, W. D., Banhidy, F., Lawlor, D. & Czeizel, A. E. (2013). Vitamin D and pre-eclampsia: original data, systematic review and meta-analysis. *Ann Nutr Metab* **63**, 331-40.
- [924] Tabesh, M., Salehi-Abargouei, A., Tabesh, M. & Esmaillzadeh, A. (2013). Maternal vitamin D status and risk of pre-eclampsia: a systematic review and meta-analysis. *J Clin Endocrinol Metab* **98,** 3165-73.
- [925] Bakacak, M., Serin, S., Ercan, O., Kostu, B., Avci, F., Kilinc, M., Kiran, H. & Kiran, G. (2015). Comparison of Vitamin D levels in cases with preeclampsia, eclampsia and healthy pregnant women. *Int J Clin Exp Med* **8**, 16280-6.
- [926] Barrera, D., Diaz, L., Noyola-Martinez, N. & Halhali, A. (2015). Vitamin D and Inflammatory Cytokines in Healthy and Preeclamptic Pregnancies. *Nutrients* **7**, 6465-90.
- [927] Gargari, B. P., Tabrizi, F. P. F., Sadien, B., Jafarabadi, M. A. & Farzadi, L. (2015). Vitamin D Status Is Related to Oxidative Stress But Not High-Sensitive C-Reactive Protein in Women with Pre-Eclampsia. *Gynecol Obstet Invest*.
- [928] Harvey, N. C., Holroyd, C., Ntani, G., Javaid, K., Cooper, P., Moon, R., Cole, Z., Tinati, T., Godfrey, K., Dennison, E., Bishop, N. J., Baird, J. & Cooper, C. (2014). Vitamin D supplementation in pregnancy: a systematic review. *Health Technol Assess* **18**, 1-190.
- [929] van Weert, B., van den Berg, D., Hrudey, E. J., Oostvogels, A. J. J. M., de Miranda, E. & Vrijkotte, T. G. M. (2016). Is first trimester vitamin D status in nulliparous women associated with pregnancy related hypertensive disorders? *Midwifery* 34, 117-22.
- [930] Kiely, M. E., Zhang, J. Y., Kinsella, M., Khashan, A. S. & Kenny, L. C. (2016). Vitamin D status is associated with utero-placental dysfunction in a large prospective pregnancy cohort with low 25(OH)D<sub>3</sub> and ubiquitous 3-epi-25(OH)D<sub>3</sub> and 25(OH)D<sub>2</sub>. *Am J Clin Nutr*, in press.
- [931] Palacios, C., De-Regil, L. M., Lombardo, L. K. & Peña-Rosas, J. P. (2016). Vitamin D supplementation during pregnancy: Updated meta-analysis on maternal outcomes. *J Steroid Biochem Mol Biol*.
- [932] Bahado-Singh, R. O., Syngelaki, A., Akolekar, R., Mandal, R., Bjondahl, T. C., Han, B., Dong, E., Bauer, S., Alpay-Savasan, Z., Graham, S., Turkoglu, O., Wishart, D. S. & Nicolaides, K. H. (2015). Validation of metabolomic models for prediction of early-onset preeclampsia. *Am J Obstet Gynecol* **213**, 530 e1-530 e10.
- [933] Melland-Smith, M., Ermini, L., Chauvin, S., Craig-Barnes, H., Tagliaferro, A., Todros, T., Post, M. & Caniggia, I. (2015). Disruption of sphingolipid metabolism augments ceramide-induced autophagy in preeclampsia. *Autophagy* 11, 653-69.
- [934] Funke, C., Schneider, S. A., Berg, D. & Kell, D. B. (2013). Genetics and iron in the systems biology of Parkinson's disease and some related disorders. *Neurochem Internat* **62**, 637-652.
- [935] Manyonda, I. T., Slater, D. M., Fenske, C., Hole, D., Choy, M. Y. & Wilson, C. (1998). A role for noradrenaline in pre-eclampsia: towards a unifying hypothesis for the pathophysiology. *Br J Obstet Gynaecol* **105**, 641-8.
- [936] Ferreira-de-Almeida, J. A., Amenta, F., Cardoso, F. & Polonia, J. J. (1998). Association of circulating endothelium and noradrenaline with increased calcium-channel binding sites in the placental bed in pre-eclampsia. *Br J Obstet Gynaecol* **105**, 1104-12.

- [937] Freestone, P. P., Haigh, R. D., Williams, P. H. & Lyte, M. (1999). Stimulation of bacterial growth by heat-stable, norepinephrine-induced autoinducers. *FEMS Microbiol Lett* **172**, 53-60.
- [938] Lyte, M. & Ernst, S. (1992). Catecholamine-induced growth of Gram-negative bacteria. *Life Sci.* **50**, 203-212.
- [939] Lyte, M. (2004). Microbial endocrinology and infectious disease in the 21st century. *Trends Microbiol* **12**, 14-20.
- [940] Lyte, M. (2014). The effect of stress on microbial growth. Anim Health Res Rev 15, 172-4.
- [941] Reissbrodt, R., Rienaecker, I., Romanova, J. M., Freestone, P. P. E., Haigh, R. D., Lyte, M., Tschäpe, H. & Williams, P. H. (2002). Resuscitation of *Salmonella enterica* serovar *typhimurium* and enterohemorrhagic *Escherichia coli* from the viable but nonculturable state by heat-stable enterobacterial autoinducer. *Appl Environ Microbiol* **68**, 4788-94.
- [942] Freestone, P. P., Lyte, M., Neal, C. P., Maggs, A. F., Haigh, R. D. & Williams, P. H. (2000). The mammalian neuroendocrine hormone norepinephrine supplies iron for bacterial growth in the presence of transferrin or lactoferrin. *J Bacteriol* **182**, 6091-8.
- [943] Freestone, P. P. E., Haigh, R. D., Williams, P. H. & Lyte, M. (2003). Involvement of enterobactin in norepinephrine-mediated iron supply from transferrin to enterohaemorrhagic *Escherichia coli*. *FEMS Microbiol Lett* **222**, 39-43.
- [944] O'Donnell, P. M., Aviles, H., Lyte, M. & Sonnenfeld, G. (2006). Enhancement of *in vitro* growth of pathogenic bacteria by norepinephrine: importance of inoculum density and role of transferrin. *Appl Environ Microbiol* **72**, 5097-9.
- [945] Kvetnansky, R., Sabban, E. L. & Palkovits, M. (2009). Catecholaminergic systems in stress: structural and molecular genetic approaches. *Physiol Rev* **89**, 535-606.
- [946] Ribas Ripoll, V., Romay, E., Brunelli, L., Pastorelli, R., Goma, G., Navas, A., Artigas, A. & Ferrer, R. (2015). Metabolite analysis in sepsis through conditional independence maps. *Conf Proc IEEE Eng Med Biol Soc* **2015**, 6477-80.
- [947] Su, L., Huang, Y., Zhu, Y., Xia, L., Wang, R., Xiao, K., Wang, H., Yan, P., Wen, B., Cao, L., Meng, N., Luan, H., Liu, C., Li, X. & Xie, L. (2014). Discrimination of sepsis stage metabolic profiles with an LC/MS-MS-based metabolomics approach. *BMJ Open Respir Res* **1**, e000056.
- [948] Eggers, A. E. (2015). A suggestion about the cause of inflammation in acute atherosis complicating poor placentation in preeclampsia. *Med Hypotheses* **85,** 718-9.
- [949] D'Anna, R., Baviera, G., Scilipoti, A., Leonardi, I. & Leo, R. (2000). The clinical utility of serum uric acid measurements in pre-eclampsia and transient hypertension in pregnancy. *Panminerva Med* **42**, 101-3.
- [950] Williams, K. P. & Galerneau, F. (2002). The role of serum uric acid as a prognostic indicator of the severity of maternal and fetal complications in hypertensive pregnancies. *J Obstet Gynaecol Can* **24**, 628-32.
- [951] Roberts, J. M., Bodnar, L. M., Lain, K. Y., Hubel, C. A., Markovic, N., Ness, R. B. & Powers, R. W. (2005). Uric acid is as important as proteinuria in identifying fetal risk in women with gestational hypertension. *Hypertension* **46**, 1263-9.
- [952] Bainbridge, S. A. & Roberts, J. M. (2008). Uric acid as a pathogenic factor in preeclampsia. *Placenta* **29,** S67-S72.
- [953] Koopmans, C. M., van Pampus, M. G., Groen, H., Aarnoudse, J. G., van den Berg, P. P. & Mol, B. W. J. (2009). Accuracy of serum uric acid as a predictive test for maternal complications in pre-eclampsia: bivariate meta-analysis and decision analysis. *Eur J Obstet Gynecol Reprod Biol* **146**, 8-14.
- [954] Agarwal, V., Gupta, B. K., Vishnu, A., Mamtatyagi, Shiprasolanki & Kiran, J. (2014). Association of Lipid Profile and Uric Acid with Pre-eclampsia of Third Trimester in Nullipara Women. *J Clin Diagn Res* **8**, CC04-7.
- [955] Pereira, K. N., Knoppka, C. K. & da Silva, J. E. (2014). Association between uric acid and severity of preeclampsia. *Clin Lab* **60**, 309-14.
- [956] Lim, K. H., Friedman, S. A., Ecker, J. L., Kao, L. & Kilpatrick, S. J. (1998). The clinical utility of serum uric acid measurements in hypertensive diseases of pregnancy. *Am J Obstet Gynecol* **178**, 1067-71.

- [957] Cnossen, J. S., de Ruyter-Hanhijarvi, H., van der Post, J. A., Mol, B. W., Khan, K. S. & ter Riet, G. (2006). Accuracy of serum uric acid determination in predicting pre-eclampsia: a systematic review. *Acta Obstet Gynecol Scand* **85**, 519-25.
- [958] Powers, R. W., Bodnar, L. M., Ness, R. B., Cooper, K. M., Gallaher, M. J., Frank, M. P., Daftary, A. R. & Roberts, J. M. (2006). Uric acid concentrations in early pregnancy among preeclamptic women with gestational hyperuricemia at delivery. *Am J Obs Gynecol* **194**, 160-166.
- [959] Thangaratinam, S., Ismail, K. M., Sharp, S., Coomarasamy, A., Khan, K. S. & Tests in Prediction of Preeclampsia Severity review, g. (2006). Accuracy of serum uric acid in predicting complications of preeclampsia: a systematic review. *BJOG* **113**, 369-78.
- [960] Chen, Q., Lau, S., Tong, M., Wei, J., Shen, F., Zhao, J. & Zhao, M. (2016). Serum uric acid may not be involved in the development of preeclampsia. *Journal of Human Hypertension* **30**, 136-140.
- [961] Eltzschig, H. K., Sitkovsky, M. V. & Robson, S. C. (2012). Purinergic signaling during inflammation. *N Engl J Med* **367**, 2322-33.
- [962] Kaddurah-Daouk, R., Zhu, H., Sharma, S., Bogdanov, M., Rozen, S. G., Matson, W., Oki, N. O., Motsinger-Reif, A. A., Churchill, E., Lei, Z., Appleby, D., Kling, M. A., Trojanowski, J. Q., Doraiswamy, P. M., Arnold, S. E. & Pharmacometabolomics Research Network. (2013). Alterations in metabolic pathways and networks in Alzheimer's disease. *Transl Psychiatry* 3, e244.
- [963] McFarland, N. R., Burdett, T., Desjardins, C. A., Frosch, M. P. & Schwarzschild, M. A. (2013). Postmortem Brain Levels of Urate and Precursors in Parkinson's Disease and Related Disorders. *Neurodegener Dis* **12**, 189-198.
- [964] Esteve, C., Jones, E. A., Kell, D. B., Boutin, H. & McDonnell, L. A. (2016). Mass spectral imaging shows major derangements in neurogranin and in purine metabolism in the triple-knockout 3xTg Alzheimer-like mouse model. *Front Neurosci*, submitted.
- [965] Lyngdoh, T., Vuistiner, P., Marques-Vidal, P., Rousson, V., Waeber, G., Vollenweider, P. & Bochud, M. (2012). Serum uric acid and adiposity: deciphering causality using a bidirectional Mendelian randomization approach. *PloS one* **7**, e39321.
- [966] Martinon, F. (2010). Mechanisms of uric acid crystal-mediated autoinflammation. *Immunol Rev* **233**, 218-32.
- [967] Orengo, J. M., Leliwa-Sytek, A., Evans, J. E., Evans, B., van de Hoef, D., Nyako, M., Day, K. & Rodriguez, A. (2009). Uric acid is a mediator of the *Plasmodium falciparum*-induced inflammatory response *PloS one* **4**.
- [968] Kool, M., Soullie, T., van Nimwegen, M., Willart, M. A., Muskens, F., Jung, S., Hoogsteden, H. C., Hammad, H. & Lambrecht, B. N. (2008). Alum adjuvant boosts adaptive immunity by inducing uric acid and activating inflammatory dendritic cells. *J Exp Med* **205**, 869-82.
- [969] Chalcraft, K. R. (2013). Comprehensive metabolomics analysis of peanut allergy and peanut-induced analphylaxis. *PhD thesis*.
- [970] Chalcraft, K. R., Kong, J., Waserman, S., Jordana, M. & McCarry, B. E. (2014). Comprehensive metabolomic analysis of peanut-induced anaphylaxis in a murine model. *Metabolomics* **10**, 452-460.
- [971] Kong, J., Chalcraft, K., Mandur, T. S., Jimenez-Saiz, R., Walker, T. D., Goncharova, S., Gordon, M. E., Naji, L., Flader, K., Larche, M., Chu, D. K., Waserman, S., McCarry, B. & Jordana, M. (2015). Comprehensive metabolomics identifies the alarmin uric acid as a critical signal for the induction of peanut allergy. *Allergy* **70**, 495-505.
- [972] van Deventer, S. J. H., Büller, H. R., ten Cate, J. W., Aarden, L. A., Hack, C. E. & Sturk, A. (1990). Experimental endotoxemia in humans: analysis of cytokine release and coagulation, fibrinolytic, and complement pathways. *Blood* **76**, 2520-6.
- [973] Johnson, K., Aarden, L., Choi, Y., De Groot, E. & Creasey, A. (1996). The proinflammatory cytokine response to coagulation and endotoxin in whole blood. *Blood* **87**, 5051-60.
- [974] American College of Chest Physicians, National Institute of Allergy and Infectious Disease & National Heart Lung and Blood Institute. (1997). From the bench to the bedside: the future of sepsis research. Executive summary of an American College of Chest Physicians, National Institute of Allergy and Infectious Disease, and National Heart, Lung, and Blood Institute Workshop. *Chest* 111, 744-53.

- [975] Pernerstorfer, T., Stohlawetz, P., Hollenstein, U., Dzirlo, L., Eichler, H. G., Kapiotis, S., Jilma, B. & Speiser, W. (1999). Endotoxin-induced activation of the coagulation cascade in humans: effect of acetylsalicylic acid and acetaminophen. *Arterioscler Thromb Vasc Biol* **19**, 2517-23.
- [976] Pernerstorfer, T., Hollenstein, U., Hansen, J. B., Stohlawetz, P., Eichler, H. G., Handler, S., Speiser, W. & Jilma, B. (2000). Lepirudin blunts endotoxin-induced coagulation activation. *Blood* **95**, 1729-34.
- [977] Dellinger, R. P. (2003). Inflammation and coagulation: implications for the septic patient. *Clin Infect Dis* **36**, 1259-65.
- [978] Schouten, M., Wiersinga, W. J., Levi, M. & van der Poll, T. (2008). Inflammation, endothelium, and coagulation in sepsis. *J Leukoc Biol* **83**, 536-45.
- [979] Anas, A. A., Wiersinga, W. J., de Vos, A. F. & van der Poll, T. (2010). Recent insights into the pathogenesis of bacterial sepsis. *Neth J Med* **68**, 147-52.
- [980] Saracco, P., Vitale, P., Scolfaro, C., Pollio, B., Pagliarino, M. & Timeus, F. (2011). The coagulopathy in sepsis: significance and implications for treatment. *Pediatr Rep* **3**, e30.
- [981] Tsao, C. M., Ho, S. T. & Wu, C. C. (2015). Coagulation abnormalities in sepsis. *Acta Anaesthesiol Taiwan* **53**, 16-22.
- [982] Armstrong, M. T., Rickles, F. R. & Armstrong, P. B. (2013). Capture of lipopolysaccharide (endotoxin) by the blood clot: a comparative study. *PLoS One* **8**, e80192.
- [983] Al-ofi, E., Coffelt, S. B. & Anumba, D. O. (2014). Fibrinogen, an endogenous ligand of Toll-like receptor 4, activates monocytes in pre-eclamptic patients. *J Reprod Immunol* **103**, 23-8.
- [984] Chatterjee, T., Maitra, D., Chakravarty, T. & Datta, A. G. (1978). Studies on plasma fibrinogen level in pre-eclampsia and eclampsia. *Experientia* **34**, 562-3.
- [985] Manten, G. T. R., Sikkema, J. M., Franx, A., Hameeteman, T. M., Visser, G. H. A., de Groot, P. G. & Voorbij, H. A. M. (2003). Increased high molecular weight fibrinogen in pre-eclampsia. *Thromb Res* **111**, 143-7.
- [986] Üstün, Y., Engin-Üstün, Y. & Kamacı, M. (2005). Association of fibrinogen and C-reactive protein with severity of preeclampsia. *Eur J Obstet Gynecol Reprod Biol* **121**, 154-8.
- [987] Sersam, L. W. (2009). Inflammatory markers in pre-eclampsia and related conditions. *Int J Gynaecol Obstet* **105**, 69-70.
- [988] Perry, K. G. & Martin, J. N. (1992). Abnormal Hemostasis and Coagulopathy in Preeclampsia and Eclampsia. *Clinical Obstetrics and Gynecology* **35**, 338-350.
- [989] Metz, J., Cincotta, R., Francis, M., Derosa, L. & Balloch, A. (1994). Screening for Consumptive Coagulopathy in Preeclampsia. *International Journal of Gynecology & Obstetrics* **46**, 3-9.
- [990] Gilabert, J., Estellés, A., Grancha, S., España, F. & Aznar, J. (1995). Fibrinolytic system and reproductive process with special reference to fibrinolytic failure in pre-eclampsia. *Hum Reprod* 10 Suppl 2, 121-31.
- [991] He, S., Bremme, K. & Blomback, M. (1997). Acquired deficiency of antithrombin in association with a hypercoagulable state and impaired function of liver and/or kidney in preeclampsia. *Blood Coagul Firbinol* **8**, 232-8.
- [992] Hladunewich, M., Karumanchi, S. A. & Lafayette, R. (2007). Pathophysiology of the clinical manifestations of preeclampsia. *Clin J Am Soc Nephrol* **2**, 543-9.
- [993] Kher, A., Bauersachs, R. & Nielsen, J. D. (2007). The management of thrombosis in pregnancy: role of low-molecular-weight heparin. *Thromb Haemost* **97**, 505-13.
- [994] Battinelli, E. M., Marshall, A. & Connors, J. M. (2013). The role of thrombophilia in pregnancy. *Thrombosis* **2013**, 516420.
- [995] Jahromi, B. N. & Rafiee, S. H. (2009). Coagulation Factors in Severe Preeclampsia. *Iranian Red Crescent Medical Journal* **11**, 321-324.
- [996] Ducloy-Bouthors, A. S. (2010). Clotting disorders and preeclampsia (in French). *Ann Fr Anesth Reanim* **29**, e121-34.
- [997] Martínez-Zamora, M. A., Tassies, D., Carmona, F., Espinosa, G., Cervera, R., Reverter, J. C. & Balasch, J. (2010). Clot lysis time and thrombin activatable fibrinolysis inhibitor in severe preeclampsia with or without associated antiphospholipid antibodies. *J Reprod Immunol* **86**, 133-40.
- [998] Dusse, L. M., Rios, D. R. A., Pinheiro, M. B., Cooper, A. J. & Lwaleed, B. A. (2011). Pre-eclampsia: relationship between coagulation, fibrinolysis and inflammation. *Clin Chim Acta* **412**, 17-21.

- [999] Godoi, L. C., Gomes, K. B., Alpoim, P. N., Carvalho, M. d. G., Lwaleed, B. A. & Sant'Ana Dusse, L. M. (2012). Preeclampsia: the role of tissue factor and tissue factor pathway inhibitor. *J Thromb Thrombolysis* **34**, 1-6.
- [1000] Han, L., Liu, X., Li, H., Zou, J., Yang, Z., Han, J., Huang, W., Yu, L., Zheng, Y. & Li, L. (2014). Blood coagulation parameters and platelet indices: changes in normal and preeclamptic pregnancies and predictive values for preeclampsia. *PLoS One* **9**, e114488.
- [1001] Koh, S. C. L., Anandakumar, C., Montan, S. & Ratnam, S. S. (1993). Plasminogen activators, plasminogen activator inhibitors and markers of intravascular coagulation in pre-eclampsia. *Gynecol Obstet Invest* **35**, 214-21.
- [1002] Kaneko, T. & Wada, H. (2011). Diagnostic criteria and laboratory tests for disseminated intravascular coagulation. *J Clin Exp Hematop* **51,** 67-76.
- [1003] Levi, M. & van der Poll, T. (2013). Disseminated intravascular coagulation: a review for the internist. *Int Emerg Med* **8**, 23-32.
- [1004] Asakura, H. (2014). Classifying types of disseminated intravascular coagulation: clinical and animal models. *J Intensive Care* **2**, 20.
- [1005] Wada, H., Matsumoto, T., Yamashita, Y. & Hatada, T. (2014). Disseminated intravascular coagulation: testing and diagnosis. *Clin Chim Acta* **436**, 130-4.
- [1006] Zeerleder, S., Hack, C. E. & Wuillemin, W. A. (2005). Disseminated intravascular coagulation in sepsis. *Chest* **128**, 2864-75.
- [1007] Acikgoz, S., Akduman, D., Eskici, Z. M., Can, M., Mungan, G., Guven, B., Comert, F. & Sumbuloglu, V. (2012). Thrombocyte and erythrocyte indices in sepsis and disseminated intravascular coagulation. *J Med Biochem* **31**, 60-64.
- [1008] Okamoto, K., Tamura, T. & Sawatsubashi, Y. (2016). Sepsis and disseminated intravascular coagulation. *J Intensive Care* **4,** 23.
- [1009] Semeraro, N., Ammollo, C. T., Semeraro, F. & Colucci, M. (2010). Sepsis-associated disseminated intravascular coagulation and thromboembolic disease. *Mediterr J Hematol Infect Dis* **2**, e2010024.
- [1010] Wu, L. C., Lin, X. & Sun, H. (2012). Tanshinone IIA protects rabbits against LPS-induced disseminated intravascular coagulation (DIC). *Acta Pharmacol Sin* **33**, 1254-9.
- [1011] Xu, M., Dong, M. Q., Cao, F. L., Liu, M. L., Wang, Y. X., Dong, H. Y., Huang, Y. F., Liu, Y., Wang, X. B., Zhang, B., Zhao, P. T., Luo, Y., Niu, W., Cui, Y. & Li, Z. C. (2009). Tanshinone IIA reduces lethality and acute lung injury in LPS-treated mice by inhibition of PLA2 activity. *Eur J Pharmacol* **607**, 194-200.
- [1012] Iba, T., Ito, T., Maruyama, I., Jilma, B., Brenner, T., Muller, M. C., Juffermans, N. P. & Thachil, J. (2016). Potential diagnostic markers for disseminated intravascular coagulation of sepsis. *Blood Rev* **30**, 149-55.
- [1013] D'Elia, A. V., Fabbro, D., Driul, L., Barillari, G., Marchesoni, D. & Damante, G. (2011). Plasminogen activator inhibitor-1 gene polymorphisms in pre-eclampsia. *Semin Thromb Hemost* **37**, 97-105.
- [1014] Buurma, A. J., Turner, R. J., Driessen, J. H., Mooyaart, A. L., Schoones, J. W., Bruijn, J. A., Bloemenkamp, K. W. M., Dekkers, O. M. & Baelde, H. J. (2013). Genetic variants in pre-eclampsia: a meta-analysis. *Hum Reprod Update* **19**, 289-303.
- [1015] Morgan, J. A., Bombell, S. & McGuire, W. (2013). Association of plasminogen activator inhibitor-type 1 (-675 4G/5G) polymorphism with pre-eclampsia: systematic review. *PLoS One* **8**, e56907.
- [1016] Kell, D. B. & Pretorius, E. (2016). Substoichiometric molecular control and amplification of the initiation and nature of amyloid fibril formation: lessons from and for blood clotting. bioRxiv preprint. bioRxiv, 054734.
- [1017] Akassoglou, K., Adams, R. A., Bauer, J., Mercado, P., Tseveleki, V., Lassmann, H., Probert, L. & Strickland, S. (2004). Fibrin depletion decreases inflammation and delays the onset of demyelination in a tumor necrosis factor transgenic mouse model for multiple sclerosis. *Proc Natl Acad Sci U S A* 101, 6698-703.
- [1018] Levi, M., van der Poll, T. & Buller, H. R. (2004). Bidirectional relation between inflammation and coagulation. *Circulation* **109**, 2698-704.
- [1019] Flick, M. J., LaJeunesse, C. M., Talmage, K. E., Witte, D. P., Palumbo, J. S., Pinkerton, M. D., Thornton, S. & Degen, J. L. (2007). Fibrin(ogen) exacerbates inflammatory joint disease through a mechanism linked to the integrin alphaMbeta2 binding motif. *J Clin Invest* **117**, 3224-35.

- [1020] Jennewein, C., Tran, N., Paulus, P., Ellinghaus, P., Eble, J. A. & Zacharowski, K. (2011). Novel aspects of fibrin(ogen) fragments during inflammation. *Mol Med* **17**, 568-73.
- [1021] Jennewein, C., Paulus, P. & Zacharowski, K. (2011). Linking inflammation and coagulation: novel drug targets to treat organ ischemia. *Curr Opin Anaesthesiol* **24,** 375-80.
- [1022] Schuliga, M. (2015). The inflammatory actions of coagulant and fibrinolytic proteases in disease. *Mediators Inflamm* **2015**, 437695.
- [1023] Liu, B., Moloney, A., Meehan, S., Morris, K., Thomas, S. E., Serpell, L. C., Hider, R., Marciniak, S. J., Lomas, D. A. & Crowther, D. C. (2011). Iron promotes the toxicity of amyloid beta peptide by impeding its ordered aggregation. *J Biol Chem* **286**, 4248-56.
- [1024] Meyer-Luehmann, M., Spires-Jones, T. L., Prada, C., Garcia-Alloza, M., de Calignon, A., Rozkalne, A., Koenigsknecht-Talboo, J., Holtzman, D. M., Bacskai, B. J. & Hyman, B. T. (2008). Rapid appearance and local toxicity of amyloid-beta plaques in a mouse model of Alzheimer's disease. *Nature* **451**, 720-4.
- [1025] Minter, M. R., Taylor, J. M. & Crack, P. J. (2016). The contribution of neuroinflammation to amyloid toxicity in Alzheimer's disease. *J Neurochem* **136**, 457-74.
- [1026] Miranda, S., Opazo, C., Larrondo, L. F., Munoz, F. J., Ruiz, F., Leighton, F. & Inestrosa, N. C. (2000). The role of oxidative stress in the toxicity induced by amyloid beta-peptide in Alzheimer's disease. *Progr Neurobiol* **62**, 633-648.
- [1027] Rival, T., Page, R. M., Chandraratna, D. S., Sendall, T. J., Ryder, E., Liu, B., Lewis, H., Rosahl, T., Hider, R., Camargo, L. M., Shearman, M. S., Crowther, D. C. & Lomas, D. A. (2009). Fenton chemistry and oxidative stress mediate the toxicity of the beta-amyloid peptide in a *Drosophila* model of Alzheimer's disease. *Eur J Neurosci* **29**, 1335-1347.
- [1028] Özcan, F., Turak, O., Durak, A., Işleyen, A., Uçar, F., Giniş, Z., Uçar, F., Basar, F. N. & Aydoğdu, S. (2013). Red cell distribution width and inflammation in patients with non-dipper hypertension. *Blood Press* **22**, 80-5.
- [1029] Heilmann, L., Mattheck, C. & Kurz, E. (1977). Changes in Blood Rheology and Their Influence on Oxygen Diffusion in Normal and Pathological Pregnancies. *Arch Gynäkol* **223**, 283-298.
- [1030] Stoeff, S., Dikov, I., Vretenarska, M., Jovtchev, S., Trifonova, N. & Penev, M. (2003). Quantitative utility assessment of some biophysical and rheological tests used in hypertension research. 1. relevance analyses of the test parameters. *C. R. Acad Sci Bulg* **56**, 95-100.
- [1031] Kurt, R. K., Aras, Z., Silfeler, D. B., Kunt, C., Islimye, M. & Kosar, O. (2015). Relationship of red cell distribution width with the presence and severity of preeclampsia. *Clin Appl Thromb Hemost* **21**, 128-31.
- [1032] Abdullahi, H., Osman, A., Rayis, D. A., Gasim, G. I., Imam, A. M. & Adam, I. (2014). Red blood cell distribution width is not correlated with preeclampsia among pregnant Sudanese women. *Diagn Pathol* **9**, 29.
- [1033] McMahon, C. J., Hopkins, S., Vail, A., King, A. T., Smith, D., Illingworth, K. J., Clark, S., Rothwell, N. J. & Tyrrell, P. J. (2013). Inflammation as a predictor for delayed cerebral ischemia after aneurysmal subarachnoid haemorrhage. *J Neurointerv Surg* 5, 512-517.
- [1034] Buys, A. V., Van Rooy, M. J., Soma, P., Van Papendorp, D., Lipinski, B. & Pretorius, E. (2013). Changes in red blood cell membrane structure in type 2 diabetes: a scanning electron and atomic force microscopy study. *Cardiovasc Diabetol* **12**, 25.
- [1035] Pretorius, E. & Lipinski, B. (2013). Thromboembolic ischemic stroke changes red blood cell morphology. *Cardiovasc Pathol* **22**, 241-242.
- [1036] Pretorius, E. & Lipinski, B. (2013). Iron alters red blood cell morphology. Blood 121, 9.
- [1037] Swanepoel, A. C. & Pretorius, E. (2012). Scanning electron microscopy analysis of erythrocytes in thromboembolic ischemic stroke. *Int J Lab Hematol* **34**, 185-91.
- [1038] Lang, F., Lang, K. S., Lang, P. A., Huber, S. M. & Wieder, T. (2006). Mechanisms and significance of eryptosis. *Antioxid Redox Signal* **8**, 1183-92.
- [1039] Lang, F., Gulbins, E., Lerche, H., Huber, S. M., Kempe, D. S. & Föller, M. (2008). Eryptosis, a window to systemic disease. *Cell Physiol Biochem* **22**, 373-80.
- [1040] Lang, F. & Qadri, S. M. (2012). Mechanisms and significance of eryptosis, the suicidal death of erythrocytes. *Blood Purif* **33**, 125-30.

- [1041] Lang, F., Lang, E. & Foller, M. (2012). Physiology and pathophysiology of eryptosis. *Transfus Med Hemother* **39**, 308-14.
- [1042] Lang, E. & Lang, F. (2015). Triggers, inhibitors, mechanisms, and significance of eryptosis: the suicidal erythrocyte death. *Biomed Res Int* **2015**, 513518.
- [1043] Lang, E., Zelenak, C., Eberhard, M., Bissinger, R., Rotte, A., Ghashghaeinia, M., Lupescu, A., Lang, F. & Qadri, S. M. (2015). Impact of cyclin-dependent kinase CDK4 inhibition on eryptosis. *Cell Physiol Biochem* 37, 1178-86.
- [1044] Qadri, S. M., Donkor, D. A., Bhakta, V., Eltringham-Smith, L. J., Dwivedi, D. J., Moore, J. C., Pepler, L., Ivetic, N., Nazi, I., Fox-Robichaud, A. E., Liaw, P. C. & Sheffield, W. P. (2016). Phosphatidylserine externalization and procoagulant activation of erythrocytes induced by *Pseudomonas aeruginosa* virulence factor pyocyanin. *J Cell Mol Med* **20**, 710-20.
- [1045] Lang, E., Bissinger, R., Gulbins, E. & Lang, F. (2015). Ceramide in the regulation of eryptosis, the suicidal erythrocyte death. *Apoptosis* **20**, 758-67.
- [1046] Hansson, S. R., Nääv, Å. & Erlandsson, L. (2014). Oxidative stress in preeclampsia and the role of free fetal hemoglobin. *Front Physiol* **5**, 516.
- [1047] Rossi, A. C. & Mullin, P. M. (2011). Prevention of pre-eclampsia with low-dose aspirin or vitamins C and E in women at high or low risk: a systematic review with meta-analysis. *Eur J Obstet Gynecol Reprod Biol* **158**, 9-16.
- [1048] Askie, L. M., Duley, L., Henderson-Smart, D. J., Stewart, L. A. & PARIS Collaborative Group. (2007). Antiplatelet agents for prevention of pre-eclampsia: a meta-analysis of individual patient data. *Lancet* **369**, 1791-8.
- [1049] Roberts, J. M. & Catov, J. M. (2007). Aspirin for pre-eclampsia: compelling data on benefit and risk. *Lancet* **369**, 1765-6.
- [1050] Villa, P. M., Kajantie, E., Räikkönen, K., Pesonen, A. K., Hämäläinen, E., Vainio, M., Taipale, P., Laivuori, H. & PREDO Study group. (2013). Aspirin in the prevention of pre-eclampsia in high-risk women: a randomised placebo-controlled PREDO Trial and a meta-analysis of randomised trials. *BJOG* **120**, 64-74.
- [1051] Bujold, E., Morency, A. M., Roberge, S., Lacasse, Y., Forest, J. C. & Giguère, Y. (2009). Acetylsalicylic acid for the prevention of preeclampsia and intra-uterine growth restriction in women with abnormal uterine artery Doppler: a systematic review and meta-analysis. *J Obstet Gynaecol Can* **31**, 818-26.
- [1052] Roberge, S., Giguere, Y., Villa, P., Nicolaides, K., Vainio, M., Forest, J. C., von Dadelszen, P., Vaiman, D., Tapp, S. & Bujold, E. (2012). Early administration of low-dose aspirin for the prevention of severe and mild preeclampsia: a systematic review and meta-analysis. *Am J Perinatol* 29, 551-6.
- [1053] Roberge, S., Demers, S. & Bujold, E. (2013). Initiation of aspirin in early gestation for the prevention of pre-eclampsia. *BJOG* **120**, 773-4.
- [1054] Bergeron, T. S., Roberge, S., Carpentier, C., Sibai, B., McCaw-Binns, A. & Bujold, E. (2016). Prevention of Preeclampsia with Aspirin in Multiple Gestations: A Systematic Review and Meta-analysis. *Am J Perinatol* **33**, 605-10.
- [1055] Roberge, S., Sibai, B., McCaw-Binns, A. & Bujold, E. (2016). Low-Dose Aspirin in Early Gestation for Prevention of Preeclampsia and Small-for-Gestational-Age Neonates: Meta-analysis of Large Randomized Trials. *Am J Perinatol*.
- [1056] Hofmeyr, G. J., Belizán, J. M., von Dadelszen, P. & Calcium Pre-eclampsia Study Group. (2014). Low-dose calcium supplementation for preventing pre-eclampsia: a systematic review and commentary. *BJOG* **121**, 951-7.
- [1057] Leslie, K., Thilaganathan, B. & Papageorghiou, A. (2011). Early prediction and prevention of pre-eclampsia. *Best Pract Res Clin Obstet Gynaecol* **25**, 343-54.
- [1058] Thangaratinam, S., Langenveld, J., Mol, B. W. & Khan, K. S. (2011). Prediction and primary prevention of pre-eclampsia. *Best Pract Res Clin Obstet Gynaecol* **25**, 419-33.
- [1059] Gillon, T. E., Pels, A., von Dadelszen, P., MacDonell, K. & Magee, L. A. (2014). Hypertensive disorders of pregnancy: a systematic review of international clinical practice guidelines. *PLoS One* **9**, e113715.
- [1060] Cottrell, E. C. & Sibley, C. P. (2015). From Pre-Clinical Studies to Clinical Trials: Generation of Novel Therapies for Pregnancy Complications. *Int J Mol Sci* **16**, 12907-24.

- [1061] Altman, D., Carroli, G., Duley, L., Farrell, B., Moodley, J., Neilson, J., Smith, D. & Magpie Trial Collaboration, G. (2002). Do women with pre-eclampsia, and their babies, benefit from magnesium sulphate? The Magpie Trial: a randomised placebo-controlled trial. *Lancet* **359**, 1877-90.
- [1062] Duley, L. (2005). Evidence and practice: the magnesium sulphate story. *Best Pract Res Clin Obstet Gynaecol* **19**, 57-74.
- [1063] Pratt, J. J., Niedle, P. S., Vogel, J. P., Oladapo, O. T., Bohren, M., Tunçalp, Ö. & Gülmezoglu, A. M. (2016). Alternative regimens of magnesium sulfate for treatment of preeclampsia and eclampsia: a systematic review of non-randomized studies. *Acta Obstet Gynecol Scand* **95**, 144-56.
- [1064] Koren, G., Pastuszak, A. & Ito, S. (1998). Drugs in pregnancy. N Engl J Med 338, 1128-37.
- [1065] Everett, T. R., Wilkinson, I. B. & Lees, C. C. (2012). Drug development in preeclampsia: a 'no go' area? *J Matern Fetal Neonatal Med* **25**, 50-2.
- [1066] Vazquez, J. C. & Abalos, E. (2011). Treatments for symptomatic urinary tract infections during pregnancy. *Cochrane Database Syst Rev*, CD002256.
- [1067] Mathew, D., Khan, K., Thornton, J. G. & Todros, T. (2007). Antibiotics for preventing hypertensive diseases in pregnancy (Protocol). *Cochrane Database Syst Rev 2007*, CD006841.
- [1068] Guerrier, G. & D'Ortenzio, E. (2013). The Jarisch-Herxheimer reaction in leptospirosis: a systematic review. *PLoS One* **8**, e59266.
- [1069] See, S., Scott, E. K. & Levin, M. W. (2005). Penicillin-induced Jarisch-Herxheimer reaction. *Ann Pharmacother* **39**, 2128-30.
- [1070] Pound, M. W. & May, D. B. (2005). Proposed mechanisms and preventative options of Jarisch-Herxheimer reactions. *J Clin Pharm Ther* **30**, 291-5.
- [1071] Prins, J. M., van Deventer, S. J. H., Kuijper, E. J. & Speelman, P. (1994). Clinical relevance of antibiotic-induced endotoxin release. *Antimicrob Agents Chemother* **38**, 1211-8.
- [1072] Kirikae, T., Nakano, M. & Morrison, D. C. (1997). Antibiotic-induced endotoxin release from bacteria and its clinical significance. *Microbiol Immunol* **41**, 285-94.
- [1073] Holzheimer, R. G. (2001). Antibiotic induced endotoxin release and clinical sepsis: a review. *J Chemother* **13 Spec No 1,** 159-72.
- [1074] Lepper, P. M., Held, T. K., Schneider, E. M., Bölke, E., Gerlach, H. & Trautmann, M. (2002). Clinical implications of antibiotic-induced endotoxin release in septic shock. *Intensive Care Med* 28, 824-33.
- [1075] Andersson, J. A., Fitts, E. C., Kirtley, M. L., Ponnusamy, D., Peniche, A. G., Dann, S. M., Motin, V. L., Chauhan, S., Rosenzweig, J. A., Sha, J. & Chopra, A. K. (2016). New role for FDA-approved drugs in combating antibiotic-resistant bacteria. *Antimicrob Agents Chemother*.
- [1076] Poston, L., Briley, A. L., Seed, P. T., Kelly, F. J. & Shennan, A. H. (2006). Vitamin C and vitamin E in pregnant women at risk for pre-eclampsia (VIP trial): randomised placebo-controlled trial. *Lancet* **367**, 1145-54.
- [1077] Ly, C., Yockell-Lelievre, J., Ferraro, Z. M., Arnason, J. T., Ferrier, J. & Gruslin, A. (2015). The effects of dietary polyphenols on reproductive health and early development. *Hum Reprod Update* **21**, 228-48.
- [1078] Liang, S. T., Wong, V. C., So, W. W., Ma, H. K., Chan, V. & Todd, D. (1985). Homozygous alphathalassaemia: clinical presentation, diagnosis and management. A review of 46 cases. *Br J Obstet Gynaecol* **92**, 680-4.
- [1079] Tungwiwat, W., Fucharoen, S., Fucharoen, G., Ratanasiri, T. & Sanchaisuriya, K. (2006). Development and application of a real-time quantitative PCR for prenatal detection of fetal alpha(0)-thalassemia from maternal plasma. *Ann N Y Acad Sci* **1075**, 103-7.
- [1080] Senden, I. P., de Groot, C. J., Steegers, E. A., Bertina, R. M. & Swinkels, D. W. (2004). Preeclampsia and the C282Y mutation in the hemochromatosis (HFE) gene. *Clin Chem* **50**, 973-4.
- [1081] Perron, N. R. & Brumaghim, J. L. (2009). A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. *Cell Biochem Biophys* **53**, 75-100.
- [1082] Dodd, J. M., McLeod, A., Windrim, R. C. & Kingdom, J. (2010). Antithrombotic therapy for improving maternal or infant health outcomes in women considered at risk of placental dysfunction. *Cochrane Database Syst Rev*, CD006780.

- [1083] Li, Y., Wu, Y., Gong, X., Shi, X., Qiao, F. & Liu, H. (2012). Low molecular weight heparin decreases the permeability of glomerular endothelial cells when exposed to pre-eclampsia serum in vitro. *Nephrology (Carlton)* **17**, 754-9.
- [1084] Dodd, J. M., McLeod, A., Windrim, R. C. & Kingdom, J. (2013). Antithrombotic therapy for improving maternal or infant health outcomes in women considered at risk of placental dysfunction. *Cochrane Database Syst Rev* **7**, CD006780.
- [1085] Darmochwał-Kolarz, D., Kolarz, B., Korzeniewski, M., Kimber-Trojnar, Z., Patro-Malysza, J., Mierzynski, R., Przegalinska-Kalamucka, M. & Oleszczuk, J. (2016). A Prevention of Pre-eclampsia with the Use of Acetylsalicylic Acid and Low-molecular Weight Heparin Molecular Mechanisms. *Curr Pharm Biotechnol* 17, 624-8.
- [1086] Zhang, Y., Liu, F., Chen, S. & Zhong, M. (2015). Low-molecular-weight heparin protects kidney through an anti-apoptotic mechanism in a rat pre-eclamptic model. *Eur J Obstet Gynecol Reprod Biol* **188**, 51-5.
- [1087] Roberge, S., Demers, S., Nicolaides, K. H., Bureau, M., Côté, S. & Bujold, E. (2016). Prevention of preeclampsia by low-molecular-weight heparin in addition to aspirin: a meta-analysis. *Ultrasound Obstet Gynecol* **47**, 548-53.
- [1088] Eggers, A. E. (2015). A hypothesis about how to achieve anticoagulation without bleeding. *Med Hypotheses* **85**, 720-2.
- [1089] Kell, D. B. (2016). How drugs pass through biological cell membranes a paradigm shift in our understanding? *Beilstein Magazine* **2**.
- [1090] Park, K., Lee, S., Ahn, H. S. & Kim, D. (2009). Predicting the multi-modal binding propensity of small molecules: towards an understanding of drug promiscuity. *Mol Biosyst* **5**, 844-53.
- [1091] Pérez-Nueno, V. I., Venkatraman, V., Mavridis, L. & Ritchie, D. W. (2012). Detecting drug promiscuity using Gaussian ensemble screening. *J Chem Inf Model* **52**, 1948-61.
- [1092] Tarcsay, A. & Keserü, G. M. (2013). Contributions of molecular properties to drug promiscuity. *J Med Chem* **56**, 1789-95.
- [1093] Arrowsmith, C. H., Audia, J. E., Austin, C., Baell, J., Bennett, J., Blagg, J., Bountra, C., Brennan, P. E., Brown, P. J., Bunnage, M. E., Buser-Doepner, C., Campbell, R. M., Carter, A. J., Cohen, P., Copeland, R. A., Cravatt, B., Dahlin, J. L., Dhanak, D., Edwards, A. M., Frye, S. V., Gray, N., Grimshaw, C. E., Hepworth, D., Howe, T., Huber, K. V. M., Jin, J., Knapp, S., Kotz, J. D., Kruger, R. G., Lowe, D., Mader, M. M., Marsden, B., Mueller-Fahrnow, A., Muller, S., O'Hagan, R. C., Overington, J. P., Owen, D. R., Rosenberg, S. H., Roth, B., Ross, R., Schapira, M., Schreiber, S. L., Shoichet, B., Sundström, M., Superti-Furga, G., Taunton, J., Toledo-Sherman, L., Walpole, C., Walters, M. A., Willson, T. M., Workman, P., Young, R. N. & Zuercher, W. J. (2015). The promise and peril of chemical probes. Nat Chem Biol 11, 536-41.
- [1094] Niphakis, M. J., Lum, K. M., Cognetta, A. B., Correia, B. E., Ichu, T. A., Olucha, J., Brown, S. J., Kundu, S., Piscitelli, F., Rosen, H. & Cravatt, B. F. (2015). A Global Map of Lipid-Binding Proteins and Their Ligandability in Cells. *Cell* **161**, 1668-1680.
- [1095] Mestres, J., Gregori-Puigjané, E., Valverde, S. & Solé, R. V. (2009). The topology of drug-target interaction networks: implicit dependence on drug properties and target families. *Mol Biosyst* 5, 1051-7.
- [1096] Hopkins, A. L. (2008). Network pharmacology: the next paradigm in drug discovery. *Nat Chem Biol* **4**, 682-690
- [1097] Berger, S. I. & Iyengar, R. (2009). Network analyses in systems pharmacology. *Bioinformatics* **25**, 2466-72.
- [1098] van der Graaf, P. H. & Benson, N. (2011). Systems pharmacology: bridging systems biology and pharmacokinetics-pharmacodynamics (PKPD) in drug discovery and development. *Pharm Res* **28**, 1460-4.
- [1099] Cucurull-Sanchez, L., Spink, K. G. & Moschos, S. A. (2012). Relevance of systems pharmacology in drug discovery. *Drug Discov Today* **17**, 665-670.
- [1100] Prentice, R. L., Langer, R. D., Stefanick, M. L., Howard, B. V., Pettinger, M., Anderson, G. L., Barad, D., Curb, J. D., Kotchen, J., Kuller, L., Limacher, M. & Wactawski-Wende, J. (2006). Combined analysis of

- Women's Health Initiative observational and clinical trial data on postmenopausal hormone treatment and cardiovascular disease. *Am J Epidemiol* **163**, 589-99.
- [1101] Couzin, J. (2008). Cholesterol veers off script. Science 322, 220-3.
- [1102] Peterson, R. T. (2008). Chemical biology and the limits of reductionism. Nat Chem Biol 4, 635-8.
- [1103] Robinson, J. G. (2008). Models for describing relations among the various statin drugs, low-density lipoprotein cholesterol lowering, pleiotropic effects, and cardiovascular risk. *Am J Cardiol* **101**, 1009-15.
- [1104] Endo, A. (2010). A historical perspective on the discovery of statins. *Proc Jpn Acad Ser B Phys Biol Sci* **86,** 484-93.
- [1105] Wagner, B. K., Kitami, T., Gilbert, T. J., Peck, D., Ramanathan, A., Schreiber, S. L., Golub, T. R. & Mootha, V. K. (2008). Large-scale chemical dissection of mitochondrial function. *Nat Biotechnol* **26**, 343-351.
- [1106] Libby, P. & Aikawa, M. (2002). Stabilization of atherosclerotic plaques: new mechanisms and clinical targets. *Nat Med* **8**, 1257-62.
- [1107] Liao, J. K. (2002). Beyond lipid lowering: the role of statins in vascular protection. *Int J Cardiol* **86,** 5-18.
- [1108] Undas, A., Brozek, J. & Musial, J. (2002). Anti-inflammatory and antithrombotic effects of statins in the management of coronary artery disease. *Clin Lab* **48**, 287-96.
- [1109] Weitz-Schmidt, G. (2002). Statins as anti-inflammatory agents. Trends Pharmacol Sci 23, 482-6.
- [1110] Blanco-Colio, L. M., Tuñon, J., Martin-Ventura, J. L. & Egido, J. (2003). Anti-inflammatory and immunomodulatory effects of statins. *Kidney Int* **63**, 12-23.
- [1111] Kwak, B. R., Mulhaupt, F. & Mach, F. (2003). Atherosclerosis: anti-inflammatory and immunomodulatory activities of statins. *Autoimmun Rev* **2**, 332-8.
- [1112] Steffens, S. & Mach, F. (2004). Anti-inflammatory properties of statins. Semin Vasc Med 4, 417-22.
- [1113] Jain, M. K. & Ridker, P. M. (2005). Anti-inflammatory effects of statins: clinical evidence and basic mechanisms. *Nat Rev Drug Discov* **4,** 977-87.
- [1114] Abeles, A. M. & Pillinger, M. H. (2006). Statins as antiinflammatory and immunomodulatory agents: a future in rheumatologic therapy? *Arthritis Rheum* **54,** 393-407.
- [1115] Endres, M. (2006). Statins: potential new indications in inflammatory conditions. *Atheroscler Suppl* **7,** 31-5.
- [1116] Li, J. J., Zheng, X. & Li, J. (2007). Statins may be beneficial for patients with slow coronary flow syndrome due to its anti-inflammatory property. *Med Hypotheses* **69**, 333-7.
- [1117] Mira, E. & Manes, S. (2009). Immunomodulatory and anti-inflammatory activities of statins. *Endocr Metab Immune Disord Drug Targets* **9,** 237-47.
- [1118] Dinarello, C. A. (2010). Anti-inflammatory Agents: Present and Future. Cell 140, 935-50.
- [1119] Bu, D. X., Griffin, G. & Lichtman, A. H. (2011). Mechanisms for the anti-inflammatory effects of statins. *Curr Opin Lipidol* **22**, 165-70.
- [1120] Antonopoulos, A. S., Margaritis, M., Lee, R., Channon, K. & Antoniades, C. (2012). Statins as antiinflammatory agents in atherogenesis: molecular mechanisms and lessons from the recent clinical trials. *Curr Pharm Des* **18**, 1519-30.
- [1121] Liappis, A. P., Kan, V. L., Rochester, C. G. & Simon, G. L. (2001). The effect of statins on mortality in patients with bacteremia. *Clin Infect Dis* **33**, 1352-7.
- [1122] Terblanche, M., Almog, Y., Rosenson, R. S., Smith, T. S. & Hackam, D. G. (2006). Statins: panacea for sepsis? *Lancet Infect Dis* **6**, 242-8.
- [1123] Falagas, M. E., Makris, G. C., Matthaiou, D. K. & Rafailidis, P. I. (2008). Statins for infection and sepsis: a systematic review of the clinical evidence. *J Antimicrob Chemother* **61,** 774-85.
- [1124] Sun, H. Y. & Singh, N. (2009). Antimicrobial and immunomodulatory attributes of statins: relevance in solid-organ transplant recipients. *Clin Infect Dis* **48**, 745-55.
- [1125] Kozarov, E., Padro, T. & Badimon, L. (2014). View of statins as antimicrobials in cardiovascular risk modification. *Cardiovasc Res* **102**, 362-74.
- [1126] Sandek, A., Utchill, S. & Rauchhaus, M. (2007). The endotoxin-lipoprotein hypothesis an update. *Archives of Medical Science* **3,** S81-S90.

- [1127] Ukinc, K., Ersoz, H. O., Erem, C., Hacihasanoglu, A. B. & Karti, S. S. (2009). Effects of one year simvastatin and atorvastatin treatments on acute phase reactants in uncontrolled type 2 diabetic patients. *Endocrine* **35**, 380-8.
- [1128] Mascitelli, L. & Goldstein, M. R. (2012). Might the beneficial effects of statin drugs be related to their action on iron metabolism? *QJM* **105**, 1225-9.
- [1129] Kanugula, A. K., Gollavilli, P. N., Vasamsetti, S. B., Karnewar, S., Gopoju, R., Ummanni, R. & Kotamraju, S. (2014). Statin-induced inhibition of breast cancer proliferation and invasion involves attenuation of iron transport: intermediacy of nitric oxide and antioxidant defence mechanisms. *FEBS J* **281**, 3719-38.
- [1130] Lecarpentier, E., Morel, O., Fournier, T., Elefant, E., Chavatte-Palmer, P. & Tsatsaris, V. (2012). Statins and pregnancy: between supposed risks and theoretical benefits. *Drugs* **72**, 773-88.
- [1131] Costantine, M. M., Cleary, K., Eunice Kennedy Shriver National Institute of Child, H. & Human Development Obstetric--Fetal Pharmacology Research Units, N. (2013). Pravastatin for the prevention of preeclampsia in high-risk pregnant women. *Obstet Gynecol* **121**, 349-53.
- [1132] Downing, J. W., Baysinger, C. L., Johnson, R. F. & Paschall, R. L. (2013). Review: Potential druggable targets for the treatment of early onset preeclampsia. *Pregnancy Hypertens* **3**, 203-10.
- [1133] Carver, A. R., Tamayo, E., Perez-Polo, J. R., Saade, G. R., Hankins, G. D. V. & Costantine, M. M. (2014). The effect of maternal pravastatin therapy on adverse sensorimotor outcomes of the offspring in a murine model of preeclampsia. *Int J Dev Neurosci* **33**, 33-40.
- [1134] Cindrova-Davies, T. (2014). The therapeutic potential of antioxidants, ER chaperones, NO and H2S donors, and statins for treatment of preeclampsia. *Front Pharmacol* **5**, 119.
- [1135] Girardi, G. (2014). Can statins prevent pregnancy complications? J Reprod Immunol 101-102, 161-7.
- [1136] McDonnold, M., Tamayo, E., Kechichian, T., Gamble, P., Longo, M., Hankins, G. D. V., Saade, G. R. & Costantine, M. M. (2014). The effect of prenatal pravastatin treatment on altered fetal programming of postnatal growth and metabolic function in a preeclampsia-like murine model. *Am J Obstet Gynecol* **210**, 542 e1-7.
- [1137] Ramma, W. & Ahmed, A. (2014). Therapeutic potential of statins and the induction of heme oxygenase-1 in preeclampsia. *J Reprod Immunol* **101-102**, 153-60.
- [1138] Saad, A. F., Kechichian, T., Yin, H., Sbrana, E., Longo, M., Wen, M., Tamayo, E., Hankins, G. D., Saade, G. R. & Costantine, M. M. (2014). Effects of pravastatin on angiogenic and placental hypoxic imbalance in a mouse model of preeclampsia. *Reprod Sci* **21**, 138-45.
- [1139] Staff, A. C., Johnsen, G. M., Dechend, R. & Redman, C. W. G. (2014). Preeclampsia and uteroplacental acute atherosis: immune and inflammatory factors. *J Reprod Immunol* 101-102, 120-6.
- [1140] Ahmed, A. & Ramma, W. (2015). Unravelling the theories of pre-eclampsia: are the protective pathways the new paradigm? *Br J Pharmacol* **172**, 1574-86.
- [1141] Bauer, A. J., Banek, C. T., Needham, K., Gillham, H., Capoccia, S., Regal, J. F. & Gilbert, J. S. (2013). Pravastatin attenuates hypertension, oxidative stress, and angiogenic imbalance in rat model of placental ischemia-induced hypertension. *Hypertension* **61**, 1103-10.
- [1142] Carver, A. R., Andrikopoulou, M., Lei, J., Tamayo, E., Gamble, P., Hou, Z., Zhang, J., Mori, S., Saade, G. R., Costantine, M. M. & Burd, I. (2014). Maternal pravastatin prevents altered fetal brain development in a preeclamptic CD-1 mouse model. *PLoS One* **9**, e100873.
- [1143] Lefkou, E., Mamopoulos, A., Fragakis, N., Dagklis, T., Vosnakis, C., Nounopoulos, E., Rousso, D. & Girardi, G. (2014). Clinical improvement and successful pregnancy in a preeclamptic patient with antiphospholipid syndrome treated with pravastatin. *Hypertension* **63**, e118-9.
- [1144] Brownfoot, F. C., Tong, S., Hannan, N. J., Binder, N. K., Walker, S. P., Cannon, P., Hastie, R., Onda, K. & Kaitu'u-Lino, T. J. (2015). Effects of Pravastatin on Human Placenta, Endothelium, and Women With Severe Preeclampsia. *Hypertension* **66**, 687-97; discussion 445.
- [1145] Chaiworapongsa, T., Romero, R., Korzeniewski, S. J., Chaemsaithong, P., Hernandez-Andrade, E., Segars, J. H., DeCherney, A. H., McCoy, M. C., Kim, C. J., Yeo, L. & Hassan, S. S. (2016). Pravastatin to prevent recurrent fetal death in massive perivillous fibrin deposition of the placenta (MPFD). *J Matern Fetal Neonatal Med* **29**, 855-62.

- [1146] Downing, J. (2010). Sildenafil for the treatment of preeclampsia. *Hypertens Pregnancy* **29,** 248-50; author reply 251-2.
- [1147] Kakigano, A., Tomimatsu, T., Mimura, K., Kanayama, T., Fujita, S., Minato, K., Kumasawa, K., Taniguchi, Y., Kanagawa, T., Endo, M., Ishihara, T., Namba, T., Mizushima, T. & Kimura, T. (2015). Drug Repositioning for Preeclampsia Therapeutics by In Vitro Screening: Phosphodiesterase-5 Inhibitor Vardenafil Restores Endothelial Dysfunction via Induction of Placental Growth Factor. *Reprod Sci* 22, 1272-80.
- [1148] Ramesar, S. V., Mackraj, I., Gathiram, P. & Moodley, J. (2011). Sildenafil citrate decreases sFlt-1 and sEng in pregnant I-NAME treated Sprague-Dawley rats. *Eur J Obstet Gynecol Reprod Biol* **157**, 136-40.
- [1149] Herraiz, S., Pellicer, B., Serra, V., Cauli, O., Cortijo, J., Felipo, V. & Pellicer, A. (2012). Sildenafil citrate improves perinatal outcome in fetuses from pre-eclamptic rats. *BJOG* **119**, 1394-402.
- [1150] Nassar, A. H., Masrouha, K. Z., Itani, H., Nader, K. A. & Usta, I. M. (2012). Effects of sildenafil in Nomega-nitro-L-arginine methyl ester-induced intrauterine growth restriction in a rat model. *Am J Perinatol* **29**, 429-34.
- [1151] Stanley, J. L., Andersson, I. J., Poudel, R., Rueda-Clausen, C. F., Sibley, C. P., Davidge, S. T. & Baker, P. N. (2012). Sildenafil citrate rescues fetal growth in the catechol-O-methyl transferase knockout mouse model. *Hypertension* **59**, 1021-8.
- [1152] George, E. M., Palei, A. C., Dent, E. A. & Granger, J. P. (2013). Sildenafil attenuates placental ischemia-induced hypertension. *Am J Physiol Regul Integr Comp Physiol* **305**, R397-403.
- [1153] Stanley, J. L., Sulek, K., Andersson, I. J., Davidge, S. T., Kenny, L. C., Sibley, C. P., Mandal, R., Wishart, D. S., Broadhurst, D. I. & Baker, P. N. (2015). Sildenafil Therapy Normalizes the Aberrant Metabolomic Profile in the Comt(-/-) Mouse Model of Preeclampsia/Fetal Growth Restriction. *Sci Rep* **5**, 18241.
- [1154] Karasu, E., Kayacan, N., Sadan, G. & Dinc, B. (2011). Different effects of different phosphodiesterase type-5 inhibitors in pre-eclampsia. *Pregnancy Hypertens* **1,** 231-7.
- [1155] Karasu, E., Kayacan, N., Sadan, G. & Dinc, B. (2012). Endothelial dysfunction in the human umbilical artery due to preeclampsia can be prevented by sildenafil. *Clin Exp Hypertens* **34,** 79-85.
- [1156] Ganzevoort, W., Alfirevic, Z., von Dadelszen, P., Kenny, L., Papageorghiou, A., van Wassenaer-Leemhuis, A., Gluud, C., Mol, B. W. & Baker, P. N. (2014). STRIDER: Sildenafil Therapy In Dismal prognosis Early-onset intrauterine growth Restriction--a protocol for a systematic review with individual participant data and aggregate data meta-analysis and trial sequential analysis. *Syst Rev* 3, 23.
- [1157] Abalos, E., Duley, L. & Steyn, D. W. (2014). Antihypertensive drug therapy for mild to moderate hypertension during pregnancy. *Cochrane Database Syst Rev* **2**, CD002252.
- [1158] Mahmud, H., Föller, M. & Lang, F. (2008). Stimulation of erythrocyte cell membrane scrambling by methyldopa. *Kidney Blood Press Res* **31**, 299-306.
- [1159] Firoz, T., Magee, L. A., MacDonell, K., Payne, B. A., Gordon, R., Vidler, M., von Dadelszen, P. & Community Level Interventions for Pre-eclampsia Working Group. (2014). Oral antihypertensive therapy for severe hypertension in pregnancy and postpartum: a systematic review. *BJOG* 121, 1210-8; discussion 1220.
- [1160] Magee, L. A., Namouz-Haddad, S., Cao, V., Koren, G. & von Dadelszen, P. (2015). Labetalol for hypertension in pregnancy. *Expert Opin Drug Saf* **14**, 453-61.
- [1161] Ishimaru, T., Ishida, J., Nakamura, S., Hashimoto, M., Matsukura, T., Nakamura, A., Kunita, S., Sugiyama, F., Yagami, K. & Fukamizu, A. (2012). Short-term suppression of the renin-angiotensin system in mice associated with hypertension during pregnancy. *Mol Med Rep* **6**, 28-32.
- [1162] Marshall, T. G., Lee, R. E. & Marshall, F. E. (2006). Common angiotensin receptor blockers may directly modulate the immune system via VDR, PPAR and CCR2b. *Theor Biol Med Model* **3**, 1.
- [1163] Ziegler, E. J., Fisher, C. J., Jr., Sprung, C. L., Straube, R. C., Sadoff, J. C., Foulke, G. E., Wortel, C. H., Fink, M. P., Dellinger, R. P., Teng, N. N. & et al. (1991). Treatment of gram-negative bacteremia and septic shock with HA-1A human monoclonal antibody against endotoxin. A randomized, double-blind, placebo-controlled trial. The HA-1A Sepsis Study Group. *N Engl J Med* **324**, 429-36.

- [1164] Derkx, B., Wittes, J. & McCloskey, R. (1999). Randomized, placebo-controlled trial of HA-1A, a human monoclonal antibody to endotoxin, in children with meningococcal septic shock. European Pediatric Meningococcal Septic Shock Trial Study Group. *Clin Infect Dis* **28**, 770-7.
- [1165] Baumgartner, J. D. & Glauser, M. P. (1993). Immunotherapy of endotoxemia and septicemia. *Immunobiology* **187**, 464-77.
- [1166] Yentis, S. M., Soni, N. & Riches, P. G. (1994). *In vitro* effects of HA-1A (Centoxin) on cytokine production in whole blood from intensive care unit patients. *Br J Anaesth* **73**, 805-11.
- [1167] Marks, L. (2012). The birth pangs of monoclonal antibody therapeutics: the failure and legacy of Centoxin. *MAbs* **4**, 403-12.
- [1168] Helmerhorst, E. J., Maaskant, J. J. & Appelmelk, B. J. (1998). Anti-lipid A monoclonal antibody centoxin (HA-1A) binds to a wide variety of hydrophobic ligands. *Infect Immun* **66**, 870-3.
- [1169] Tan, N. S., Ng, M. L., Yau, Y. H., Chong, P. K. W., Ho, B. & Ding, J. L. (2000). Definition of endotoxin binding sites in horseshoe crab factor C recombinant sushi proteins and neutralization of endotoxin by sushi peptides. *FASEB J* 14, 1801-13.
- [1170] Tan, N. S., Ho, B. & Ding, J. L. (2000). High-affinity LPS binding domain(s) in recombinant factor C of a horseshoe crab neutralizes LPS-induced lethality. *FASEB J* **14**, 859-70.
- [1171] Ding, J. L., Li, P. & Ho, B. (2008). The Sushi peptides: structural characterization and mode of action against Gram-negative bacteria. *Cell Mol Life Sci* **65**, 1202-19.
- [1172] Yau, Y. H., Ho, B., Tan, N. S., Ng, M. L. & Ding, J. L. (2001). High therapeutic index of factor C Sushi peptides: potent antimicrobials against *Pseudomonas aeruginosa*. *Antimicrob Agents Chemother* **45**, 2820-5.
- [1173] Leptihn, S., Har, J. Y., Chen, J., Ho, B., Wohland, T. & Ding, J. L. (2009). Single molecule resolution of the antimicrobial action of quantum dot-labeled sushi peptide on live bacteria. *BMC Biol* **7**, 22.
- [1174] Leptihn, S., Guo, L., Frecer, V., Ho, B., Ding, J. L. & Wohland, T. (2010). One step at a time: action mechanism of Sushi 1 antimicrobial peptide and derived molecules. *Virulence* **1**, 42-4.
- [1175] Li, P., Sun, M., Wohland, T., Ho, B. & Ding, J. L. (2006). The molecular mechanism of interaction between sushi peptide and *Pseudomonas* endotoxin. *Cell Mol Immunol* **3**, 21-8.
- [1176] Li, P., Sun, M., Wohland, T., Yang, D., Ho, B. & Ding, J. L. (2006). Molecular mechanisms that govern the specificity of Sushi peptides for Gram-negative bacterial membrane lipids. *Biochemistry* **45**, 10554-62.
- [1177] Frecer, V., Ho, B. & Ding, J. L. (2004). *De novo* design of potent antimicrobial peptides. *Antimicrob Agents Chemother* **48**, 3349-57.
- [1178] Bhattacharjya, S. (2010). *De novo* designed lipopolysaccharide binding peptides: structure based development of antiendotoxic and antimicrobial drugs. *Curr Med Chem* **17**, 3080-93.
- [1179] Brantsæter, A. L., Myhre, R., Haugen, M., Myking, S., Sengpiel, V., Magnus, P., Jacobsson, B. & Meltzer, H. M. (2011). Intake of probiotic food and risk of preeclampsia in primiparous women: the Norwegian Mother and Child Cohort Study. *Am J Epidemiol* **174**, 807-15.
- [1180] Gluckman, P., Beedle, A. & Hanson, M. (2009). *Principles of evolutionary medicine*. Oxford University Press, Oxford.
- [1181] Hemmings, D. G. (2006). Signal transduction underlying the vascular effects of sphingosine 1-phosphate and sphingosylphosphorylcholine. *Naunyn Schmiedebergs Arch Pharmacol* **373**, 18-29.
- [1182] Hemmings, D. G., Hudson, N. K., Halliday, D., O'Hara, M., Baker, P. N., Davidge, S. T. & Taggart, M. J. (2006). Sphingosine-1-phosphate acts via rho-associated kinase and nitric oxide to regulate human placental vascular tone. *Biol Reprod* **74**, 88-94.
- [1183] Teijaro, J. R., Walsh, K. B., Cahalan, S., Fremgen, D. M., Roberts, E., Scott, F., Martinborough, E., Peach, R., Oldstone, M. B. & Rosen, H. (2011). Endothelial cells are central orchestrators of cytokine amplification during influenza virus infection. *Cell* **146**, 980-91.
- [1184] Oldstone, M. B. A., Teijaro, J. R., Walsh, K. B. & Rosen, H. (2013). Dissecting influenza virus pathogenesis uncovers a novel chemical approach to combat the infection. *Virology* **435**, 92-101.
- [1185] Maceyka, M. & Spiegel, S. (2014). Sphingolipid metabolites in inflammatory disease. *Nature* **510**, 58-67.

- [1186] Oldstone, M. B. A. & Rosen, H. (2014). Cytokine storm plays a direct role in the morbidity and mortality from influenza virus infection and is chemically treatable with a single sphingosine-1-phosphate agonist molecule. *Curr Top Microbiol Immunol* **378**, 129-47.
- [1187] Walsh, K. B., Teijaro, J. R., Brock, L. G., Fremgen, D. M., Collins, P. L., Rosen, H. & Oldstone, M. B. A. (2014). Animal model of respiratory syncytial virus: CD8+ T cells cause a cytokine storm that is chemically tractable by sphingosine-1-phosphate 1 receptor agonist therapy. *J Virol* 88, 6281-93.
- [1188] Arish, M., Husein, A., Kashif, M., Saleem, M., Akhter, Y. & Rub, A. (2016). Sphingosine-1-phosphate signaling: unraveling its role as a drug target against infectious diseases. *Drug Discov Today* **21**, 133-42.
- [1189] Zhang, J., Dunk, C. E. & Lye, S. J. (2013). Sphingosine signalling regulates decidual NK cell angiogenic phenotype and trophoblast migration. *Hum Reprod* **28**, 3026-37.
- [1190] Nagamatsu, T., Iwasawa-Kawai, Y., Ichikawa, M., Kawana, K., Yamashita, T., Osuga, Y., Fujii, T. & Schust, D. J. (2014). Emerging roles for lysophospholipid mediators in pregnancy. *Am J Reprod Immunol* **72**, 182-91.
- [1191] Romanowicz, L. & Bańkowski, E. (2010). Altered sphingolipid composition in Wharton's jelly of pre-eclamptic newborns. *Pathobiology* **77**, 78-87.
- [1192] Romanowicz, L. & Bańkowski, E. (2010). Sphingolipids of human umbilical cord vein and their alteration in preeclampsia. *Mol Cell Biochem* **340**, 81-9.
- [1193] Romanowicz, L. & Bańkowski, E. (2010). Lipid compounds of human Wharton's jelly and their alterations in preeclampsia. *Int J Exp Pathol* **91,** 1-9.
- [1194] Li, Q., Pan, Z., Wang, X., Gao, Z., Ren, C. & Yang, W. (2014). miR-125b-1-3p inhibits trophoblast cell invasion by targeting sphingosine-1-phosphate receptor 1 in preeclampsia. *Biochem Biophys Res Commun* **453**, 57-63.
- [1195] Baig, S., Lim, J. Y., Fernandis, A. Z., Wenk, M. R., Kale, A., Su, L. L., Biswas, A., Vasoo, S., Shui, G. & Choolani, M. (2013). Lipidomic analysis of human placental syncytiotrophoblast microvesicles in adverse pregnancy outcomes. *Placenta* **34**, 436-42.
- [1196] Carrillo-Vico, A., Lardone, P. J., Naji, L., Fernández-Santos, J. M., Martín-Lacave, I., Guerrero, J. M. & Calvo, J. R. (2005). Beneficial pleiotropic actions of melatonin in an experimental model of septic shock in mice: regulation of pro-/anti-inflammatory cytokine network, protection against oxidative damage and anti-apoptotic effects. *J Pineal Res* **39**, 400-408.
- [1197] Gitto, E., Pellegrino, S., Gitto, P., Barberi, I. & Reiter, R. J. (2009). Oxidative stress of the newborn in the pre- and postnatal period and the clinical utility of melatonin. *J Pineal Res* **46**, 128-39.
- [1198] Hobson, S. R., Lim, R., Gardiner, E. E., Alers, N. O. & Wallace, E. M. (2013). Phase I pilot clinical trial of antenatal maternally administered melatonin to decrease the level of oxidative stress in human pregnancies affected by pre-eclampsia (PAMPR): study protocol. *BMJ Open* **3**, e003788.
- [1199] Westh, H., Lisby, G., Breysse, F., Boddinghaus, B., Chomarat, M., Gant, V., Goglio, A., Raglio, A., Schuster, H., Stuber, F., Wissing, H. & Hoeft, A. (2009). Multiplex real-time PCR and blood culture for identification of bloodstream pathogens in patients with suspected sepsis. *Clin Microbiol Infect* **15**, 544-51.
- [1200] Yang, B., Wang, Y. & Qian, P. Y. (2016). Sensitivity and correlation of hypervariable regions in 16S rRNA genes in phylogenetic analysis. *BMC Bioinformatics* **17**, 135.
- [1201] Khamis, A., Raoult, D. & La Scola, B. (2005). Comparison between *rpoB* and 16S rRNA gene sequencing for molecular identification of 168 clinical isolates of *Corynebacterium*. *J Clin Microbiol* **43**, 1934-6.
- [1202] Adékambi, T., Drancourt, M. & Raoult, D. (2009). The *rpoB* gene as a tool for clinical microbiologists. *Trends Microbiol* **17,** 37-45.
- [1203] Roux, S., Enault, F., Bronner, G. & Debroas, D. (2011). Comparison of 16S rRNA and protein-coding genes as molecular markers for assessing microbial diversity (Bacteria and Archaea) in ecosystems. *FEMS Microbiol Ecol* **78**, 617-28.
- [1204] Liu, W., Li, L., Khan, M. A. & Zhu, F. (2012). Popular molecular markers in bacteria. *Mol Gen Mikrobiol Virusol*, 14-7.
- [1205] Větrovský, T. & Baldrian, P. (2013). The variability of the 16S rRNA gene in bacterial genomes and its consequences for bacterial community analyses. *PLoS One* **8**, e57923.

- [1206] Das, S., Dash, H. R., Mangwani, N., Chakraborty, J. & Kumari, S. (2014). Understanding molecular identification and polyphasic taxonomic approaches for genetic relatedness and phylogenetic relationships of microorganisms. *J Microbiol Meth* **103**, 80-100.
- [1207] Cody, A. J., McCarthy, N. D., Jansen van Rensburg, M., Isinkaye, T., Bentley, S. D., Parkhill, J., Dingle, K. E., Bowler, I. C., Jolley, K. A. & Maiden, M. C. (2013). Real-time genomic epidemiological evaluation of human *Campylobacter* isolates by use of whole-genome multilocus sequence typing. *J Clin Microbiol* 51, 2526-34.
- [1208] Lazar, L., Rigó, J., Jr., Nagy, B., Balogh, K., Makó, V., Cervenak, L., Mézes, M., Prohászka, Z. & Molvarec, A. (2009). Relationship of circulating cell-free DNA levels to cell-free fetal DNA levels, clinical characteristics and laboratory parameters in preeclampsia. *BMC Med Genet* **10**, 120.
- [1209] Galbiati, S., Causarano, V., Pinzani, P., Francesca, S., Orlando, C., Smid, M., Pasi, F., Castiglioni, M. T., Cavoretto, P., Rovere-Querini, P., Pedroni, S., Calza, S., Ferrari, M. & Cremonesi, L. (2010). Evaluation of a panel of circulating DNA, RNA and protein potential markers for pathologies of pregnancy. Clin Chem Lab Med 48, 791-4.
- [1210] Miranda, M. L., Macher, H. C., Munoz-Hernandez, R., Vallejo-Vaz, A., Moreno-Luna, R., Villar, J., Guerrero, J. M. & Stiefel, P. (2013). Role of circulating cell-free DNA levels in patients with severe preeclampsia and HELLP syndrome. *Am J Hypertens* **26**, 1377-80.
- [1211] Oudejans, C. B. M. (2015). Maternal plasma RNA sequencing. Clin Biochem 48, 942-7.
- [1212] Hudecova, I. (2015). Digital PCR analysis of circulating nucleic acids. Clin Biochem 48, 948-56.
- [1213] Coates, A. R. M. & Hu, Y. (2006). New strategies for antibacterial drug design: targeting non-multiplying latent bacteria. *Drugs R D* 7, 133-51.
- [1214] Coates, A. R., Halls, G. & Hu, Y. (2011). Novel classes of antibiotics or more of the same? *Br J Pharmacol* **163**, 184-94.
- [1215] Hu, Y., Shamaei-Tousi, A., Liu, Y. & Coates, A. (2010). A new approach for the discovery of antibiotics by targeting non-multiplying bacteria: a novel topical antibiotic for staphylococcal infections. *PLoS One* **5**, e11818.
- [1216] Hu, Y., Liu, A., Ortega-Muro, F., Alameda-Martin, L., Mitchison, D. & Coates, A. (2015). High-dose rifampicin kills persisters, shortens treatment duration, and reduces relapse rate *in vitro* and *in vivo*. Front Microbiol **6**, 641.
- [1217] Laxminarayan, R., Duse, A., Wattal, C., Zaidi, A. K., Wertheim, H. F., Sumpradit, N., Vlieghe, E., Hara, G. L., Gould, I. M., Goossens, H., Greko, C., So, A. D., Bigdeli, M., Tomson, G., Woodhouse, W., Ombaka, E., Peralta, A. Q., Qamar, F. N., Mir, F., Kariuki, S., Bhutta, Z. A., Coates, A., Bergstrom, R., Wright, G. D., Brown, E. D. & Cars, O. (2013). Antibiotic resistance the need for global solutions. *Lancet Infect Dis* 13, 1057-98.
- [1218] Krishnan, R., Tsubery, H., Proschitsky, M. Y., Asp, E., Lulu, M., Gilead, S., Gartner, M., Waltho, J. P., Davis, P. J., Hounslow, A. M., Kirschner, D. A., Inouye, H., Myszka, D. G., Wright, J., Solomon, B. & Fisher, R. A. (2014). A bacteriophage capsid protein provides a general amyloid interaction motif (GAIM) that binds and remodels misfolded protein assemblies. *J Mol Biol* **426**, 2500-19.
- [1219] Barber, M., Tait, R. C., Scott, J., Rumley, A., Lowe, G. D. & Stott, D. J. (2004). Dementia in subjects with atrial fibrillation: hemostatic function and the role of anticoagulation. *J Thromb Haemost* **2**, 1873-8.
- [1220] Murthy, S. B., Jawaid, A., Qureshi, S. U., Schulz, P. E. & Schulz, P. E. (2009). The apolipoprotein 2 allele in Alzheimer's disease: suggestions for a judicious use of antiplatelet and anticoagulant medications. *J Am Geriatr Soc* **57**, 1124-5.
- [1221] Ahn, H. J., Glickman, J. F., Poon, K. L., Zamolodchikov, D., Jno-Charles, O. C., Norris, E. H. & Strickland, S. (2014). A novel Abeta-fibrinogen interaction inhibitor rescues altered thrombosis and cognitive decline in Alzheimer's disease mice. *J Exp Med* **211**, 1049-62.
- [1222] Ankarcrona, M., Winblad, B., Monteiro, C., Fearns, C., Powers, E. T., Johansson, J., Westermark, G. T., Presto, J., Ericzon, B. G. & Kelly, J. W. (2016). Current and future treatment of amyloid diseases. *J Intern Med*.
- [1223] Marshall, B. J. (1994). Helicobacter pylori. Am J Gastroenterol 89, S116-28.